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NB100-106 Protocol

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Western blot Protocol for Ogg1 Antibody (NB100-106)

Western Blot Procedure

- 1. Run 50 ug of protein on a 4-20% Tris-glycine mini-gel at 125V for 90 minutes.
- 2. Equilibrate gel, nitrocellulose membrane, Whatman paper, and blotting pads in transfer buffer for 15 minutes.
- 3. Transfer protein to the membrane at 25V for 90 minutes.
- 4. Allow membrane to air-dry.
- 5. Block membrane with 1XPBS/3% BSA for 1 hour at room temperature (23-27 degrees C).
- 6. Wash membrane twice, for 5 minutes each, with 1XPBS/0.05% Tween-20 (PBST).
- 7. Incubate membrane with NB100-106 (anti-hOGG1), diluted in 1XPBS/1% BSA, for 1 hour at room temperature.
- 8. Wash membrane once for 15 minutes, then four times for 5 minutes each, with PBST.
- 9. Incubate membrane with goat anti-rabbit IgG-HRP, diluted in 1XPBS/1% BSA, for 1 hour at room temperature.
- 10. Wash membrane once for 15 minutes, then four times for 5 minutes each, with PBST.

11. Detect cross-reacting proteins using Renaissance Chemiluminescence Reagent Plus kit from NEN Life Sciences.