



ELISA PRODUCT INFORMATION & MANUAL

15-deoxy-Delta 12, 14-PGJ2

NBP2-61285

Enzyme-linked Immunosorbent Assay for
quantitative detection of 15-deoxy-Delta 12, 14-
PGJ2. For research use only. Not for diagnostic or
therapeutic procedures.

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Product Manual



Please read
entire booklet
before
proceeding with
the assay.



Carefully note
the handling
and storage
conditions of
each kit
component.



Please contact
Novus
Biologicals
Technical
Support if
necessary.

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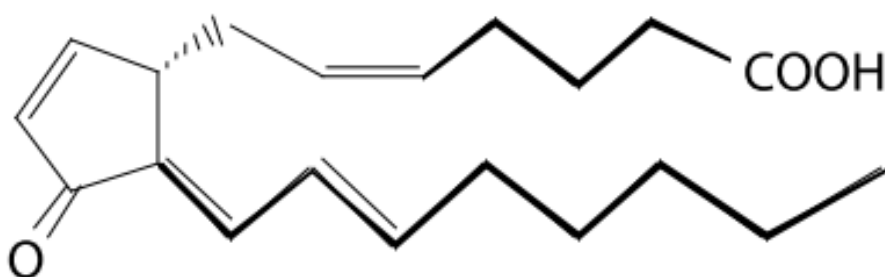
DESCRIPTION

The 15-deoxy- $\Delta^{12,14}$ -PGJ₂ ELISA kit is a competitive immunoassay for the quantitative determination of 15-deoxy- $\Delta^{12,14}$ -Prostaglandin J₂ (15-d-PGJ₂) in biological fluids. Please read the complete kit insert before performing this assay. The kit uses a polyclonal antibody to 15-d-PGJ₂ to bind, in a competitive manner, the 15-d-PGJ₂ in the sample or an alkaline phosphatase molecule which has 15-d-PGJ₂ covalently attached to it. After a simultaneous incubation, the excess reagents are washed away and substrate is added. After a short incubation time the enzyme reaction is stopped and the yellow color generated read on a microplate reader at 405nm. The intensity of the bound yellow color is inversely proportional to the concentration of 15-d-PGJ₂ in either standards or samples. The measured optical density is used to calculate the concentration of 15-d-PGJ₂. For further explanation of the principles and practice of immunoassays please see the excellent books by Chard¹³ or Tijssen¹⁴

INTRODUCTION

15-deoxy- $\Delta^{12,14}$ -Prostaglandin J₂ is one of the ultimate dehydration products of PGD₂. PGD₂ is formed from PGH₂ which itself is synthesized from arachidonic acid by the enzyme prostaglandin synthase¹⁻⁴. In the aqueous solution PGD₂ forms PGJ₂⁵. Albumin or other serum proteins present in the sample will convert PGD₂ to the isomeric compound, Δ^{12} -PGJ₂⁶⁻⁸. In males about 152 ng of Δ^{12} -PGJ₂ is excreted in urine per day, and in women about half this amount is excreted⁹. Excellent recent reviews^{10,11} of the biological actions of these compounds have been published. 15-d-PGJ₂ has been shown as an inducer of adipogenesis and an activator of the γ isoform of the peroxisome proliferation activated receptor (PPAR γ)¹².

15-deoxy- $\Delta^{12,14}$ -Prostaglandin J₂





Handle
with care



Avoid
freeze /
thaw cycles

SAFETY WARNINGS & PRECAUTIONS

FOR RESEARCH USE ONLY. NOT FOR USE IN DIAGNOSTIC PROCEDURES.

- Some kit components contain azide, which may react with lead or copper plumbing. When disposing of reagents always flush with large volumes of water to prevent azide build-up.
- Stop solution is a solution of trisodium phosphate. This solution is caustic; care should be taken in use.
- The activity of the alkaline phosphatase conjugate is dependent on the presence of Mg^{2+} and Zn^{2+} ions. The activity of the conjugate is affected by concentrations of chelators ($>10mM$) such as EDTA and EGTA.
- We test this kit's performance with a variety of samples, however it is possible that high levels of interfering substances may cause variation in assay results.
- The 15-deoxy- $\Delta^{12,14}$ -PGJ₂ standard provided is supplied in ethanolic buffer at a pH optimized to maintain 15-d-PGJ₂ integrity. Care should be taken handling this material because of the known and unknown effects of prostaglandins.

MATERIALS SUPPLIED

- 1. Goat anti-Rabbit IgG Microtiter Plate, One Plate 96 Wells.**
A plate using break-apart strips coated with goat antibody specific to rabbit IgG.
- 2. 15-d-PGJ₂ ELISA Conjugate, 5ml.**
A blue solution of alkaline phosphatase conjugated with 15-d-PGJ₂.
- 3. 15-d-PGJ₂ ELISA Antibody, 5ml.**
A yellow solution of a polyclonal antibody to 15-d-PGJ₂.
- 4. Assay Buffer, 27ml.**
Tris buffered saline, containing proteins and sodium azide as preservative.
- 5. Wash Buffer Concentrate, 27ml.**
Tris buffered saline containing detergents.
- 6. 15-deoxy- $\Delta^{12,14}$ -Prostaglandin J₂ Standard, 0.5ml.**
A solution of 1,000,000pg/ml 15-d-PGJ₂.
- 7. p-Npp Substrate, 20ml.**
A solution of p-nitrophenyl phosphate in buffer. Ready to use.
- 8. Stop Solution, 5ml.**
A solution of trisodium phosphate in water. Keep tightly capped. Caution: Caustic.
- 9. 15-d-PGJ₂ Assay Layout Sheet, 1 each.**
- 10. Plate Sealer, 2 each.**



Reagents
require
separate
storage
conditions.

STORAGE

All components of this kit, except the conjugate and standard, are stable at 4°C until the kit's expiration date. The conjugate and standard must be stored frozen at -20°C.

OTHER MATERIALS NEEDED

1. Deionized or distilled water.
2. Precision pipets for volumes between 5µl and 1,000µl.
3. Repeater pipets for dispensing 50µl and 200µl.
4. Disposable beaker for diluting buffer concentrates.
5. Graduated cylinders.
6. A Microplate shaker.
7. Adsorbent paper for blotting.
8. A 37°C Incubator.
9. Microplate reader capable of reading at 405nm, preferably with correction at between 570 and 590nm.

SAMPLE HANDLING

The 15-deoxy- $\Delta^{12,14}$ -PGJ₂ ELISA is compatible with 15-d-PGJ₂ samples in a wide range of matrices. Samples diluted sufficiently into Assay Buffer can be read directly from the standard curve. Please refer to the Sample Recovery recommendations on page 11 for details of suggested dilutions. However, the end user must verify that the recommended dilutions are appropriate for their samples. Samples containing rabbit IgG may interfere with the assay. Some samples normally have very low levels of 15-d-PGJ₂ present and extraction may be necessary for accurate measurement. A suitable extraction procedure is outlined below:

Materials Needed

1. 15-d-PGJ₂ Standard to allow extraction efficiency to be accurately determined.
2. 2M hydrochloric acid, deionized water, ethanol, hexane and ethyl acetate.
3. 200mg C18 Reverse Phase Extraction Columns.

Procedure

1. Acidify the plasma, urine or tissue homogenate by addition of 2M HCl to pH of 3.5. Approximately 50 μ l of HCl will be needed per ml of plasma. Allow to sit at 4°C for 15 minutes. Centrifuge samples in a microcentrifuge for 2 minutes to remove any precipitate.
2. Prepare the C18 reverse phase column by washing with 10ml of ethanol followed by 10ml of deionized water.
3. Apply the sample under a slight positive pressure to obtain a flow rate of about 0.5ml / minute. Wash the column with 10ml of water, followed by 10ml of 15% ethanol, and finally 10ml hexane. Elute the sample from the column by addition of 10ml ethyl acetate.
4. If analysis is to be carried out immediately, evaporate samples under a stream of nitrogen. Add at least 250 μ l of Assay Buffer to the dried samples. Vortex well, then allow to sit for 5 minutes at room temperature. Repeat twice. If analysis is to be delayed, store samples as the eluted ethyl acetate solutions at -80°C until the immunoassay is to be run. Evaporate the organic solvent under a stream of nitrogen prior to running assay and reconstitute as above.

Please refer to references 15-18 for details of extraction protocols.

PROCEDURAL NOTES

1. Do not mix components from different kit lots or use reagents beyond the kit expiration date.
2. Allow all reagents to warm to room temperature for at least 30 minutes before opening.
3. Standards can be made up in either glass or plastic tubes.
4. Pre-rinse the pipet tip with reagent, use fresh pipet tips for each sample, standard and reagent.
5. Pipet standards and samples to the bottom of the wells.
6. Add the reagents to the side of the well to avoid contamination.
7. This kit uses break-apart microtiter strips, which allow the user to measure as many samples as desired. Unused wells must be kept desiccated at 4°C in the sealed bag provided. The wells should be used in the frame provided.
8. Care must be taken to minimize contamination by endogenous alkaline phosphatase. Contaminating alkaline phosphatase activity, especially in the substrate solution, may lead to high blanks. Care should be taken not to touch pipet tips and other items that are used in the assay with bare hands.
9. Prior to the addition of substrate, ensure that there is no residual wash buffer in the wells. Any remaining wash buffer may cause variation in the assay results.

REAGENT PREPARATION

1. 15-d-PGJ₂ Standard

Allow the 1,000,000pg/ml 15-d-PGJ₂ standard solution to warm to room temperature. Label six 12 x 75mm glass tubes #1 through #6. Pipet 800µl of standard diluent (Assay Buffer or Tissue Culture Media) into tube #1. Pipet 750µl of standard diluent into tubes #2 through #6. Add 200µl of the 1,000,000pg/ml standard to tube #1. Vortex thoroughly. Add 250µl of tube #1 to tube #2 and vortex thoroughly. Continue this for tubes #2 through #6. The concentration of 15-d-PGJ₂ in tubes #1 through #6 will be 200,000, 50,000, 12,500 3,125, 781 and 195pg/ml respectively. See 15-d-PGJ₂ Assay Layout Sheet for dilution details. Diluted Standards should be used within 60 minutes of preparation.

2. 15-d-PGJ₂ Conjugate

Allow the conjugate to warm to room temperature. Any unused conjugate should be aliquoted and re-frozen at or below -20°C. Avoid repeated freeze/thaws of the aliquots.

3. Conjugate 1:10 Dilution for Total Activity Measurement

Prepare the Conjugate 1:10 Dilution by diluting 50µl of the supplied conjugate with 450µl of Assay Buffer. The dilution should be used within 3 hours of preparation. This 1:10 dilution is intended for use in the Total Activity wells ONLY.

4. Wash Buffer

Prepare the Wash Buffer by diluting 5ml of the supplied concentrate with 95ml of deionized water. This can be stored at room temperature until the kit expiration date, or for 3 months, whichever is earlier.

ASSAY PROCEDURE

Bring all reagents to room temperature for at least 30 minutes prior to opening. All standards and samples should be run in duplicate.

1. Refer to the Assay Layout Sheet to determine the number of wells to be used and put any remaining wells with the desiccant back into the pouch and seal the ziploc. Store unused wells at 4°C.
2. Pipet 100µl of standard diluent (Assay Buffer or Tissue Culture Media) into the NSB and the Bo (0pg/ml Standard) wells.
3. Pipet 100µl of Standards #1 through #6 into the appropriate wells.
4. Pipet 100µl of the Samples into the appropriate wells.
5. Pipet 50µl of Assay Buffer into the NSB wells.
6. Pipet 50µl of blue Conjugate into each well, except the Total Activity (TA) and Blank wells.
7. Pipet 50µl of yellow Antibody into each well, except the Blank, TA and NSB wells.

NOTE: Every well used should be Green in color except the NSB wells which should be Blue. The Blank and TA wells are empty at this point and have no color.

8. Incubate the plate at room temperature on a plate shaker for 2 hours at ~500rpm. The plate may be covered with the plate sealer provided, if so desired.
9. Empty the contents of the wells and wash by adding 400µl of wash solution to every well. Repeat the wash 2 more times for a total of 3 Washes.
10. After the final wash empty or aspirate the wells and firmly tap the plate on a lint free paper towel to remove remaining wash buffer.
11. Add 5µl of the light blue Conjugate 1:10 Dilution (see Step 3, Reagent Preparation, on Page 8) to the TA wells.
12. Add 200µl of the pNpp Substrate solution to every well. Cover plate with the plate sealer provided and incubate at 37°C for 3 hours.
13. Add 50µl of Stop Solution to every well. This stops the reaction and the plate should be read immediately.
14. Blank the plate reader against the Blank wells, read the optical density at 405nm preferably with correction between 570 and 590nm. If the plate reader is not able to be blanked against the Blank wells, manually subtract the mean optical density of the Blank wells from all readings.

CALCULATION OF RESULTS

Several options are available for the calculation of the concentration of 15-d-PGJ₂ in the samples. We recommend that the data be handled by an immunoassay software package utilizing a 4 parameter logistic curve fitting program. If this sort of data reduction software is not readily available, the concentration of 15-d-PGJ₂ can be calculated as follows:

1. Calculate the average net Optical Density (OD) bound for each standard and sample by subtracting the average NSB OD from the average OD bound:

$$\text{Average Net OD} = \text{Average Bound OD} - \text{Average NSB OD}$$

2. Calculate the binding of each pair of standard wells as a percentage of the maximum binding wells (Bo), using the following formula:

$$\text{Percent Bound} = \frac{\text{Net OD}}{\text{Net Bo OD}} \times 100$$

3. Plot Percent Bound versus Concentration of 15-d-PGJ₂ for the standards. Approximate a straight line through the points. The concentration of 15-d-PGJ₂ in the unknowns can be determined by interpolation.

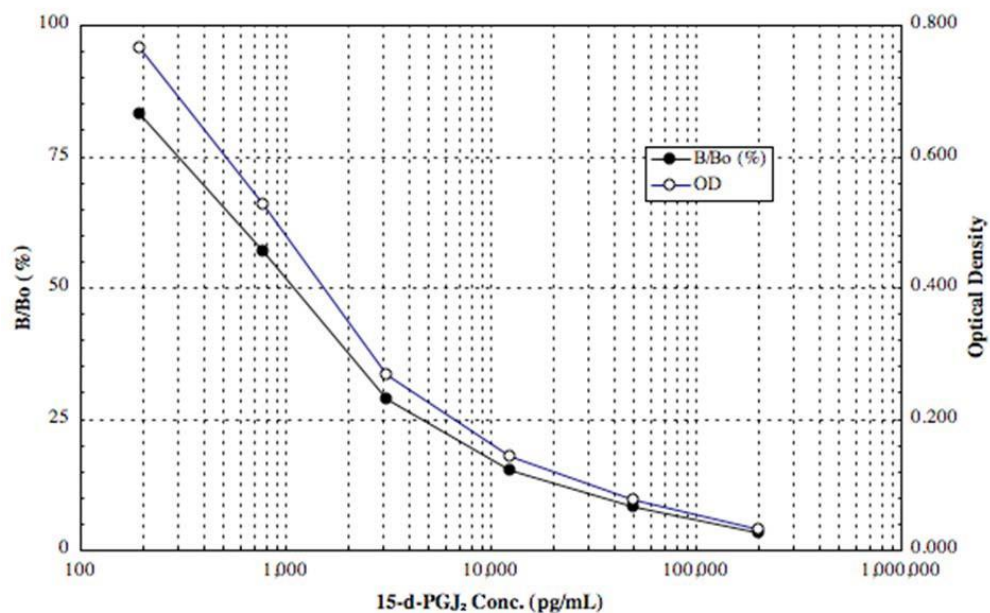
TYPICAL RESULTS

The results shown below are for illustration only and should not be used to calculate results from another assay.

Sample	Mean OD (-Blank)	Average Net OD	Percent Bound	15-d-PGJ ₂ (pg/mL)
Blank OD	(0.134)			
TA	2.696			
NSB	0.009	0.000	0.00%	
Bo	0.932	0.923	100%	0
S1	0.034	0.031	3.4%	200,000
S2	0.086	0.077	8.3%	50,000
S3	0.151	0.142	15.4%	12,500
S4	0.276	0.267	28.9%	3,125
S5	0.535	0.526	57.0%	781
S6	0.775	0.766	83.0%	195
Unknown 1	0.624	0.615	66.6%	486
Unknown 2	0.102	0.093	10.1%	23,638

TYPICAL STANDARD CURVES

A typical standard curve is shown below. This curve must not be used to calculate 15-d-PGJ₂ concentrations; each user must run a standard curve for each assay.



Typical Quality Control Parameters

Total Activity Added =	2.696 x 10 x 10 = 269.6
%NSB =	0.0%
%Bo/TA =	0.3%
Quality of Fit =	0.9997 (Calculated from 4 parameter logistic curve fit)
20% Intercept =	7,636pg/ml
50% Intercept =	1,104pg/ml
80% Intercept =	247pg/ml

PERFORMANCE CHARACTERISTICS

The following parameters for this kit were determined using the guidelines listed in the National Committee for Clinical Laboratory Standards (NCCLS) Evaluation Protocols¹⁹.

Sensitivity

Sensitivity was calculated by determining the average optical density bound for sixteen (16) wells run as Bo, and comparing to the average optical density for sixteen (16) wells run with Standard #6. The detection limit was determined as the concentration of 15-d-PGJ₂ measured at two (2) standard deviations from the zero along the standard curve.

Average Optical Density for the Bo = 1.300 ± 0.030 (2.3%)

Average Optical Density for Standard #6 = 0.983 ± 0.050 (5.1%)

Delta Optical Density (0-195pg/ml) = 0.318

2 SD's of the Zero Standard = 2 x 0.030 = 0.060

Sensitivity = $\frac{0.060}{0.318} \times 195\text{pg/ml}$ **36.8pg/ml**

Linearity

A sample containing 13,011pg/ml 15-d-PGJ₂ was diluted 4 times 1:2 in the kit Assay Buffer and measured in the assay. The data was plotted graphically as actual 15-d-PGJ₂ concentration versus measured 15-d-PGJ₂ concentration.

The line obtained had a slope of 0.974 and a correlation coefficient of 0.998.

Precision

Intra-assay precision was determined by taking samples containing low, medium and high concentrations of 15-d-PGJ₂ and running these samples multiple times (n=16) in the same assay. Inter-assay precision was determined by measuring three samples with low, medium and high concentrations of 15-d-PGJ₂ in multiple assays (n=8).

The precision numbers listed below represent the percent coefficient of variation for the concentrations of 15-d-PGJ₂ determined in these assays as calculated by a 4 parameter logistic curve fitting program.

	15-d-PGJ ₂ (pg/mL)	Intra-assay %CV	Inter-assay %CV
Low	1,136	5.6	
Medium	3,062	5.7	
High	4,394	7.4	
Low	1,195		15.7
Medium	4,406		13.0
High	7,219		14.5

Cross Reactivities

The cross reactivities for a number of related eicosanoid compounds was determined by dissolving the cross reactant (purity checked by N.M.R. and other analytical methods) in Assay Buffer at concentrations from 500,000 to 5pg/ml. These samples were then measured in the 15-d-PGJ₂ assay, and the measured 15-d-PGJ₂ concentration at 50% B/Bo calculated. The % cross reactivity was calculated by comparison with the actual concentration of cross reactant in the sample and expressed as a percentage.

Compound	Cross Reactivity
15-deoxy - Δ 12,14-PGJ ₂	100%
PGJ ₂	49.2%
Δ 12-PGJ ₂	5.99%
PGD ₂	4.92%
Arachidonic Acid	0.03%
PGE ₂	<0.01%
PGF ₂ α	<0.01%
9 α ,11 β -PGF ₂ α	<0.01%
Thromboxane B ₂	<0.01%
2-Arachidonoylglycerol	<0.01%
Anandamide	<0.01%

SAMPLE RECOVERIES

Please refer to pages 6-8 for Sample Handling recommendations and Standard preparation. 15-d-PGJ₂ concentrations were measured in a variety of different samples including tissue culture media, human saliva, human urine and porcine plasma. 15-d-PGJ₂ was spiked into the samples which were diluted with the appropriate diluent and assayed in the kit. The following results were obtained:

Sample	% Recovery*	Recommended Dilution*
Tissue Culture Media	97.4	1:2
Human Saliva	94.4	None
Human Urine	113.9	1:16
Porcine Plasma	105.0	≥1:4

* See Sample Handling instructions on page 6 for details.

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