IgA (Guinea Pig) ELISA Kit

Catalog Number KA2032

96 assays

Version: 03

Intended for research use only

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Introduction

Intended Use

The IgA (Guinea Pig) ELISA Kit is a highly sensitive two-site enzyme linked immunoassay (ELISA) for measuring IgA in biological samples of Guinea Pigs.

Principle of the Assay

The principle of the double antibody sandwich ELISA is represented in Figure 1. In this assay the IgA present in samples reacts with the anti-IgA antibodies which have been adsorbed to the surface of polystyrene microtiter wells. After the removal of unbound proteins by washing, anti-IgA antibodies conjugated with horseradish peroxidase (HRP), are added. These enzyme-labeled antibodies form complexes with the previously bound IgA. Following another washing step, the enzyme bound to the immunosorbent is assayed by the addition of a chromogenic substrate, 3,3',5,5'- tetramethylbenzidine (TMB). The quantity of bound enzyme varies directly with the concentration of IgA in the sample tested; thus, the absorbance, at 450 nm, is a measure of the concentration of IgA in the test sample. The quantity of IgA in the test sample can be interpolated from the standard curve constructed from the standards, and corrected for sample dilution.

Figure 1.

Anti-IgA Antibody Bound To Solid Phase

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Standards and Samples Added

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IgA * Anti-IgA Complexes Formed

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Unbound Sample Proteins Removed

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Anti-IgA-HRP Conjugate Added

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Anti-IgA-HRP * IgA * Anti-IgA Complexes Formed

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Unbound Anti-IgA-HRP Removed

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Chromogenic Substrate Added

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Determine Bound Enzyme Activity

General Information

Materials Supplied

List of component

Component	Amount
Diluent Concentrate (Running Buffer): One bottle containing 5X concentrated diluent running buffer.	50 mL
Wash Solution Concentrate: One bottle containing 20X concentrated wash solution.	50 mL
Enzyme-Antibody Conjugate 100X: One vial affinity-purified anti-guinea pig IgA antibody conjugated with horseradish peroxidase in stabilizing buffer.	150 µL
Chromogenic Substrate Solution: One vial containing 3,3',5,5'-tetramethybenzidine (TMB) and hydrogen peroxide in citric acid buffer at pH 3.3.	12 mL
Stop Solution: One vial containing 0.3 M sulfuric acid. WARNING: Avoid contact with skin	12 mL
Anti- Guinea Pig IgA ELISA micro plate: Twelve removable eight (8) well micro well strips in well holder frame. Each well is coated with affinity-purified anti- Guinea Pig IgA.	96 wells
Guinea Pig IgA Calibrator: One vial containing a lyophilized Guinea Pig IgA Calibrator.	1 vial

Storage Instruction

The expiration date for the package is stated on the box label.

- ✓ Diluent

  The 5X Diluent Concentrate is stable until the expiration date. The 1X working solution is stable for at least one week from the date of preparation. Both solutions should be stored at 4-8°C.

- ✓ Wash Solution

  The 20X Wash Solution Concentrate is stable until the expiration date. The 1X working solution is stable for at least one week from the date of preparation. Both solutions can be stored at room temperature (16-25°C) or at 4-8°C.

- ✓ Enzyme-Antibody Conjugate

  Undiluted horseradish peroxidase anti-IgA conjugate should be stored at 4-8°C and diluted immediately prior to use. The working conjugate solution is stable for up to 1 hour when stored in the dark.

- ✓ Chromogen-Substrate Solution

  The Substrate Solution should be stored at 4-8°C and is stable until the expiration date.

- ✓ Stop Solution

  The Stop Solution should be stored at 4-8°C and is stable until the expiration date.

- ✓ Anti-Guinea Pig IgA ELISA Micro Plate

  Anti-Guinea Pig IgA coated wells are stable until the expiration date, and should be stored at 4-8°C in the sealed foil pouch with desiccant pack.

✓ Guinea Pig IgA Standards

The lyophilized Guinea Pig IgA Calibrator should be stored at 4°C or frozen until reconstituted. The reconstituted calibrator should be aliquoted out and stored frozen (avoid multiple freeze-thaw cycles). The working standard solutions should be prepared immediately prior to use and are stable for up to 8 hours.

Materials Required but Not Supplied

- ✓ Precision pipettes (2 µL to 200 µL) for making and dispensing dilutions
- ✓ Test tubes
- ✓ Microplate washer/aspirator
- ✓ Distilled or Deionized H_2O
- ✓ Microtitre Plate reader
- ✓ Assorted glassware for the preparation of reagents and buffer solutions
- ✓ Timer

Precautions for Use

✓ Precausion

For any sample that might contain pathogens, care must be taken to prevent contact with open wounds.

✓ Additives and Preservatives

No additives or preservatives are necessary to maintain the integrity of the specimen. Avoid azide contamination.

✓ Known interfering substances

Azide and thimerosal at concentrations higher than 0.1% inhibits the enzyme reaction.

✓ Limitation of the procedure

- Reliable and reproducible results will be obtained when the assay procedure is carried out with a complete understanding of the information contained in the package insert instructions and with adherence to good laboratory practice.
- Factors that might affect the performance of the assay include proper instrument function, cleanliness of glassware, quality of distilled or deionized water, and accuracy of reagent and sample pipettings.

Assay Protocol

Reagent Preparation

- ✓ Diluent Concentrate

  The Diluent solution supplied is a 5X concentrate and must be diluted 1/5 (1 part buffer concentrate, 4 parts dH_2O) with distilled or de-ionized water.
- ✓ Wash Solution Concentrate

  The Wash Solution supplied is a 20X concentrate and must be diluted 1/20 with distilled or de-ionized water (1 part buffer concentrate, 19 parts dH_2O). Crystal formation in the concentrate is not uncommon when storage temperatures are low. Warming of the concentrate to 30-35°C before dilution can dissolve crystals.
- ✓ Enzyme-Antibody Conjugate

  Calculate the required amount of working conjugate solution for each microtiter plate test strip by adding 10 µL Enzyme-Antibody Conjugate to 990 µL of 1X Diluent for each test strip to be used for testing. Mix uniformly, but gently. Avoid foaming.
- ✓ Chromogen-Substrate Solution

  Ready to use as supplied.
- ✓ Stop Solution

  Ready to use as supplied.
- ✓ Anti-Guinea Pig IgA ELISA Micro Plate

  Ready to use as supplied. Unseal microtiter pouch and remove plate from pouch. Remove all strips and wells that will not be used in the assay and place back in pouch and re-seal. Take clear plastic tape and cover tops of strips to avoid particulates from contaminating wells.
- ✓ Guinea Pig IgA Standards

  Add 1.0 mL of distilled or de-ionized water to the Guinea Pig IgA Calibrator and mix gently until dissolved. The Calibrator is now at a concentration of 1.85 µg/mL (the reconstituted calibrator should be aliquoted and frozen if future use is intended). Guinea Pig IgA standards need to be prepared immediately prior to use (see the following chart). Mix well between each step. Avoid foaming.

Standard	ng/mL	Volume added to 1x Diluent	Volume of 1x Diluent
6	200	80 µL of Guinea Pig IgA Calibrator	660 µl
5	100	300 µl standard 6	300 µl
4	50	300 µl standard 5	300 µl
3	25	300 µl standard 4	300 µl
2	12.5	300 µl standard 3	300 µl
1	6.25	300 µl standard 2	300 µl
0	0		600 µl

Sample Preparation

✓ Specimen Collection and Handling

Blood should be collected by venipuncture and the serum separated from the cells, after clot formation, by centrifugation. Care should be taken to minimize hemolysis, excessive hemolysis can impact your results. Specimens may be shipped at room temperature and then stored refrigerated at 2-8°C if testing is to take place within one week after collection. If testing is to take place later than one week, specimens should be stored at -20°C. Avoid repeated freeze-thaw cycles.

✓ Dilution of Serum Samples

The assay for quantification of IgA in samples requires that each test sample be diluted before use. For a single step determination a dilution of serum at 1/2,000 is appropriate for most samples. For absolute quantification, samples that yield results outside the range of the standard curve, a lesser or greater dilution might be required. If unsure of sample level, a serial dilution with one or two representative samples before running the entire plate is highly recommended.

To prepare a 1/2,000 dilution of sample, transfer 5 µL of sample to 495 µL of 1X diluent. This gives you a 1/100 dilution. Next, dilute the 1/100 samples by transferring 20 µL, to 380 µL of 1X diluent. You now have a 1/2,000 dilution of your sample. Mix thoroughly at each stage.

Assay Procedure

1. Bring all reagents to room temperature before use.
2. Pipette 100 µL of
   Standard 0 (0.0 ng/ml) in duplicate
   Standard 1 (6.25 ng/ml) in duplicate
   Standard 2 (12.5 ng/ml) in duplicate
   Standard 3 (25 ng/ml) in duplicate
   Standard 4 (50 ng/ml) in duplicate
   Standard 5 (100 ng/ml) in duplicate
   Standard 6 (200 ng/ml) in duplicate
3. Pipette 100 µL of sample (in duplicate) into pre-designated wells.
4. Incubate the micro titer plate at room temperature for twenty (20 ± 2) minutes. Keep plate covered and level during incubation.
5. Following incubation, aspirate the contents of the wells.
6. Completely fill each well with appropriately diluted Wash Solution and aspirate. Repeat three times, for a total of four washes. If washing manually: completely fill wells with wash buffer, invert the plate and pour/shake out the contents in a waste container. Follow this by sharply striking the wells on absorbent

paper to remove residual buffer. Repeat 3 times for a total of four washes.

7. Pipette 100 µL of appropriately diluted Enzyme-Antibody Conjugate to each well. Incubate at room temperature for twenty (20 ± 2) minutes. Keep plate covered and level during incubation.
8. Wash and blot the wells as described in Steps 5/6.
9. Pipette 100 µL of TMB Substrate Solution into each well.
10. Incubate in the dark at room temperature for precisely ten (10) minutes.
11. After ten minutes, add 100 µL of Stop Solution to each well.
12. Determine the absorbance (450 nm) of the contents of each well. Calibrate the plate reader to manufacturer's specification.

✓ Stability of the final reaction mixture

The absorbance of the final reaction mixture can be measured up to 2 hours after the addition of the Stop Solution. However, good laboratory practice dictates that the measurement be made as soon as possible.

Data Analysis

Calculation of Results

1. Subtract the average background value from the test values for each sample.
2. Using the results observed for the standards construct a standard curve. The appropriate curve fit is that of a four-parameter logistics curve, although a second order polynomial (quadratic) or other curve fits may also be used.
3. Interpolate test sample values from standard curve. Correct for sera dilution factor to arrive at IgA concentration in original sample.

Resources

Plate Layout

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6								
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3								
2								
1								
	A	B	C	D	E	F	G	H