

## PRODUCT INFORMATION & ELISA MANUAL

# CD84/SLAMF5 Antibody Pair [HRP]

NBP2-79439

## Sample Insert for reference use only

Matched Antibody Pair utilized in an Enzyme-linked Immunosorbent Assay for quantitative detection of Human CD84/SLAMF5.

For research use only.

Not for diagnostic or therapeutic procedures.

www.novusbio.com - P: 303.730.1950 - P: 888.506.6887 - F: 303.730.1966 - technical@novusbio.com

Novus kits are guaranteed for 6 months from date of receipt

#### **BACKGROUND**

The CD2 family receptors are type I transmembrane glycoproteins belonging to immunoglobulin (Ig) superfamily characterized by a membrane-proximal Ig constant 2 (C2) domain and a membrane-distal variable (V) domain that is responsible for ligand recognition. CD84, also known as LY9B and SLAMF5, is a homophilic member of the SLAM (signaling lymphocyte activation molecule) subfamily of the CD2 family. The SLAM family receptorsmediate signal transduction through the interaction of its ITSM (immunoreceptor tyrosine-based switch motifs) in the intracellular region and the SH2 domain of adaptor molecules SAP (SLAM-associated protein) and EAT-2 (EWS-activated transcript 2), and accordingly modulate both adaptive and innate immune responses. The CD84-CD84 interaction was independent of its cytoplasmic tail. Thus, CD84 is its own ligand and acts as a costimulatory molecule. CD84 is expressed on cells from almost all hematopoietic lineages and on CD34+ hematopoietic progenitor cells, suggesting that CD84 serves as a marker for committed hematopoietic progenitor cells.

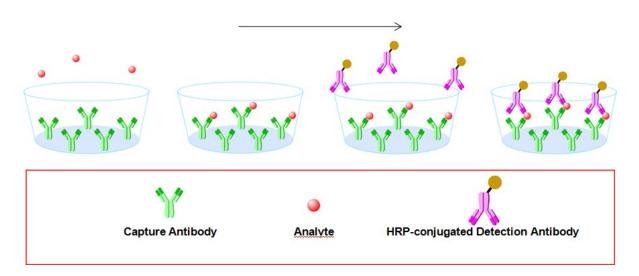
#### PRINCIPLE OF THE TEST

The Novus Biologicals CD84/SLAMF5 Antibody Pair [HRP] is asolid phase sandwich ELISA (Enzyme-Linked ImmunosorbentAssay). It utilizes a monoclonal antibody specific for CD84/SLAMF coatedon a 96-well plate. Standards and samples are added to the wells, and any CD84/SLAMF presentbinds to the immobilized antibody. The wells are washed and a horseradish peroxidase conjugated mouse anti-CD84/SLAMF monoclonal antibody is then added, producing an antibody-antigen-antibody"sandwich". The wells are again washed and TMB substrate solution is loaded, which produces color in proportion to the amount of CD84/SLAMF presentin the sample. To end the enzyme reaction, the stop solution is added and absorbances of the microwell are read at 450 nm.

#### INTENDED USE

- ◆ The human CD84/SLAMF5 Antibody Pair [HRP] is for the quantitative determination of human CD84/SLAMF.
- ◆This CD84/SLAMF5 Antibody Pair [HRP] contains the basic components required for the development of sandwich ELISAs.

#### ASSAY PROCEDURE SUMMARY



This antibody pair has been configured for research use only and is not to be used in diagnostic procedures.

#### MATERIALS PROVIDED

Bring all reagents to room temperature before use.

**Capture Antibody** - 0.2 mg/mL of rabbit anti-CD84/SLAMF monoclonal antibody. Dilute to a working concentration of 2.0  $\mu$ g/mL in CBS before coating.

**Detection Antibody** - 0.5 mg/mL mouse antiCD84/SLAMF conjugated to horseradish-peroxidase (HRP). Dilute to working concentration of 0.5  $\mu$ g/mL in detection antibody dilution buffer before use.

**Standard** – Each vial contains 400 ng of recombinant CD84/SLAMF. Reconstitute standard powder with 1 mL detection antibody dilution buffer. After reconstitution, store at -20°C to -80°C in a manual defrost freezer. A seven-point standard curve usi ng 2-fold serial dilutions in sample dilution buffer, and a high standard of 10000 pg/mL is recommended.

#### SOLUTIONS REQUIRED

**CBS** - 0.05M Na<sub>2</sub>CO<sub>3</sub>, 0.05M NaHCO<sub>3</sub>, pH 9.6, 0.2 µm filtered

**TBS** - 20 mM Tris, 150 mM NaCl, pH 7.4

**Wash Buffer** - 0.05% Tween20 in TBS, pH 7.2 - 7.4

Blocking Buffer - 2% BSA in Wash Buffer

Sample dilution buffer - 0.1% BSA in wash buffer, pH 7.2 - 7.4, 0.2 µm filtered

Detection antibody dilution buffer - 0.5% BSA in wash buffer, pH 7.2 - 7.4, 0.2 µm filtered

Substrate Solution: To achieve best assay results, fresh substrate solution is recommended

Substrate stock solution - 10mg / ml TMB ( Tetramethylbenzidine ) in DMSO

Substrate dilution buffer - 0.05M Na<sub>2</sub>HPO<sub>4</sub> and 0.025M citric acid; adjust pH to 5.5

**Substrate working solution** - For each plate dilute 250  $\mu$ l substrate stock solution in 25ml substrate dilution buffer and then add 80  $\mu$ l 0.75%  $H_2O_2$ , mix it well

Stop Solution - 2 N H<sub>2</sub>SO<sub>4</sub>

#### **PRECAUTION**

The Stop Solution suggested for use with this antibody pair is an acid solution. Wear eye, hand, face, and clothing protection when using this material.

#### **STORAGE**

**Capture Antibody**: Aliquot and store at  $-20^{\circ}$ C to  $-80^{\circ}$ C for up to 6 months from date of receipt. Avoid repeated freeze-thaw cycles.

**Detection Antibody**: Protect it from prolonged exposure to light. Aliquot and store at  $-20^{\circ}$ C to  $-80^{\circ}$ C and for up to 6 months from date of receipt. Avoid repeated freeze-thaw cycles.

**Standard**: Store lyophilized standard at -20  $^{\circ}$ C to -80  $^{\circ}$ C for up to 6 months from date of receipt. Aliquot and store the reconstituted standard at -80  $^{\circ}$ C for up to 1 month. Avoid repeated freeze-thaw cycles.

#### **GENERAL ELISA PROTOCOL**

#### **Plate Preparation**

- 1. Dilute the capture antibody to the working concentration in CBS. Immediately coat a 96-well microplate with 100 $\mu$ L per well of the diluted capture antibody. Seal the plate and incubate overnight at 4°C.
- 2. Aspirate each well and wash with at least  $300\mu$ I wash buffer, repeating the process two times for a total of three washes. Complete removal of liquid at each step is essential for good performance. After the last wash, remove any remaining wash buffer by inverting the plate and blotting it against clean paper towels. 3.Block plates by adding 300  $\mu$ L of blocking buffer to each well. Incubate at room temperature for a minimum of 1 hour.
- 4.Repeat the aspiration/wash as in step 2. The plates are now ready for sample addition.

#### **Assay Procedure**

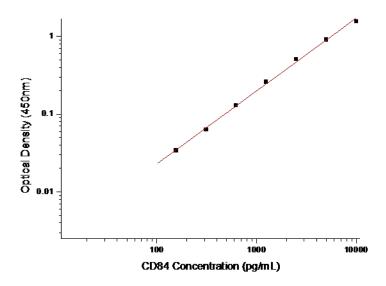
- 1. Add 100  $\mu$ L of sample or standards in sample dilution buffer per well. Seal the plate and incubate 2 hours at room temperature.
- 2. Repeat the aspiration/wash as in step 2 of plate preparation.
- 3. Add  $100~\mu L$  of the detection antibody, diluted in antibody dilution buffer, to each well. Seal the plate and incubate 1 hour at room temperature.
- 4. Repeat the aspiration/wash as in step 2 of plate preparation.
- $5. Add\ 200\ \mu L$  of substrate solution to each well. Incubate for 20 minutes at room temperature ( **if substrate solution is not as requested, the incubation time should be optimized** ). Avoid placing the plate in direct light.
- 6. Add 50 µL of stop solution to each well. Gently tap the plate to ensure thorough mixing.
- 7. Determine the optical density of each well immediately, using a microplate reader set to 450 nm.

#### CALCULATION OF RESULTS

- Calculate the mean absorbance for each set of duplicate standards, controls and samples. Subtract the mean zero standard absorbance from each.
- Construct a standard curve by plotting the mean absorbance for each standard on the y-axis against the concentration on the x-axis and draw a best fit curve through the points on the graph.
- •To determine the concentration of the unknowns, find the unknowns' mean absorbance value on the y-axis and draw a horizontal line to the standard curve. At the point of intersection, draw a vertical line to the x-axis and read the concentration. If samples have been diluted, the concentration read from the standard curve must be multiplied by the dilution factor.
- Alternatively, computer-based curve-fitting statistical software may also be employed to calculate the concentration of the sample.

#### **TYPICAL DATA**

This standard curve is only for demonstration purposes. A standard curve should be generated for each assay.



Concentration (pg/ml)	Zero standard subtracted OD
0	0.000
156.25	0.034
312.5	0.063
625	0.130
1250	0.260
2500	0.509
5000	0.904
10000	1.551

#### PERFORMANCE CHARACTERISTIC

#### **SENSITIVITY**

The minimum detectable dose of Human CD84/SLAMF was determined to be approximately 156.25 pg/ml. This is defined as at least three times standard deviations above the mean optical density of 10 replicates of the zero standard.

### **TROUBLE SHOOTING**

Problems	Possible Sources	Solutions			
	Incorrect or no Detection Antibody was added	Add appropriate Detection Antibody and continue			
No signal	Substrate solution was not added	Add substrate solution and continue			
	Incorrect storage condition	Check if the kit is stored at recommended condition and used before expiration date			
	Standard was incompletely reconstituted or was inappropriately stored	Aliquot reconstituted standard and store at -80 $^{\circ}\mathrm{C}$			
Poor Standard	Imprecise / inaccurate pipetting	Check / calibrate pipettes			
Curve	Incubations done at inappropriate temperature, timing or agitation	Follow the general ELISA protocol			
	Background wells were contaminated	Avoid cross contamination by using the sealer appropriately			
	The concentration of antigen in samples was too low	Enriching samples to increase the concentration of antigen			
Poor detection value	Samples were ineffective	Check if the samples are stored at cold environment. Detect samples in timely manner			
High Background	Insufficient washes	Use multichannel pipettes without touching the reagents on the plate			
	insufficient wasnes	Increase cycles of washes and soaking time between washes			
	TMB Substrate Solution was contaminated	TMB Substrate Solution should be clear and colorless prior to addition to wells			
	Materials were contaminated.	Use clean plates, tubes and pipettes tips			
Non specificity	Samples were contaminated	Avoid cross contamination of samples			
Non-specificity	The concentration of samples was too high	Try higher dilution rate of samples			

	ELISA Plate Template											
	1	2	3	4	5	6	7	8	9	10	11	12
Α												
В												
С												
D												
E												
F												
G												
Н												

## Human CD84/SLAMF5 Antibody Pair [HRP] Notes