

## Introduction and Background

### A. Overview

The protein **CD80** (Cluster of Differentiation **80**) is a molecule found on activated B cells and monocytes which provides a costimulatory signal necessary for T cell activation and survival. It is also known as **B7.1**. The cDNA for B7-1 predicts a type I membrane protein, i.e., one synthesized with a signal peptide that is cleaved upon translocation across the endoplasmic membrane. The protein is predicted to contain 2 extracellular domains structurally similar to those of Ig, a hydrophobic transmembrane region, and a short cytoplasmic domain[1]. The CD80 and CD86 (601020) genes encode B7-1 and B7-2, respectively, which are structurally similar members of the immunoglobulin superfamily expressed on a variety of hematopoietic cell types.[2] stated that B7-1 and B7-2 provide a costimulatory signal to T cells by interacting with CD28 and CTLA4. The standard product used in this kit is recombinant B7-1 · D37—K245 · which is composed of two single chains acids with the dipolymer.

### B. Test Principle

Cd80 (Mouse) ELISA Kit ( Cat # KA0991 ) was based on standard sandwich enzyme-linked immune-sorbent assay technology. Mouse CD80 specific-specific polyclonal antibodies were precoated onto 96-well plates. The mouse specific detection polyclonal antibodies were biotinylated. The test samples and biotinylated detection antibodies were added to the wells subsequently and then followed by washing with PBS or TBS buffer. Avidin-Biotin-Peroxidase Complex was added and unbound conjugates were washed away with PBS or TBS buffer. HRP substrate TMB was used to visualize HRP enzymatic reaction. TMB was catalyzed by HRP to produce a blue color product that changed into yellow after adding acidic stop solution. The density of yellow is proportional to the mouse CD80 amount of sample captured in plate.

### C. Notice for Application of Kit

- ✓ Before using Kit, spin tubes and bring down all components to bottom of tube.
- ✓ Duplicate well assay was recommended for both standard and sample testing.
- ✓ Don't let 96-well plate dry, dry plate will inactivate active components on plate.
- ✓ In order to avoid marginal effect of plate incubation due to temperature difference (reaction may be stronger in the marginal wells), it is suggested that the diluted ABC and TMB solution will be pre-warmed in 37°C for 30 min before using.

✓

### D. Application

For quantitative detection of mouse CD80 in sera, body fluids, tissue lysates or cell culture supernates. The significance of detection in sera, plasma, body fluids is undetermined.

## Material and Method

### A. List of component

1. Lyophilized recombinant mouse CD80 standard: 10ng/tube×2.
2. One 96-well plate precoated with anti- mouse CD80 antibody.
3. Sample diluent buffer: 30 ml
4. Biotinylated anti- mouse CD80 antibody: 130µl, dilution 1:100.
5. Antibody diluent buffer: 12ml.
6. Avidin-Biotin-Peroxidase Complex (ABC): 130µl, dilution 1:100.
7. ABC diluent buffer: 12ml.
8. TMB color developing agent: 10ml.
9. TMB stop solution: 10ml.

### B. Additional Required Materials But Not Provided

1. Microplate reader in standard size.
2. Automated plate washer.
3. Adjustable pipettes and pipette tips. Multichannel pipettes are recommended in the condition of large amount of samples in the detection.
4. Clean tubes and Eppendorf tubes.
5. Washing buffer (neutral PBS or TBS).

#### Preparation of 0.01M TBS:

Add 1.2g Tris, 8.5g NaCl; 450µl of purified acetic acid or 700µl of concentrated hydrochloric acid to 1000ml H<sub>2</sub>O and adjust pH to 7.2-7.6. Finally, adjust the total volume to 1L.

#### Preparation of 0.01 M PBS:

Add 8.5g sodium chloride, 1.4g Na<sub>2</sub>HPO<sub>4</sub> and 0.2g NaH<sub>2</sub>PO<sub>4</sub> to 1000ml distilled water and adjust pH to 7.2-7.6. Finally, adjust the total volume to 1L.

### C. Preparation of reagent

#### Plate Washing

Discard the solution in the plate without touching the side walls. Blot the plate onto paper towels or other absorbent material. Soak each well with at least 0.3 ml PBS or TBS buffer for 1~2 minutes. Repeat this process two additional times for a total of THREE washes.

**Note:** For automated washing, aspirate all wells and wash THREE times with PBS or TBS buffer, overfilling wells with PBS or TBS buffer. Blot the plate onto paper towels or other absorbent material.

#### Sample Preparation and Storage

Store samples to be assayed within 24 hours at 2-8°C. For long-term storage, aliquot and freeze samples at -20°C.

Avoid repeated freeze-thaw cycles.

- **Cell culture supernate, tissue lysate or body fluids:** Remove particulates by centrifugation, analyze immediately or aliquot and store at -20°C
- **Serum:** Allow the serum to clot in a serum separator tube (about 30 min) at room temperature. Centrifuge at approximately 2000 X g for 20 min. Analyze the serum immediately or aliquot and store frozen at -20°C.
- **Plasma:** Collect plasma using heparin, EDTA, citrate as an anticoagulant. Centrifuge at 2-8°C for 20 min at 2000 x g within 30 min of collection. Analyze immediately or aliquot and store frozen at -20°C.

### **Sample Dilution Guideline**

The user needs to estimate the concentration of the target protein in the sample and select a proper dilution factor so that the diluted target protein concentration falls near the middle of the linear regime in the standard curve. Dilute the sample using the provided diluent buffer. The following is a guideline for sample dilution. Several trials may be necessary in practice. **The sample must be well mixed with the diluents buffer.**

- **High target protein concentration (40-400ng/ml).** The working dilution is 1:100. i.e. Add 3µl sample into 297µl sample diluent buffer.
- **Medium target protein concentration (4-40ng/ml).** The working dilution is 1:10. i.e. Add 25µl sample into 225 µl sample diluent buffer.
- **Low target protein concentration (62.5-4000pg/ml).** The working dilution is 1:2. i.e. Add 100 µl sample to 100 µl sample diluent buffer.
- **Very Low target protein concentration (≤62.5pg/ml).** No dilution necessary, or the working dilution is 1:2.

### **Reagent Preparation and Storage**

1. Reconstitution of the mouse CD80 standard: CD80 standard solution should be prepared no more than 2 hours prior to the experiment. Two tubes of CD80 standard (10ng per tube) are included in each kit. Use one tube for each experiment.
  - a. 10,000pg/ml of mouse CD80 standard solution: Add 1 ml sample diluent buffer into one tube, keep the tube at room temperature for 10 min and mix thoroughly.
  - b. 4000pg/ml of mouse CD80 standard solution: Add 0.4 ml of the above 10,000pg/ml CD80 standard solution into 0.6 ml sample Eppendorf tubes and mix thoroughly.
  - c. 2000pg/ml→62.5pg/ml of mouse CD80 standard solutions: Label 6 Eppendorf tubes with 2000pg/ml , 1000pg/ml , 500pg/ml , 250pg/ml , 125pg/ml , 62.5pg/ml, respectively. Aliquot 0.3 ml of the sample diluent buffer into each tube. Add 0.3 ml of the above 4000pg/ml CD80 standard solution into 1st tube and mix. Transfer 0.3 ml from 1st tube to 2nd tube and mix. Transfer 0.3 ml from 2nd tube to

3rd tube and mix, and so on.

**Note:** The standard solutions are best used within 12 hours. The 10 ng/ml standard solution may be stored at 4°C for up to 12 hours, or at -20°C for up to 48 hours. Avoid repeated freeze-thaw cycles.

2. Preparation of biotinylated anti-mouse CD80 antibody working solution: The solution should be prepared no more than 2 hours prior to the experiment.
  - a. The total volume should be: 0.1ml/well x (the number of wells). (Allowing 0.1-0.2 ml more than total volume)
  - b. Biotinylated anti-mouse CD80 antibody should be diluted in 1:99 with the antibody diluent buffer and mixed thoroughly.
3. Preparation of Avidin-Biotin-Peroxidase Complex (ABC) working solution: The solution should be prepared no more than 1 hour prior to the experiment.
  - a. The total volume should be: 0.1ml/well x (the number of wells). (Allowing 0.1-0.2 ml more than total volume)
  - b. Avidin- Biotin-Peroxidase Complex (ABC) should be diluted in 1:99 with the ABC dilution buffer and mixed thoroughly.

#### **D. Stability and storage**

Store at 2-8°C for frequent use, at -20°C for infrequent use.

#### **E. Expiration**

Four months at 4°C and eight months at -20°C.

#### **F. Protocol**

The ABC working solution and TMB color developing agent must be kept warm at 37°C for 30 min before use. When diluting samples and reagents, they must be mixed completely and evenly. Standard CD80 detection curve should be prepared for each experiment. The user will decide sample dilution fold by crude estimation of CD80 amount in samples.

1. Aliquot 0.1ml per well of the 4000pg/ml , 2000pg/ml , 1000pg/ml , 500pg/ml , 250pg/ml , 125pg/ml , 62.5pg/ml mouse CD80 standard solutions into the precoated 96-well plate. Add 0.1ml of the sample diluent buffer into the control well (Zero well). Add 0.1ml of each properly diluted sample of mouse sera, plasma, body fluids, tissue lysates or cell culture supernatants to each empty well. **See “Sample Dilution Guideline” above for details.** We recommend that each mouse CD80 standard solution and each sample is measured in duplicate.
2. Seal the plate with the cover and incubate at 37°C for 90 min.
3. Remove the cover, discard plate content, and blot the plate onto paper towels or other absorbent material. Do NOT let the wells completely dry at any time.
4. Add 0.1ml of biotinylated anti-mouse CD80 antibody working solution into each well and incubate the

plate at 37°C for 60 min.

5. Wash the plate three times with 0.01M TBS or 0.01M PBS, and each time let washing buffer stay in the wells for 1 min. Discard the washing buffer and blot the plate onto paper towels or other absorbent material.
6. Add 0.1ml of prepared ABC working solution into each well and incubate the plate at 37°C for 30 min.
7. Wash plate 5 times with 0.01M TBS or 0.01M PBS, and each time let washing buffer stay in the wells for 1-2 min. Discard the washing buffer and blot the plate onto paper towels or other absorbent material.
8. Add 90 µl of prepared TMB color developing agent into each well and incubate plate at 37°C for 15-20 min (shades of blue can be seen in the wells with the four most concentrated mouse CD80 standard solutions; the other wells show no obvious color).
9. Add 0.1ml of prepared TMB stop solution into each well. The color changes into yellow immediately.
10. Read the O.D. absorbance at 450nm in a microplate reader within 30 min after adding the stop solution.

For calculation, (the relative O.D.<sub>450</sub>) = (the O.D.<sub>450</sub> of each well) – (the O.D.<sub>450</sub> of Zero well). The standard curve can be plotted as the relative O.D.<sub>450</sub> of each standard solution (Y) vs. the respective concentration of the standard solution (X). The mouse CD80 concentration of the samples can be interpolated from the standard curve.

**Note:** if the samples measured were diluted, multiply the dilution factor to the concentrations from interpolation to obtain the concentration before dilution.

### Summary

1. Add samples and standards and incubate the plate at 37°C for 90 min. Do not wash.
2. Add biotinylated antibodies and incubate the plate at 37°C for 60 min. Wash plate 3 times with 0.01M TBS.
3. Add ABC working solution and incubate the plate at 37°C for 30 min. Wash plate 5 times with 0.01M TBS.
4. Add TMB color developing agent and incubate the plate at 37°C for 15-20 min.
5. Add TMB stop solution and read.

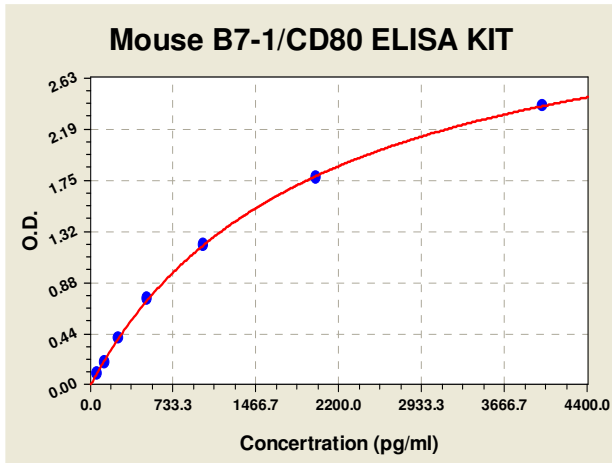
### G. Performance Characteristics

#### Typical result

Typical Data Obtained from Mouse CD80

Concentration pg/ml	0.0	62.5	125	250	500	1000	2000	4000
O.D.	0.002	0.107	0.202	0.397	0.739	1.200	1.777	2.391

(TMB reaction incubate at 37°C for 15 min)



This standard curve was generated for demonstration purpose only. **A standard curve must be run with each assay.**

**Range**

62.5pg/ml-4000pg/ml

**Sensitivity**

< 10pg/ml

**Specificity**

No detectable cross-reactivity with any other cytokine.

**References**

1. Reeves, R. H., Patch, D., Sharpe, A. H., Borriello, F., Freeman, G. J., Edelhoff, S., Distech, C. The costimulatory genes Cd80 and Cd86 are linked on mouse chromosome 16 and human chromosome 3. *Mammalian Genome* 8: 581-582, 1997.
2. Selvakumar, A., Mohanraj, B. K., Eddy, R. L., Shows, T. B., White, P. C., Dupont, B. Genomic organization and chromosomal location of the human gene encoding the B-lymphocyte activation antigen B7. *Immunogenetics* 36: 175-181, 1992.