Product Datasheet

Amine Reactive Comp-Bead 2 Population Kit NBP3-00496

Unit Size: 10 mL (2 x 5 mL)

Store at 4C in the dark. Do not freeze.

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NBP3-00496

Amine Reactive Comp-Bead 2 Population Kit

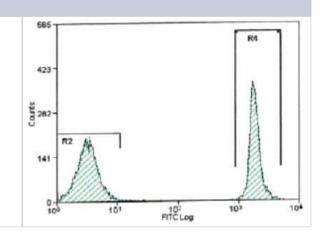
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H 7.4)

Buffer	0.016 M PBS (pH 7.4)
Product Description	
Description	 The Amine Reactive Compensation Beads are designed for labeling amine-reactive dyes (live/dead fixable viability stains) when setting compensation in flow cytometry. Ensure consistent and accurate compensation in any channel Evaluate cell viability of animal cells, bacteria, yeast, and fungi Contains both amine reactive and nonreactive beads for generating positive and negative populations Concentration: 10^7 particles / mL Particle size: 7.0 -7.9 micron Particle material: polystyrene
Kit Components	High Binding Beads - 5 mL, Negative Binding Beads - 5 mL

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Product Application Details	
Applications	Flow Cytometry
Recommended Dilutions	Flow Cytometry
Application Notes	Dye and bead concentrations may be further optimized for best results. Centrifugation force and time may need to be increased if needed. Protect the beads from light after exposure to the dye. Use immediately after staining.

Images

Amine Reactive Comp-Bead 2 Population [NBP3-00496] - Histogram. Amine Reactive Comp-Beads stained with Biolegend Zombie Green (Tm).



Procedures

Flow Cytometry Protocol for Amine Reactive Comp-Bead 2 Population Kit (NBP3-00496)

- 1. Allow beads to come to room temperature, then vortex briefly.
- 2. Add 1 drop (about 50 uL) of the High Binding beads to a 1.5 mL microcentrifuge tube.
- 3. Wash the beads by adding 0.5 mL of PBS (free of surfactant and blocker) to the microcentrifuge tube, centrifuge at 300 x G for 5 minutes, decant and repeat.
- 4. Decant and resuspend in 50 uL PBS.
- 5. Prepare the amine reactive dye according to the manufacturer's instructions.
- 6. Add 1 4 uL of the amine reactive dye to the bead suspension and vortex briefly.
- 7. Incubate for 30 minutes. Protect tube from light.
- 8. Add 1 mL of PBS to the same tube and vortex briefly.
- 9. Centrifuge at 300 x G for 5 minutes, decant and repeat.
- 10. Resuspend the beads in PBS containing 0.05% BSA with brief vortex.
- 11. Add a drop (about 50 uL) fo the Negative Binding beads to the labeled High Binding beads.
- 12. Analyze on the flow cytometer using a live gate around the singlet population in the FSC/SSC dot plot.
- 13. Create a fluorescent histogram for the appropriate detectors and perform compensation to achieve the desired results.





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