

## PRODUCT INFORMATION & ELISA MANUAL

# IL-18/IL-1F4 Antibody Pair [HRP]

NBP2-79493

## Sample Insert for reference use only

Matched Antibody Pair utilized in an Enzyme-linked Immunosorbent Assay for quantitative detection of Mouse IL-18/IL-1F4.

For research use only.

Not for diagnostic or therapeutic procedures.

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Novus kits are guaranteed for 6 months from date of receipt

#### **BACKGROUND**

Interleukin-18 (IL-18, also known as interferon-gamma inducing factor) is a proinflammatory cytokine that belongs to the IL-1 superfamily and is produced by macrophages and other cells. This cytokine can induce the IFN-gamma production of T cells. The combination of IL-18 and IL12 has been shown to inhibit IL4 dependent IgE and IgG1 production, and enhance IgG2a production of B cells. IL-18 binding protein (IL18BP) can specifically interact with this cytokine, and thus negatively regulate its biological activity. IL-18 is an IL-1-like cytokine that requires cleavage with caspase-1 to become active, was found to increase IgE production in a CD4+ T cells-, IL-4- and STAT6-dependent fashion. IL-18 and T cell receptor-mediated stimulation could induce naïve CD4+ T cells to develop into IL-4-producing cells in vitro. Thus, caspase-1 and IL-18 may be critical in regulation of IgE production in vivo, providing a potential therapeutic target for allergic disorders. IL-18 production in primary synovial cultures and purified synovial fibroblasts was, in turn, upregulated by TNF-α and IL-1β, suggesting that monokine expression can feed back to promote Th1 cell development in synovial membrane. Besides, synergistic combinations of IL-18, IL-12, and IL-15 may be of importance in sustaining both Th1 responses and monokine production in RA.

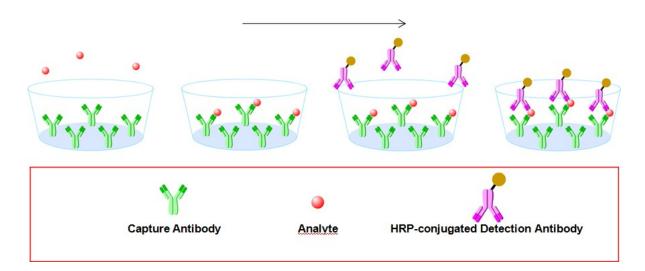
#### PRINCIPLE OF THE TEST

The IL-18/IL-1F4 Antibody Pair [HRP] is a solid phase sandwich ELISA (Enzyme-Linked Immunosorbent Assay). It utilizes a monoclonal antibody specific for Mouse IL-18/IL-1F4 coated on a 96-well plate. Standards and samples are added to the wells, and any Mouse IL-18/IL-1F4 present binds to the immobilized antibody. The wells are washed and a horseradish peroxidase conjugated rabbit anti-Mouse IL-18/IL-1F4 monoclonal antibody is then added, producing an antibody-antigen- antibody "sandwich". The wells are again washed and TMB substrate solution is loaded, which produces color in proportion to the amount of Mouse IL-18/IL-1F4 present in the sample. To end the enzyme reaction, the stop solution is added and absorbances of the microwell are read at 450 nm.

#### INTENDED USE

- ◆ The Mouse IL-18/IL-1F4 ELISA Pair Set is for the quantitative determination of Mouse IL-18/IL-1F4.
- ◆This ELISA Pair Set contains the basic components required for the development of sandwich ELISAs.

#### ASSAY PROCEDURE SUMMARY



This Pair Set has been configured for research use only and is not to be used in diagnostic procedures.

#### MATERIALS PROVIDED

Bring all reagents to room temperature before use.

**Capture Antibody** – 1 mg/mL of rabbit anti-Mouse IL-18/IL-1F4 monoclonal antibody (in PBS, pH 7.4). Dilute to a working concentration of 2 μg/mL in PBS before coating.

**Detection Antibody** - 0.2 mg/mL of rabbit anti-Mouse IL-18/IL-1F4 monoclonal antibody conjugated to horseradish-peroxidase (HRP) (in PBS, 50 % HRP-Protector, pH 7.4, store at 4  $^{\circ}$ C). Dilute to working concentration of 0.25 µg/mL in detection antibody dilution buffer before use.

**Standard** – Each vial contains 70 ng of recombinant Mouse IL-18/IL-1F4. Reconstitute with 1 mL detection antibody dilution buffer. After reconstitution, store at  $-20^{\circ}$ C to  $-80^{\circ}$ C in a manual defrost freezer. A seven-point standard curve using 2-fold serial dilutions in sample dilution buffer, and a high standard of 1500 pg/mL is recommended.

#### **SOLUTIONS REQUIRED**

**PBS** - 136.9 mM NaCl, 10.1 mM Na $_2$ HPO $_4$ , 2.7 mM KCl, 1.8 mM KH $_2$ PO $_4$ , pH 7.4, 0.2  $\mu$ m filtered

**TBS** - 20 mM Tris, 150 mM NaCl, pH 7.4

**Wash Buffer** - 0.05% Tween20 in TBS, pH 7.2 - 7.4

**Blocking Buffer** - 2% BSA in Wash Buffer

Sample dilution buffer - 0.1% BSA in wash buffer, pH 7.2 - 7.4, 0.2 µm filtered

**Detection antibody dilution buffer** - 0.5% BSA in wash buffer, pH 7.2 - 7.4, 0.2  $\mu m$  filtered

Substrate Solution: To achieve best assay results, fresh substrate solution is recommended

Substrate stock solution - 10mg / ml TMB ( Tetramethylbenzidine ) in DMSO

Substrate dilution buffer -  $0.05M\ Na_2HPO_4$  and  $0.025M\ citric$  acid; adjust pH to 5.5

**Substrate working solution** - For each plate dilute 250  $\mu$ l substrate stock solution in 25ml substrate dilution buffer and then add 80  $\mu$ l 0.75%  $H_2O_2$ , mix it well

Stop Solution - 2 N H<sub>2</sub>SO<sub>4</sub>

#### **PRECAUTION**

The Stop Solution suggested for use with this Pair Set is an acid solution. Wear eye, hand, face, and clothing protection when using this material.

#### STORAGE

**Capture Antibody**: Aliquot and store at  $-20^{\circ}$ C to  $-80^{\circ}$ C for up to 6 months from date of receipt. Avoid repeated freeze-thaw cycles.

**Detection Antibody**: Store at  $4^{\circ}$ C and protect it from prolonged exposure to light for up to 6 months from date of receipt. **DO NOT FREEZE!** 

**Standard**: Store lyophilized standard at  $-20\,^{\circ}$ C to  $-80\,^{\circ}$ C for up to 6 months from date of receipt. Aliquot and store the reconstituted standard at  $-80\,^{\circ}$ C for up to 1 month. Avoid repeated freeze-thaw cycles.

#### **GENERAL ELISA PROTOCOL**

#### **Plate Preparation**

- 1. Dilute the capture antibody to the working concentration in PBS. Immediately coat a 96-well microplate with 100 $\mu$ L per well of the diluted capture antibody. Seal the plate and incubate overnight at 4°C.
- 2. Aspirate each well and wash with at least  $300\mu$ I wash buffer, repeating the process two times for a total of three washes. Complete removal of liquid at each step is essential for good performance. After the last wash, remove any remaining wash buffer by inverting the plate and blotting it against clean paper towels. 3.Block plates by adding 300  $\mu$ L of blocking buffer to each well. Incubate at room temperature for a minimum of 1 hour.
- 4.Repeat the aspiration/wash as in step 2. The plates are now ready for sample addition.

#### **Assay Procedure**

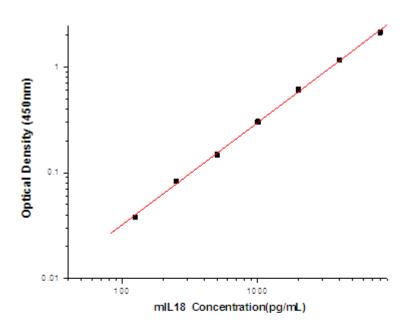
- 1. Add 100  $\mu$ L of sample or standards in sample dilution buffer per well. Seal the plate and incubate 2 hours at room temperature.
- 2. Repeat the aspiration/wash as in step 2 of plate preparation.
- 3.Add  $100~\mu L$  of the detection antibody, diluted in antibody dilution buffer, to each well. Seal the plate and incubate 1 hour at room temperature.
- 4. Repeat the aspiration/wash as in step 2 of plate preparation.
- $5. Add\ 200\ \mu L$  of substrate solution to each well. Incubate for 20 minutes at room temperature ( **if substrate solution is not as requested, the incubation time should be optimized** ). Avoid placing the plate in direct light.
- 6.Add 50 µL of stop solution to each well. Gently tap the plate to ensure thorough mixing.
- 7. Determine the optical density of each well immediately, using a microplate reader set to 450 nm.

#### CALCULATION OF RESULTS

- Calculate the mean absorbance for each set of duplicate standards, controls and samples. Subtract the mean zero standard absorbance from each.
- Construct a standard curve by plotting the mean absorbance for each standard on the y-axis against the concentration on the x-axis and draw a best fit curve through the points on the graph.
- •To determine the concentration of the unknowns, find the unknowns' mean absorbance value on the y-axis and draw a horizontal line to the standard curve. At the point of intersection, draw a vertical line to the x-axis and read the concentration. If samples have been diluted, the concentration read from the standard curve must be multiplied by the dilution factor.
- Alternatively, computer-based curve-fitting statistical software may also be employed to calculate the concentration of the sample.

#### **TYPICAL DATA**

This standard curve is only for demonstration purposes. A standard curve should be generated for each assay.



| Concentration ( pg/mL) | Zero standard subtracted OD |  |  |  |  |
|------------------------|-----------------------------|--|--|--|--|
| 0                      | 0                           |  |  |  |  |
| 23.44                  | 0.026                       |  |  |  |  |
| 46.88                  | 0.053                       |  |  |  |  |
| 93.75                  | 0.129                       |  |  |  |  |
| 187.5                  | 0.265                       |  |  |  |  |
| 375                    | 0.529                       |  |  |  |  |
| 750                    | 1.090                       |  |  |  |  |
| 1500                   | 2.114                       |  |  |  |  |

#### PERFORMANCE CHARACTERISTIC

#### **SENSITIVITY**

The minimum detectable dose of Mouse IL-18/IL-1F4 was determined to be approximately **23.44 pg/ml**. This is defined as at least three times standard deviations above the mean optical density of 10 replicates of the zero standard.

#### **TROUBLE SHOOTING**

| Problems               | Possible Sources  | Solutions  |  |  |  |
|------------------------|---|--|--|--|--|
|                        | Incorrect or no Detection Antibody was added                          | Add appropriate Detection Antibody and continue                                      |  |  |  |
| No signal              | Substrate solution was not added                                      | Add substrate solution and continue  |  |  |  |
|                        | Incorrect storage condition   | Check if the kit is stored at recommended condition and used before expiration date  |  |  |  |
| Poor Standard<br>Curve | Standard was incompletely reconstituted or was inappropriately stored | Aliquot reconstituted standard and store at -80 $^{\circ}\mathrm{C}$                 |  |  |  |
|                        | Imprecise / inaccurate pipetting                                      | Check / calibrate pipettes   |  |  |  |
|                        | Incubations done at inappropriate temperature, timing or agitation    | Follow the general ELISA protocol  |  |  |  |
|                        | Background wells were contaminated                                    | Avoid cross contamination by using the sealer appropriately                          |  |  |  |
|                        | The concentration of antigen in samples was too low                   | Enriching samples to increase the concentration of antigen                           |  |  |  |
| Poor detection value   | Samples were ineffective  | Check if the samples are stored at cold environment. Detect samples in timely manner |  |  |  |
| High Background        | Insufficient washes   | Use multichannel pipettes without touching the reagents on the plate                 |  |  |  |
|                        | insunicient wasnes  | Increase cycles of washes and soaking time between washes                            |  |  |  |
|                        | TMB Substrate Solution was contaminated                               | TMB Substrate Solution should be clear and colorless prior to addition to wells      |  |  |  |
|                        | Materials were contaminated.  | Use clean plates, tubes and pipettes tips  |  |  |  |
| Non-specificity        | Samples were contaminated   | Avoid cross contamination of samples   |  |  |  |
|                        | The concentration of samples was too high                             | Try higher dilution rate of samples  |  |  |  |

|   | ELISA Plate Template |   |   |   |   |   |   |   |   |    |    |    |
|---|----------------------|---|---|---|---|---|---|---|---|----|----|----|
|   | 1                    | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 | 12 |
| A |                      |   |   |   |   |   |   |   |   |    |    |    |
| В |                      |   |   |   |   |   |   |   |   |    |    |    |
| С |                      |   |   |   |   |   |   |   |   |    |    |    |
| D |                      |   |   |   |   |   |   |   |   |    |    |    |
| E |                      |   |   |   |   |   |   |   |   |    |    |    |
| F |                      |   |   |   |   |   |   |   |   |    |    |    |
| G |                      |   |   |   |   |   |   |   |   |    |    |    |
| Н |                      |   |   |   |   |   |   |   |   |    |    |    |

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