



Crp (Cat) ELISA Kit

Catalog Number KA2064

96 assays

Version: 01

Intended for research use only

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Introduction and Background

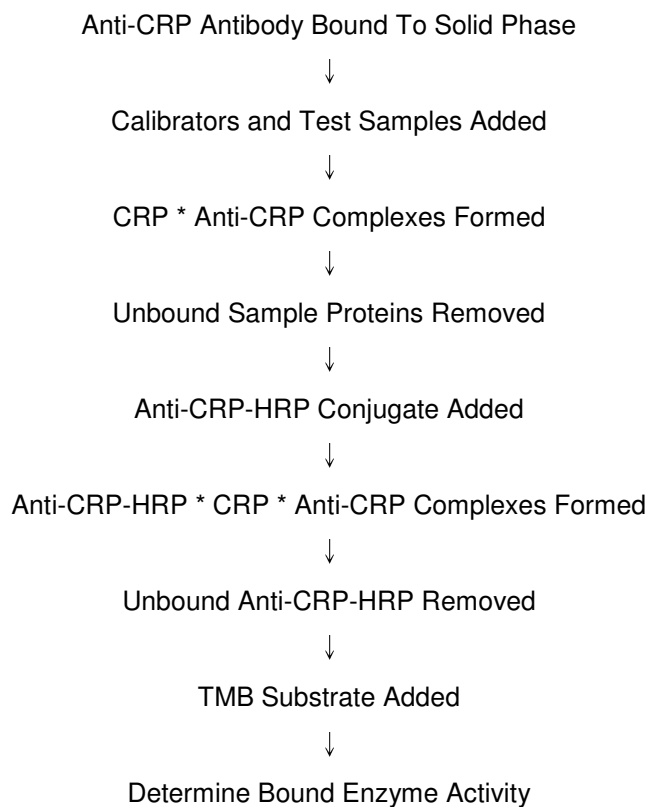
A. Introduction

Acute phase proteins are plasma proteins which increase in concentration following infection, inflammation or trauma. The first acute phase protein to be recognized was discovered in humans by Tillet and Frances in 1930. This C-reactive protein (CRP) is so named because it is able to effect precipitation of somatic C-polysaccharide of *Streptococcus pneumonia*. CRP is an alpha globulin with a mass of 110,000 to 140,000 daltons, and composed of five identical subunits, which are non-covalently assembled as a cyclic pentamer. It is synthesized in the liver and, in humans, is normally present as a trace constituent of serum at levels less than 0.3 mg/dL. The levels in serum rise quickly following acute tissue damage and also fall very rapidly once the stimulus is removed. It has been proposed that the function of CRP is to aid in complement activation, influence phagocytic cell function, and augment cell mediated cytotoxicity. Investigations over the past few years have shown that quantification of these in plasma or serum can provide valuable information in the detection, prognosis, and monitoring of disease not only in humans, but in companion animals and farm herds as well.

B. Test principle

The principle of the double antibody sandwich ELISA is represented in Figure 1. In this assay the CRP present in samples reacts with the anti-CRP antibodies which have been adsorbed to the surface of polystyrene microtiter wells. After the removal of unbound proteins by washing, anti-CRP antibodies conjugated with horseradish peroxidase (HRP), are added. These enzyme-labeled antibodies form complexes with the previously bound CRP. Following another washing step, the enzyme bound to the immunosorbent is assayed by the addition of a chromogenic substrate, 3,3',5,5'-tetramethylbenzidine (TMB). The quantity of bound enzyme varies directly with the concentration of CRP in the sample tested; thus, the absorbance, at 450 nm, is a measure of the concentration of CRP in the test sample. The quantity of CRP in the test sample can be interpolated from the calibration curve constructed from the calibrators, and corrected for sample dilution.

Figure 1.



C. Intended use

The Crp (Cat) ELISA Kit is a highly sensitive two-site enzyme-linked immunoassay (ELISA) for the quantitative determination of CRP in cat biological samples. For research use only.

Material and Method

A. Componets

1. Diluent Concentrate
One bottle containing 50 mL of a 5X concentrated diluent running buffer.
2. Wash Solution Concentrate
One bottle containing 50 mL of a 20X concentrated wash solution.
3. Enzyme-Antibody Conjugate Concentrate
One vial containing 150 μ L of a 100X concentrated affinity-purified anti-cat CRP antibody conjugated with HRP in stabilizing buffer.
4. TMB Substrate Solution
One vial containing 12 mL of TMB and hydrogen peroxide in citric acid buffer at pH 3.3.
5. Stop Solution
One vial containing 12 mL of 0.3 M sulfuric acid. WARNING: Avoid contact with skin.
6. Microtiter Plate
Twelve removable eight-well strips in well holder frame. Wells are coated with affinity-purified anti-cat CRP.
7. Cat CRP Calibrator
One vial containing a Cat CRP Calibrator.

B. Materials required but not provided

- ✓ Precision pipettes (2 μ L to 200 μ L)
- ✓ Test tubes
- ✓ Microplate washer/aspirator
- ✓ Distilled or de-ionized H₂O
- ✓ Microplate reader
- ✓ Assorted glassware for the preparation of reagents and buffer solutions
- ✓ Timer
- ✓ Vortex mixer

C. Precaution

1. Read the instructions carefully before beginning the assay.
2. This kit is for research use only.
3. Great care has been taken to ensure the quality and reliability of this product. However, it is possible that in certain cases, unusual results may be obtained due to high levels of interfering factors.
4. No additives or preservatives are necessary to maintain the integrity of the specimen. Avoid azide contamination.
5. Azide and thimerosal at concentrations higher than 0.1% inhibit the enzyme reaction.
6. Other precautions:

- ✓ Do not interchange kit components from different lots.
- ✓ Do not use kit components beyond the expiration date.
- ✓ Protect reagents from direct sunlight.
- ✓ Do not pipette by mouth.
- ✓ Do not eat, drink, smoke or apply cosmetics where reagents are used.
- ✓ Avoid all contact with the reagents by using gloves.
- ✓ Stop solution contains diluted sulfuric acid. Irritation to eyes and skin is possible. Flush with water after contact.

D. Reagent preparation

1. Diluent Concentrate

The Diluent solution supplied is a 5X concentrate and must be diluted 1:5 with distilled or de-ionized water.

2. Wash Solution Concentrate

The Wash Solution supplied is a 20X concentrate and must be diluted 1:20 with distilled or de-ionized water. Crystal formation in the concentrate is not uncommon when storage temperatures are low. Warming of the concentrate to 30-35°C before dilution can dissolve crystals.

3. Enzyme-Antibody Conjugate Concentrate

The Enzyme-Antibody Conjugate supplied is a 100X concentrate and must be diluted 1:100. The required amount of working conjugate solution for each microtiter plate is prepared by adding 100 µL Enzyme-Antibody Conjugate to 9.9 mL of 1X Diluent. Mix uniformly, but gently. Avoid foaming.

4. TMB Substrate Solution

Ready to use as supplied.

5. Stop Solution

Ready to use as supplied.

6. Microtiter Plate

Ready to use as supplied. Unseal microtiter pouch and remove plate from pouch. Remove all strips and wells that will not be used in the assay and place back in pouch and re-seal._

7. Cat CRP Calibrator

The Cat CRP Calibrator should be aliquoted out and stored frozen. The calibrator is now at a concentration of 84.3 µg/mL and needs to be diluted in 1X diluent according to the chart below for each run. Cat CRP Calibrators need to be prepared immediately prior to use (see chart below). Mix well between each step. Avoid foaming.

Calibrator	Concentration (ng/mL)	Calibrator Volume added to 1X Diluent	Volume of 1X Diluent
7	200	2 μ L Cat CRP Calibrator	841 μ L
6	100	300 μ L Calibrator 7	300 μ L
5	50	300 μ L Calibrator 6	300 μ L
4	25	300 μ L Calibrator 5	300 μ L
3	12.5	300 μ L Calibrator 4	300 μ L
2	6.25	300 μ L Calibrator 3	300 μ L
1	3.125	300 μ L Calibrator 2	300 μ L
0	0		500 μ L

E. Storage and stability

1. Complete Kit

The expiration date for the kit is stated on the outer label. The recommended storage temperature is 4°C.

Note: See long term storage recommendations below for the Cat CRP Calibrator.

2. Diluent

The 5X Diluent Concentrate is stable until the expiration date. The 1X working solution is stable for at least one week from the date of preparation. Both solutions should be stored at 4°C.

3. Wash Solution

The 20X Wash Solution Concentrate is stable until the expiration date. The 1X working solution is stable for at least one week from the date of preparation. Both solutions can be stored at room temperature (RT, 16-25°C) or at 4°C.

4. Enzyme-Antibody Conjugate

Undiluted anti-CRP-HRP conjugate should be stored at 4°C and diluted immediately prior to use. The working conjugate solution is stable for up to 8 hours.

5. TMB Substrate Solution

The TMB Substrate Solution should be stored at 4°C and is stable until the expiration date.

6. Stop Solution

The Stop Solution should be stored at 4°C and is stable until the expiration date.

7. Microtiter Plate

Anti-cat CRP coated wells are stable until the expiration date, and should be stored at 4°C in the sealed foil pouch with desiccant pack.

8. Cat CRP Calibrator

Aliquot Cat CRP calibrator and store them frozen. For storage longer than 14 days, keep frozen until the expiration date. Storage of less than 14 days can be at 4°C. The calibrator is stable until the expiration date but the working calibrator solution is stable for up to 8 hours after preparation.

F. Indication of instability

If the test is performing correctly, the results observed with the calibrator solutions should be within 20% of the expected values.

G. Specimen collection and handling

Blood should be collected by venipuncture and the serum separated from the cells, after clot formation, by centrifugation. For plasma samples, blood should be collected into a container with an anticoagulant and then centrifuged. Care should be taken to minimize hemolysis, excessive hemolysis can impact your results. Assay immediately or aliquot and store samples at -20°C . Avoid repeated freezing/thawing. For any sample that might contain pathogens, care must be taken to prevent contact with open wounds. No additives or preservatives are necessary to maintain the integrity of the specimen. Avoid azide contamination.

H. Assay protocol

✓ **Dilution of Samples**

Due to the high sensitive nature of the assay each sample should be diluted before use for a normal assay. A 1:8,000 dilution of serum or plasma is appropriate for most samples. For absolute quantification of samples that yield results outside the range of the calibration curve, a lesser or greater dilution might be required. If unsure of sample level, a serial dilution with one or two representative samples before running the entire plate is highly recommended. To prepare a 1:8,000 dilution of sample, transfer 5 μL of sample to 495 μL of 1X Diluent. This gives you a 1:100 dilution. Mix thoroughly. Next, dilute the 1:100 samples by transferring 5 μL to 395 μL of 1X Diluent. You now have a 1:8,000 dilution of your sample. Mix thoroughly at each stage.

✓ **Procedure**

1. Bring all reagents to RT before use.
2. Pipette 100 μL of
 - Calibrator 0 (0.0 ng/mL) in duplicate
 - Calibrator 1 (3.125 ng/mL) in duplicate
 - Calibrator 2 (6.25 ng/mL) in duplicate
 - Calibrator 3 (12.5 ng/mL) in duplicate
 - Calibrator 4 (25 ng/mL) in duplicate
 - Calibrator 5 (50 ng/mL) in duplicate
 - Calibrator 6 (100 ng/mL) in duplicate
 - Calibrator 7 (200 ng/mL) in duplicate
3. Pipette 100 μL of diluted sample in duplicate into pre-designated wells.
4. Incubate the Microtiter Plate at 22°C (RT) for thirty (30 ± 2) minutes. Keep plate level and covered during incubation.
5. Following incubation, aspirate the contents of the wells.
6. Completely fill each well with appropriately diluted Wash Solution and aspirate. Repeat three times, for a total of four washes. If washing manually: completely fill wells with diluted Wash Solution, invert the plate and pour/shake out the contents in a waste container. Follow this by sharply striking the wells on absorbent paper to remove residual Wash Solution. Repeat three times for a total of four washes.
7. Pipette 100 μL of appropriately diluted Enzyme-Antibody Conjugate to each well. Incubate at 22°C (RT) for thirty (30 ± 2) minutes. Keep plate covered in the dark and level during incubation.
8. Wash and blot the wells as described in Steps 4 and 5.

9. Pipette 100 μ L of TMB Substrate Solution into each well.
 10. Incubate in the dark at RT for precisely ten (10) minutes.
 11. After ten minutes, add 100 μ L of Stop Solution to each well.
 12. Determine the absorbance at 450 nm of the contents of each well. Zero the plate reader to air.
- The absorbance of the final reaction mixture can be measured up to 2 hours after the addition of the Stop Solution. However, good laboratory practice dictates that the measurement be made as soon as possible.

I. RESULTS

1. Subtract the average background value from the test values for each sample.
2. Using the results observed for the calibrators construct a calibration curve. The appropriate curve fit is that of a four-parameter logistics curve, although a second order polynomial (quadratic) or other curve fits may also be used.
3. Interpolate test sample values from calibration curve. Correct for serum dilution factor to arrive at CRP concentration in original sample.

J. Performance characteristics

In accord with good laboratory practice, the assays for specific CRP require meticulous quality control. Each laboratory should use routine quality control procedures to establish inter- and intra-assay precision and performance characteristics.

K. Limitation of the procedure

1. Reliable and reproducible results will be obtained when the assay procedure is carried out with a complete understanding of the instructions and with adherence to good laboratory practice.
2. Factors that might affect the performance of the assay include proper instrument function, cleanliness of glassware, quality of distilled or de-ionized water, washing technique, incubation time or temperature, and accuracy of reagent and sample pipettings.