

# PRODUCT INFORMATION & ELISA MANUAL

# Cytosolic Sulfotransferase 2B1/SULT2B1 Antibody Pair [HRP]

NBP2-79636

## Sample Insert for reference use only

Matched Antibody Pair utilized in an Enzyme-linked Immunosorbent Assay for quantitative detection of Human Cytosolic Sulfotransferase 2B1/SULT2B1.

For research use only.

Not for diagnostic or therapeutic procedures.

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Novus kits are guaranteed for 6 months from date of receipt

#### **BACKGROUND**

Sulfotransferase family cytosolic 2B member 1, also known as Sulfotransferase 2B1, ST2B1, Alcohol sulfotransferase, Hydroxysteroid sulfotransferase 2, SULT2B1 and HSST2, is a cytoplasm protein which belongs to the sulfotransferase 1 family. The human hydroxysteroid sulfotransferase (SULT) family is comprised of two subfamilies, SULT2A1 and SULT2B1. SULT2B1 is expressed highly in placenta, prostate and trachea. A lesser expression of SULT1B1 was observed in the small intestine and lung. SULT2B1 catalyzes the sulfate conjugation of many hormones, neurotransmitters, drugs and xenobiotic compounds. Sulfonation increases the water solubility of most compounds, and therefore their renal excretion, but it can also result in bioactivation to form active metabolites. SULT2B1 sulfates hydroxysteroids like DHEA. Isoform 1 preferentially sulfonates cholesterol. The two SULT2B1 isoforms, SULT2B1a and SULT2B1b, are encoded by a single gene as a result of alternative transcription initiation and alternative splicing. SULT2B1b catalyzes the sulfonation of 3beta-hydroxysteroid hormones and cholesterol, whereas SULT2B1a preferentially catalyzes pregnenolone sulfonation.

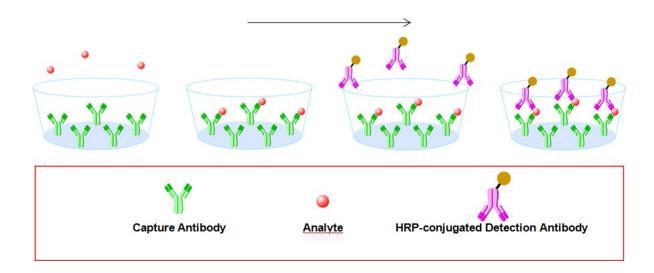
#### PRINCIPLE OF THE TEST

The Novus Biologicals Cytosolic Sulfotransferase 2B1/SULT2B1 Antibody Pair [HRP] is a solid phase sandwich ELISA (Enzyme-Linked Immunosorbent Assay). It utilizes a monoclonal antibody specific for Cytosolic Sulfotransferase 2B1/SULT2B1 coated on a 96-well plate. Standards and samples are added to the wells, and any Cytosolic Sulfotransferase 2B1/SULT2B1 present binds to the immobilized antibody. The wells are washed and a horseradish peroxidase conjugated rabbit anti-Cytosolic Sulfotransferase 2B1/SULT2B1 polyclonal antibody is then added, producing an antibody-antigen-antibody "sandwich". The wells are again washed and TMB substrate solution is loaded, which produces color in proportion to the amount of Cytosolic Sulfotransferase 2B1/SULT2B1 present in the sample. To end the enzyme reaction, the stop solution is added and absorbances of the microwell are read at 450 nm.

#### INTENDED USE

- ◆ The Human Cytosolic Sulfotransferase 2B1/SULT2B1 Antibody Pair [HRP] is for the quantitative determination of Human Cytosolic Sulfotransferase 2B1/SULT2B1.
- ◆ This Cytosolic Sulfotransferase 2B1/SULT2B1 Antibody Pair [HRP] contains the basic components required for the development of sandwich ELISAs.

#### **ASSAY PROCEDURE SUMMARY**



This antibody pair has been configured for research use only and is not to be used in diagnostic procedures.

#### MATERIALS PROVIDED

Bring all reagents to room temperature before use.

**Capture Antibody** - 0.5 mg/mL of mouse anti-Cytosolic Sulfotransferase 2B1/SULT2B1 monoclonal antibody (in PBS, pH 7.4). Dilute to a working concentration of 2  $\mu$ g/mL in CBS before coating.

**Detection Antibody** - 0.5 mg/mL rabbit anti-Cytosolic Sulfotransferase 2B1/SULT2B1 polyclonal antibody conjugated to horseradish-peroxidase (HRP) (in PBS, 50 % glycerol, pH 7.4). Dilute to working concentration of 0.5  $\mu$ g/mL in detection antibody dilution buffer before use.

**Standard** – Each vial contains 160 ng of recombinant Cytosolic Sulfotransferase 2B1/SULT2B1. Reconstitute with 1 mL detection antibody dilution buffer. After reconstitution, store at -20°C to -80°C in a manual defrost freezer. A seven-point standard curve using 2-fold serial dilutions in sample dilution buffer, and a high standard of 3 ng/mL is recommended.

#### **SOLUTIONS REQUIRED**

**CBS** - 0.05M Na<sub>2</sub>CO<sub>3</sub>, 0.05M NaHCO<sub>3</sub>, pH 9.6, 0.2 µm filtered

TBS - 20 mM Tris, 150 mM NaCl, pH 7.4

Wash Buffer - 0.05% Tween20 in TBS, pH 7.2 - 7.4

Blocking Buffer - 2% BSA in Wash Buffer

Sample dilution buffer - 0.1% BSA in wash buffer, pH 7.2 - 7.4, 0.2 µm filtered

Detection antibody dilution buffer - 0.5% BSA in wash buffer, pH 7.2 - 7.4, 0.2 µm filtered

Substrate Solution: To achieve best assay results, fresh substrate solution is recommended

Substrate stock solution - 10mg / ml TMB (Tetramethylbenzidine) in DMSO

Substrate dilution buffer - 0.05M Na₂HPO₄ and 0.025M citric acid; adjust pH to 5.5

**Substrate working solution** - For each plate dilute 250  $\mu$ l substrate stock solution in 25ml substrate dilution buffer and then add 80  $\mu$ l 0.75%  $H_2O_2$ , mix it well

Stop Solution - 2 N H<sub>2</sub>SO<sub>4</sub>

#### **PRECAUTION**

The Stop Solution suggested for use with this antibody pair is an acid solution. Wear eye, hand, face, and clothing protection when using this material.

#### **STORAGE**

**Capture Antibody**: Aliquot and store at  $-20^{\circ}$ C to  $-80^{\circ}$ C for up to 6 months from date of receipt. Avoid repeated freeze-thaw cycles.

**Detection Antibody**: Protect it from prolonged exposure to light. Aliquot and store at  $-20^{\circ}$ C to  $-80^{\circ}$ C and for up to 6 months from date of receipt. Avoid repeated freeze-thaw cycles.

**Standard**: Store lyophilized standard at -20  $^{\circ}$ C to -80  $^{\circ}$ C for up to 6 months from date of receipt. Aliquot and store the reconstituted standard at -80  $^{\circ}$ C for up to 1 month. Avoid repeated freeze-thaw cycles.

#### **GENERAL ELISA PROTOCOL**

#### **Plate Preparation**

- 1. Dilute the capture antibody to the working concentration in CBS. Immediately coat a 96-well microplate with 100µL per well of the diluted capture antibody. Seal the plate and incubate overnight at 4°C.
- 2. Aspirate each well and wash with at least 300µl wash buffer, repeating the process two times for a total of three washes. Complete removal of liquid at each step is essential for good performance. After the last wash, remove any remaining wash buffer by inverting the plate and blotting it against clean paper towels.
- 3.Block plates by adding 300 µL of blocking buffer to each well. Incubate at room temperature for a minimum of 1 hour.
- 4.Repeat the aspiration/wash as in step 2. The plates are now ready for sample addition.

#### **Assay Procedure**

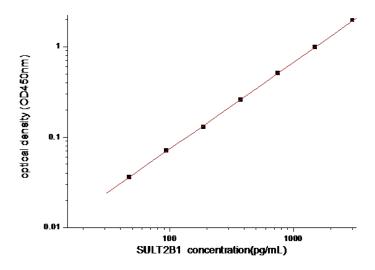
- $1.Add\ 100\ \mu L$  of sample or standards in sample dilution buffer per well. Seal the plate and incubate 2 hours at room temperature.
- 2. Repeat the aspiration/wash as in step 2 of plate preparation.
- 3. Add  $100~\mu L$  of the detection antibody, diluted in antibody dilution buffer, to each well. Seal the plate and incubate 1 hour at room temperature.
- 4. Repeat the aspiration/wash as in step 2 of plate preparation.
- 5. Add  $200~\mu L$  of substrate solution to each well. Incubate for 20~minutes at room temperature ( if substrate solution is not as requested, the incubation time should be optimized ). Avoid placing the plate in direct light.
- 6.Add 50 µL of stop solution to each well. Gently tap the plate to ensure thorough mixing.
- 7. Determine the optical density of each well immediately, using a microplate reader set to 450 nm.

#### CALCULATION OF RESULTS

- Calculate the mean absorbance for each set of duplicate standards, controls and samples. Subtract the mean zero standard absorbance from each.
- Construct a standard curve by plotting the mean absorbance for each standard on the y-axis against the concentration on the x-axis and draw a best fit curve through the points on the graph.
- •To determine the concentration of the unknowns, find the unknowns' mean absorbance value on the y-axis and draw a horizontal line to the standard curve. At the point of intersection, draw a vertical line to the x-axis and read the concentration. If samples have been diluted, the concentration read from the standard curve must be multiplied by the dilution factor.
- Alternatively, computer-based curve-fitting statistical software may also be employed to calculate the concentration of the sample.

#### **TYPICAL DATA**

This standard curve is only for demonstration purposes. A standard curve should be generated for each assay.



| Concentration (pg/ml) | Zero standard subtracted OD |
|-----------------------|-----------------------------|
| 0                     | 0.000                       |
| 47                    | 0.036                       |
| 94                    | 0.071                       |
| 188                   | 0.129                       |
| 375                   | 0.259                       |
| 750                   | 0.509                       |
| 1500                  | 0.986                       |
| 3000                  | 1.966                       |

#### PERFORMANCE CHARACTERISTIC

#### **SENSITIVITY**

The minimum detectable dose of Human Cytosolic Sulfotransferase 2B1/SULT2B1 was determined to be approximately 47 pg/ml. This is defined as at least three times standard deviations above the mean optical density of 10 replicates of the zero standard.

### **TROUBLE SHOOTING**

| Problems               | Possible Sources  | Solutions  |  |  |  |
|------------------------|---|--|--|--|--|
|                        | Incorrect or no Detection Antibody was added                          | Add appropriate Detection Antibody and continue                                      |  |  |  |
| No signal              | Substrate solution was not added                                      | Add substrate solution and continue  |  |  |  |
|                        | Incorrect storage condition   | Check if the kit is stored at recommended condition and used before expiration date  |  |  |  |
| Poor Standard<br>Curve | Standard was incompletely reconstituted or was inappropriately stored | Aliquot reconstituted standard and store at -80 $^{\circ}\mathrm{C}$                 |  |  |  |
|                        | Imprecise / inaccurate pipetting                                      | Check / calibrate pipettes   |  |  |  |
|                        | Incubations done at inappropriate temperature, timing or agitation    | Follow the general ELISA protocol  |  |  |  |
|                        | Background wells were contaminated                                    | Avoid cross contamination by using the sealer appropriately                          |  |  |  |
|                        | The concentration of antigen in samples was too low                   | Enriching samples to increase the concentration of antigen                           |  |  |  |
| Poor detection value   | Samples were ineffective  | Check if the samples are stored at cold environment. Detect samples in timely manner |  |  |  |
|                        | Insufficient washes   | Use multichannel pipettes without touching the reagents on the plate                 |  |  |  |
| High Background        | insunicient wasnes  | Increase cycles of washes and soaking time between washes                            |  |  |  |
|                        | TMB Substrate Solution was contaminated                               | TMB Substrate Solution should be clear and colorless prior to addition to wells      |  |  |  |
|                        | Materials were contaminated.  | Use clean plates, tubes and pipettes tips  |  |  |  |
| Non-specificity        | Samples were contaminated   | Avoid cross contamination of samples   |  |  |  |
|                        | The concentration of samples was too high                             | Try higher dilution rate of samples  |  |  |  |

|   | ELISA Plate Template |   |   |   |   |   |   |   |   |    |    |    |
|---|----------------------|---|---|---|---|---|---|---|---|----|----|----|
|   | 1                    | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 | 12 |
| A |                      |   |   |   |   |   |   |   |   |    |    |    |
| В |                      |   |   |   |   |   |   |   |   |    |    |    |
| С |                      |   |   |   |   |   |   |   |   |    |    |    |
| D |                      |   |   |   |   |   |   |   |   |    |    |    |
| E |                      |   |   |   |   |   |   |   |   |    |    |    |
| F |                      |   |   |   |   |   |   |   |   |    |    |    |
| G |                      |   |   |   |   |   |   |   |   |    |    |    |
|   |                      |   |   |   |   |   |   |   |   |    |    |    |

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Notes