



# Phospholipase D Assay Kit

Catalog Number KA1636

100 assays

Version: 03

Intended for research use only

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## **Introduction**

### **Intended Use**

Applications:

- ✓ Direct Assays: phospholipase D in biological samples.
- ✓ Drug Discovery/Pharmacology: effects of drugs on phospholipase D metabolism.

Features:

- ✓ Sensitive. Use 10 µL samples. Detection range: colorimetric assay 0.06 - 10 U/L, fluorimetric assay 0.04 - 1 U/L.
- ✓ Simple and High-throughput: the assay involves addition of a single working reagent and can be readily adapted to high-throughput assays for drug screening.

### **Background**

PHOSPHOLIPASE D (PLD) catalyses the hydrolysis of the phosphodiester bond of glycerophospholipids to generate phosphatidic acid and a free headgroup. Abnormalities in PLD expression have been associated with human cancers.

### **Principle of the Assay**

In this assay, PLD hydrolyzes phosphatidylcholine to choline which is determined using choline oxidase and a H<sub>2</sub>O<sub>2</sub> specific dye. The optical density of the pink colored product at 570nm or fluorescence intensity (530/585 nm) is directly proportional to the PLD activity in the sample.

## General Information

### **Materials Supplied**

List of component

Component	Amount
Assay Buffer	10 mL
Dye Reagent	120 µL
Enzyme Mix (Dried)	1 vial
Substrate	1.5 mL
Calibrator	400 µL

### **Storage Instruction**

Store all components at -20 °C. Shelf life of six months after receipt.

### **Materials Required but Not Supplied**

Pipetting devices, centrifuge tubes, clear flat-bottom uncoated 96-well plates, optical density plate reader; black flat-bottom uncoated 96-well plates, fluorescence plate reader.

### **Precautions for Use**

Reagents are for research use only. Normal precautions for laboratory reagents should be exercised while using the reagents.

## Assay Protocol

### Assay Procedure

#### ✓ Colorimetric assay

Liquid samples can be assayed directly. Solid samples should be homogenized in a suitable enzyme buffer prior to assay.

*Note: SH-containing reagents (e.g.  $\beta$ -mercaptoethanol, dithiothreitol,  $> 5 \mu\text{M}$ ), sodium azide, EDTA, and sodium dodecyl sulfate are known to interfere in this assay and should be avoided in sample preparation. If a sample is known to contain choline, it should be removed by dialysis or membrane filtration.*

1. Equilibrate all components to room temperature. Briefly centrifuge the tubes before opening. Keep thawed tubes on ice during assay. Reconstitute Enzyme Mix with 120  $\mu\text{L}$  Assay Buffer. Reconstituted Enzyme Mix is stable for 1 month when stored at  $-20^\circ\text{C}$ .
2. Calibrator: mix 33  $\mu\text{L}$  Calibrator with 187  $\mu\text{L}$   $\text{dH}_2\text{O}$  (final 300  $\mu\text{M}$  choline). Dilute calibrator in  $\text{dH}_2\text{O}$  as follows.

No	300 $\mu\text{M}$ Premix + $\text{H}_2\text{O}$	Vol ( $\mu\text{L}$ )	Calibrator ( $\mu\text{M}$ )
1	100 $\mu\text{L}$ + 0 $\mu\text{L}$	100	300
2	60 $\mu\text{L}$ + 40 $\mu\text{L}$	100	180
3	30 $\mu\text{L}$ + 70 $\mu\text{L}$	100	90
4	0 $\mu\text{L}$ + 100 $\mu\text{L}$	100	0

Transfer 10  $\mu\text{L}$  diluted standards into separate wells of a clear flat bottom 96-well plate.

Samples: transfer 10  $\mu\text{L}$  of each sample into separate wells of the plate.

3. Color reaction. Prepare enough Working Reagent by mixing, for each well, 85  $\mu\text{L}$  Assay Buffer, 1  $\mu\text{L}$  Enzyme Mix, 1  $\mu\text{L}$  Dye Reagent and 12  $\mu\text{L}$  Substrate. Add 90  $\mu\text{L}$  Working Reagent to each well.  
Tap plate to mix. Incubate at desired temperature and protect from light. At 10 and 30 min, read optical density 570nm (550-585nm).

#### ✓ Fluorimetric assay

The fluorimetric assay procedure is similar to the colorimetric procedure except that (1) 0, 9, 18 and 30  $\mu\text{M}$  calibrator and (2) a black 96-well plate are used. Read fluorescence intensity at  $\lambda_{\text{ex}} = 530 \text{ nm}$  and  $\lambda_{\text{em}} = 585 \text{ nm}$ .

## Data Analysis

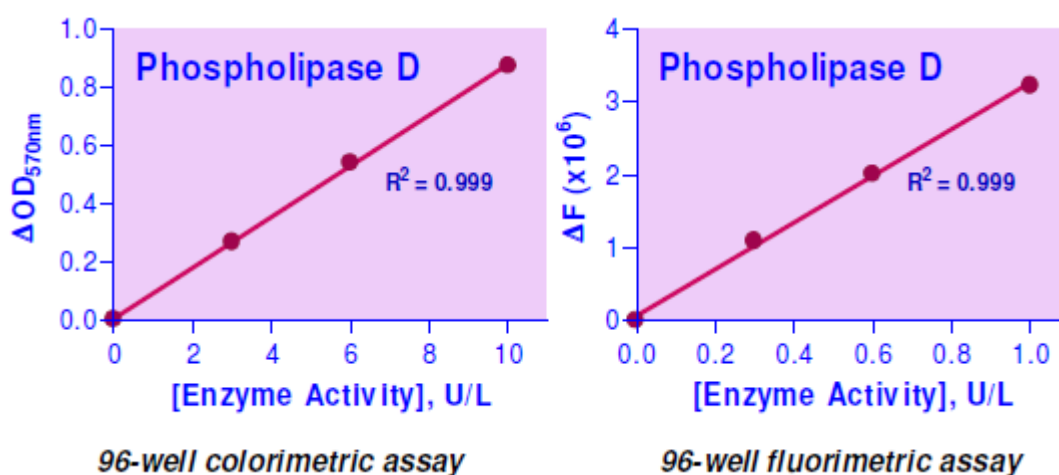
### Calculation of Results

Subtract blank value (#4) from the standard values and plot the  $\Delta OD$  or  $\Delta F$  against standard concentrations. Determine the slope and calculate the PLD activity of Sample,

$$[\text{Phospholipase D}] = \frac{R_{30} - R_{10}}{\text{Slope} \times 20} \times n \text{ (U/L)}$$

$R_{30}$  and  $R_{10}$  are optical density or fluorescence intensity readings of the Sample at 30 min and 10 min, respectively. 20 is the enzyme reaction time (30 min - 10 min).  $n$  is the sample dilution factor. *Note: if the calculated PLD activity of a sample is higher than 10 U/L in the Colorimetric Assay or 1 U/L in the Fluorimetric Assay, dilute sample in assay buffer and repeat the assay. Multiply result by the dilution factor.*

Unit definition: 1 unit of PLD catalyzes formation of 1  $\mu\text{mole}$  of choline per minute under the assay conditions (pH 7.4).



## Resources

### References

1. Su W, Chen Q, Frohman MA. (2009). Targeting phospholipase D with small-molecule inhibitors as a potential therapeutic approach for cancer metastasis. *Future Oncol.* 5(9):1477-86.
2. Carman GM, Fischl AS, Dougherty M, Maerker G. (1981). A spectrophotometric method for the assay of phospholipase D activity. *Anal Biochem.* 110(1):73-6.
3. Imamura S, Horiuti Y. (1978). Enzymatic determination of phospholipase D activity with choline oxidase. *J Biochem.* 83(3):677-80.