

LTF (Chicken) ELISA Kit

Catalog Number KA2124

96 assays

Version: 02

Intended for research use only



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Introduction

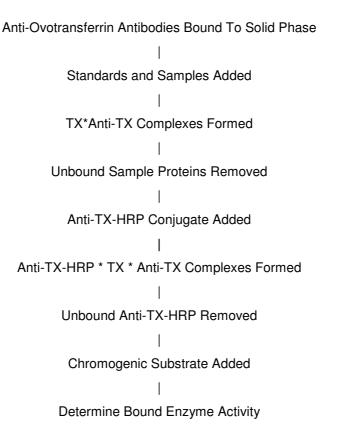
Intended Use

The LTF (Chicken) ELISA Kits are a highly sensitive two-site enzyme linked immunoassay (ELISA) for measuring Ovotransferrin in biological samples of Chickens.

Principle of the Assay

The principle of the double antibody sandwich ELISA is represented in Figure 1. In this assay the Ovotransferrin present in samples reacts with the anti-Ovotransferrin antibodies which have been adsorbed to the surface of polystyrene microtitre wells. After the removal of unbound proteins by washing, anti-Ovotransferrin antibodies conjugated with horseradish peroxidase (HRP), are added. These enzyme-labeled antibodies form complexes with the previously bound Ovotransferrin. Following another washing step, the enzyme bound to the immunosorbent is assayed by the addition of a chromogenic substrate, 3,3',5,5'-tetramethylbenzidine (TMB). The quantity of bound enzyme varies directly with the concentration of Ovotransferrin in the sample tested; thus, the absorbance, at 450 nm, is a measure of the concentration of Ovotransferrin in the test sample. The quantity of Ovotransferrin in the test sample can be interpolated from the standard curve constructed from the standards, and corrected for sample dilution.

Figure 1.





General Information

Materials Supplied

List of component

Component	Amount		
Diluent Concentrate (Running Buffer): One bottle containing 5X concentrated diluent			
running buffer.			
Wash Solution Concentrate: One bottle containing 20X concentrated wash solution			
Enzyme-Antibody Conjugate: One vial containing 100X concentrated affinity-purified	150 μL		
anti-Chicken Ovotransferrin antibody conjugated with HRP in stabilizing buffer.			
Chromogenic Substrate Solution: One vial containing 3,3',5,5'-tetramethybenzidine (TMB)			
and hydrogen peroxide in citric acid buffer at pH 3.3.	12 mL		
Stop Solution: One vial containing 0.3 M sulfuric acid. WARNING: Avoid contact with skin	12 mL		
Anti-Chicken Ovotransferrin ELISA micro plate: Twelve removable eight (8) well strips in	OC wells		
well holder frame. Wells are coated with affinity-purified anti-Chicken Ovotransferrin	96 wells		
Chicken Ovotransferrin Calibrator: One vial containing Chicken Ovotransferrin Calibrator			

Storage Instruction

Complete Kit

The expiration date for the kit is stated on the outer label.

Diluent

The 5X Diluent Concentrate is stable until the expiration date. The 1X working solution is stable for at least one week from the date of preparation. Both solutions should be stored at 4-8 ℃.

Wash Solution

The 20X Wash Solution Concentrate is stable until the expiration date. The 1X working solution is stable for at least one week from the date of preparation. Both solutions can be stored at room temperature (16-25 ℃) or at 4-8 ℃.

Enzyme-Antibody Conjugate

Undiluted horseradish peroxidase anti-Ovotransferrin conjugate should be stored at 4-8 ℃ and diluted immediately prior to use. The working conjugate solution is stable for up to 1 hour when stored in the dark.

Chromogen-Substrate Solution

The Substrate Solution should be stored at 4-8 °C and is stable until the expiration date.

Stop Solution

The Stop Solution should be stored at 4-8 °C and is stable until the expiration date.



- Anti-Chicken Ovotransferrin Elisa Micro Plate
 Anti-Chicken Ovotransferrin coated wells are stable until the expiration date, and should be stored at 4-8 ℃ in sealed foil pouch with desiccant pack.
- Chicken Ovotransferrin Calibrator

 Long Term Storage: Upon receipt, aliquot the calibrator and store them frozen. They will be stable until expiration date. Short Term Storage: the calibrator is stable for up to 14 days at 4℃. The working standard solutions should be prepared immediately prior to use and are stable for up to 8 hours.

Materials Required but Not Supplied

- ✓ Precision pipettes (2 μL to 200 μL) for making and dispensing dilutions
- ✓ Test tubes
- ✓ Microplate washer/aspirator
- ✓ Distilled or Deionized H₂O
- ✓ Microplate reader
- ✓ Assorted glassware for the preparation of reagents and buffer solutions
- ✓ Timer

Precautions for Use

Precausion

For any sample that might contain pathogens, care must be taken to prevent contact with open wounds.

Additives and Preservatives

No additives or preservatives are necessary to maintain the integrity of the specimen. Avoid azide contamination.

• Known interfering substances

Azide and thimerosal at concentrations higher than 0.1% inhibits the enzyme reaction.

- Limitation of the procedure
- ✓ Reliable and reproducible results will be obtained when the assay procedure is carried out with a complete
 understanding of the information contained in the package insert instructions and with adherence to good
 laboratory practice.
- ✓ Factors that might affect the performance of the assay include proper instrument function, cleanliness of glassware, quality of distilled or deionized water, and accuracy of reagent and sample pipettings, washing technique, incubation time or temperature.
- ✓ Do not mix or substitute reagents with those from other lots or sources.



Assay Protocol

Reagent Preparation

Diluent Concentrate

The Diluent Solution supplied is a 5X Concentrate and must be diluted 1/5 with distilled or deionized water (1 part buffer concentrate, 4 parts dH₂O).

Wash Solution Concentrate

The Wash Solution supplied is a 20X Concentrate and must be diluted 1/20 with distilled or deionized water (1 part buffer concentrate, 19 parts dH₂O). Crystal formation in the concentrate is not uncommon when storage temperatures are low. Warming of the concentrate to 30-35 ℃ before dilution can dissolve crystals.

• Enzyme-Antibody Conjugate

Calculate the required amount of working conjugate solution for each microtitre plate test strip by adding 10 µl Enzyme-Antibody Conjugate to 990 µl of 1X Diluent for each test strip to be used for testing. Mix uniformly, but gently. Avoid foaming.

• Chromogen-Substrate Solution

Ready to use as supplied.

Stop Solution

Ready to use as supplied.

• Anti-Chicken Ovotransferrin Elisa Micro Plate

Ready to use as supplied. Unseal Microtiter Pouch and remove plate from pouch. Remove all strips and wells that will not be used in the assay and place back in pouch and re-seal along with desiccant.

Chicken Ovotransferrin Calibrator

The calibrator is now at a concentration of 978 ug/ml. Chicken Ovotransferrin standards need to be prepared immediately prior to use (see chart below). Mix well between each step. Avoid foaming.

Standard	ng/mL	Volume added to 1X Diluent	Volume of 1X Diluent	
Α	9780	5 μl Chicken Ovotransferrin Calibrator	495 µl	
7	400	40 µl standard A	938 µl	
6	200	300 μl standard 7	300 µl	
5	100	300 μl standard 6	300 µl	
4	50	300 μl standard 5	300 µl	
3	25	300 μl standard 4	300 µl	
2	12.5	300 μl standard 3	300 µl	
1	6.25	300 μl standard 2	300 µl	
0	0		600 µl	



Sample Preparation

Blood should be collected by venipuncture. The serum should be separated from the cells after clot formation by centrifugation. For plasma samples, blood should be collected into a container with an anticoagulant and then centrifuged. Care should be taken to minimize hemolysis, excessive hemolysis can impact your results. Assay immediately or aliquot and store samples at -20 °C. Avoid repeated freeze thaw cycles.

Dilution of Samples

The assay for quantification of Ovotransferrin in samples requires that each test sample be diluted before use. For a single step determination a dilution of 1/10,000 is appropriate for most serum/plasma samples. For absolute quantification, samples that yield results outside the range of the standard curve, a lesser or greater dilution might be required. If unsure of sample level, a serial dilution with one or two representative samples before running the entire plate is highly recommended.

To prepare a 1/10,000 dilution of sample, transfer 5 μ l of sample to 495 μ l of 1X diluent. This gives you a 1/100 dilution. Next, dilute the 1/100 dilution by transferring 5 μ l to 495 μ l of 1X dilutent. This gives you a 1/10,000 dilution. Mix thoroughly at each step.

Assay Procedure

- 1. Bring all reagents to room temperature before use.
- 2. Pipette 100 µl of

Standard 0 (0.0 ng/ml) in duplicate

Standard 1 (6.25 ng/ml) in duplicate

Standard 2 (12.5 ng/ml) in duplicate

Standard 3 (25 ng/ml) in duplicate

Standard 4 (50 ng/ml) in duplicate

Standard 5 (100 ng/ml) in duplicate

Standard 6 (200 ng/ml) in duplicate

Standard 7 (400 ng/ml) in duplicate

- 3. Pipette 100 μ I of sample (in duplicate) into pre designated wells.
- 4. Incubate the micro titer plate at room temperature (16-25°C) for thirty (30 ± 2) minutes. Keep plate covered and level during incubation.
- 5. Following incubation, aspirate the contents of the wells.
- 6. Completely fill each well with appropriately diluted Wash Solution and aspirate. Repeat three times, for a total of four washes. If washing manually: completely fill wells with wash buffer, invert the plate then pour/shake out the contents in a waste container. Follow this by sharply striking the wells on absorbent paper to remove residual buffer. Repeat 3 times for a total of four washes.
- 7. Pipette 100 µl of appropriately diluted Enzyme-Antibody Conjugate to each well. Incubate at room



temperature (16-25 °C) for twenty (20 \pm 2) minutes. Keep plate covered in the dark and level during incubation.

- 8. Wash and blot the wells as described in Steps 5/6.
- 9. Pipette 100 µl of TMB Substrate Solution into each well.
- 10. Incubate in the dark at room temperature for precisely ten (10) minutes.
- 11. After ten minutes, add 100 µl of Stop Solution to each well.
- 12. Determine the absorbance (450 nm) of the contents of each well. Calibrate the plate reader to manufacturer's specification.

Stability of the final reaction mixture

The absorbance of the final reaction mixture can be measured up to 2 hours after the addition of the Stop Solution. However, good laboratory practice dictates that the measurement be made as soon as possible.



Data Analysis

Calculation of Results

- 1. Subtract the average background value from the test values for each sample.
- 2. Using the results observed for the standards construct a Standard Curve. The appropriate curve fit is that of a four-parameter logistics curve. A second order polynomial (quadratic) or other curve fits may also be used.
- 3. Interpolate test sample values from standard curve. Correct for sera dilution factor to arrive at the Ovotransferrin concentration in original samples.

Performance characteristics

· Indications of instability

If the test is performing correctly, the results observed with the calibrator solutions should be within 20% of the expected values.



Resources

Plate Layout

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