

Product Datasheet

BNIP3L Antibody - BSA Free

NBP1-88558

Unit Size: 0.1 ml

Store at 4C short term. Aliquot and store at -20C long term. Avoid freeze-thaw cycles.

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NBP1-88558

BNIP3L Antibody - BSA Free

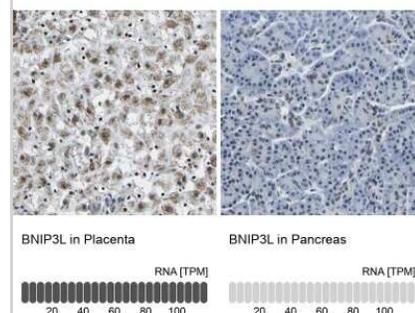
| Product Information | |
|---------------------|--|
| Unit Size | 0.1 ml |
| Concentration | Concentrations vary lot to lot. See vial label for concentration. If unlisted please contact technical services. |
| Storage | Store at 4C short term. Aliquot and store at -20C long term. Avoid freeze-thaw cycles. |
| Clonality | Polyclonal |
| Preservative | 0.02% Sodium Azide |
| Isotype | IgG |
| Purity | Immunogen affinity purified |
| Buffer | PBS (pH 7.2) and 40% Glycerol |

| Product Description | |
|---------------------|--|
| Host | Rabbit |
| Gene ID | 665 |
| Gene Symbol | BNIP3L |
| Species | Human, Mouse |
| Reactivity Notes | Mouse reactivity reported in the scientific literature (PMID: 24573672). |
| Immunogen | This antibody was developed against Recombinant Protein corresponding to amino acids: SNGNDNGNGKNGGLEHVPSSSSIHNGDMEKILLDAQHESGQSSSRGSSSHCDS PSPQEDGQIMFDVEMHTSRDHSSQSEEEVVEGEKEVEALKKSADWVSDWSS RPENIPPKEFHFRHPKRSVLSMRKSGAMK |

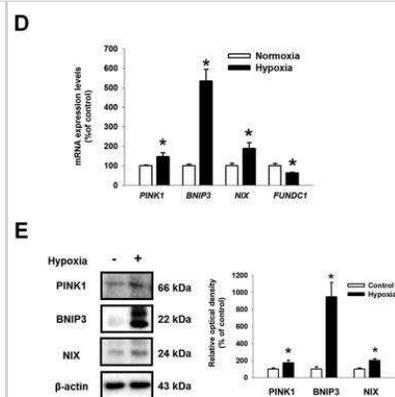
| Product Application Details | |
|-----------------------------|---|
| Applications | Western Blot, Immunocytochemistry/ Immunofluorescence, Immunohistochemistry, Immunohistochemistry-Paraffin |
| Recommended Dilutions | Western Blot Reported in scientific literature (PMID:33473105), Immunohistochemistry 1:200 - 1:500, Immunocytochemistry/ Immunofluorescence 0.25-2 ug/ml, Immunohistochemistry-Paraffin 1:200 - 1:500 |
| Application Notes | For IHC-Paraffin, HIER pH 6 retrieval is recommended. ICC/IF Fixation Permeabilization: Use PFA/Triton X-100. |

Images

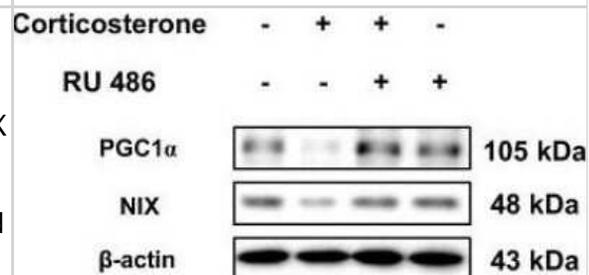
Immunohistochemistry-Paraffin: BNIP3L Antibody [NBP1-88558] - Staining in human placenta and pancreas tissues using anti-BNIP3L antibody. Corresponding BNIP3L RNA-seq data are presented for the same tissues.



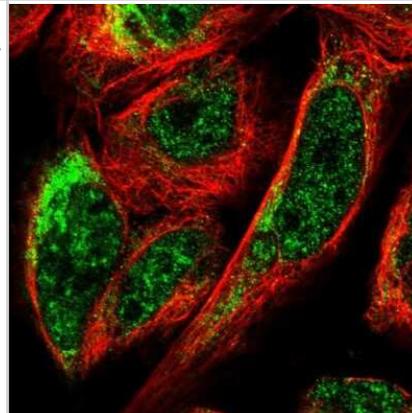
Western Blot: BNIP3L Antibody [NBP1-88558] - UCB-hMSCs were exposed to normoxia or hypoxia. (D) The mRNA expressions of PINK1, BNIP3, NIX and FUNDC1 were analyzed by quantitative real-time PCR (qPCR). n = 5. (E) The protein expressions of PINK1, BNIP3, NIX and β -actin were assessed by western blot. n = 4. *p < 0.05 versus control. Image collected and cropped by CiteAb from the following publication (<https://linkinghub.elsevier.com/retrieve/pii/S2213231717303804>) licensed under a CC-BY license.



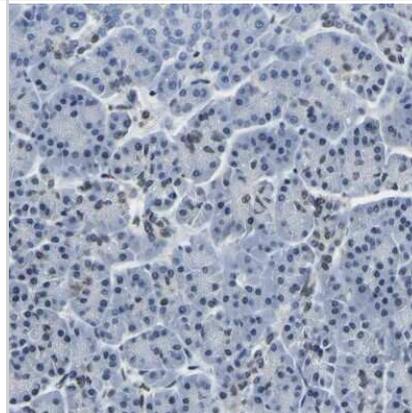
Western Blot: BNIP3L Antibody [NBP1-88558] - Vehicle or RU 486 (5 mg/kg) injected mice were presented with/without corticosterone (10 mg/kg) for 3 days. The expressions of peroxisome proliferator-activated receptor gamma coactivator 1-alpha (PGC1 alpha NBP1-04676) and NIX (BNIP3L NBP1-88558) were visualized via western blotting. Loading control is beta-actin. n = 5. Image collected and cropped by CiteAb from the following publication (<https://pubmed.ncbi.nlm.nih.gov/33473105/>) licensed under a CC-BY license.



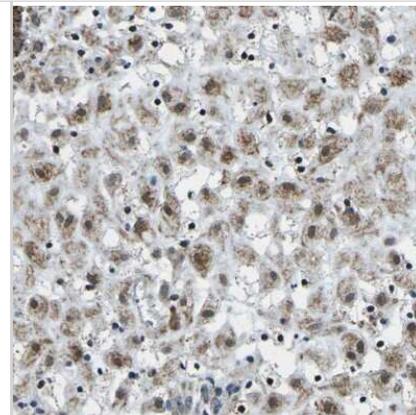
Immunocytochemistry/Immunofluorescence: BNIP3L Antibody [NBP1-88558] - Staining of human cell line U-2 OS shows localization to nuclear speckles & mitochondria. Antibody staining is shown in green.



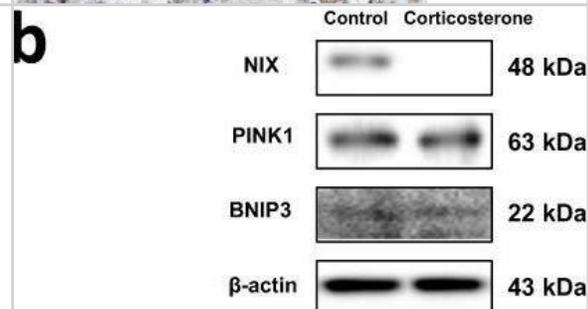
Immunohistochemistry-Paraffin: BNIP3L Antibody [NBP1-88558] - Staining of human pancreas shows low expression as expected.



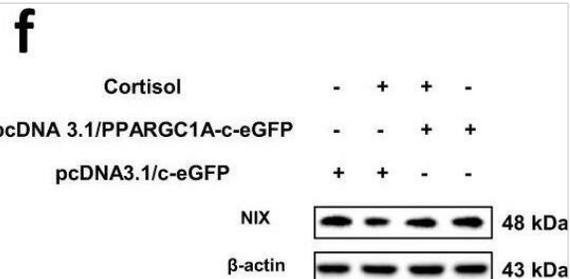
Immunohistochemistry-Paraffin: BNIP3L Antibody [NBP1-88558] - Staining of human placenta shows high expression.



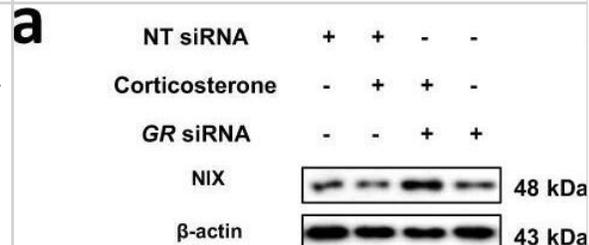
Corticosterone affects NIX-dependent mitophagy through decreasing PGC1 α in vivo. a–f Mice exposed to vehicle, corticosterone (10 mg/kg), corticosterone with phorbol 12-myristate 13-acetate (PMA pretreatment, 200 μ g/kg), or PMA alone for 7 days. a Slide samples for IHC immunostained with LAMP1 (green), TOMM20 (red), & DAPI (blue). Scale bars, 100 μ m (magnification, \times 200). n = 5. b The expressions of NIX, PTEN-induced kinase 1 (PINK1), & BCL2 interacting protein 3 (BNIP3) detected with WBt where β -actin used as a loading control. n = 5. c Slide samples for IHC immunostained with synaptophysin (green), PSD95 (red), & DAPI (blue). Scale bars, 100 μ m (magnification, \times 200). n = 5. d Synaptophysin & PSD95 detected by WBt. Loading control is β -actin. n = 5. e The mice subjected to Y-maze test to evaluate spatial memory function. n = 6. f The mice subjected to forced swim test to evaluate depression-like behavior. n = 5. g Vehicle or RU 486 (5 mg/kg) injected mice presented with/without corticosterone (10 mg/kg) for 3 days. The expressions of peroxisome proliferator-activated receptor gamma coactivator 1-alpha (PGC1 α) & NIX visualized via WB. Loading control is β -actin. n = 5. h The schematic model for mechanisms of inhibition in NIX-dependent mitophagy by glucocorticoid was shown. All blots & IF images representative. n = 5 or 6 from each animal with two technical replicates each in results of IHC & WBt. Quantitative data presented as a mean \pm S.E.M. The representative images acquired by SRRF imaging system. Two-sided two-way ANOVA was conducted except Fig. 8b, data of which analyzed by two-sided unpaired student's t test. ** indicates $p < 0.01$ versus control & ## indicates $p < 0.01$ versus corticosterone, respectively. Data provided as a Source data file. Image collected & cropped by CiteAb from the following publication (<https://pubmed.ncbi.nlm.nih.gov/33473105>), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



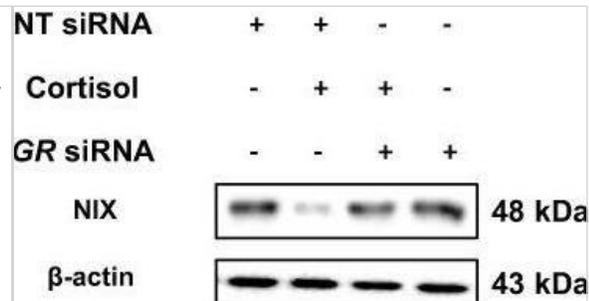
Western Blot: BNIP3L Antibody [NBP1-88558] - Role of PGC1 α in NIX-dependent mitophagy. a–e Nontargeting (NT) or GR siRNA was transfected to hippocampal neurons & SH-SY5Y cells for 24 h prior to corticosterone & cortisol for 12 h, respectively. a, b Peroxisome proliferator-activated receptor gamma coactivator 1-alpha (PGC1 α) expression was detected in western blot where β -actin was used as a loading control in both cell types. n = 5. c Colocalization of PGC1 α (red) & DAPI (blue) in hippocampal neurons was visualized with SRRF imaging system. Scale bars, 20 μ m (magnification, \times 1000). n = 5. d Colocalization of PGC1 α (green) & DAPI (blue) in SH-SY5Y was visualized with SRRF imaging system. Scale bars, 20 μ m (magnification, \times 1000). n = 5. e PGC1 α protein expressions in subcellular fraction samples were detected by western blotting. Lamin A/C & α -tubulin were used as a nuclear & cytosolic loading control, respectively. n = 5. f, g SH-SY5Y cells were transfected with pcDNA3.1/c-eGFP or pcDNA3.1/PPARGC1A-c-eGFP vector for 24 h prior to cortisol treatment for 24 h. f NIX expression was detected in western blot where β -actin was used as a loading control. n = 5. g TOMM20 levels were detected by western blot. Loading control for western blot is β -actin. n = 5. All blots & immunofluorescence images are representative. n = 5 from independent experiments with two technical replicates each. Quantitative data are presented as a mean \pm S.E.M. Two-sided two-way ANOVA was conducted. ** indicates $p < 0.01$ versus control. #, ## indicates $p < 0.05$, $p < 0.01$ versus corticosterone in hippocampal neurons & cortisol in SH-SY5Y, respectively. Data are provided as a Source data file. Image collected & cropped by CiteAb from the following publication (<https://pubmed.ncbi.nlm.nih.gov/33473105>), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



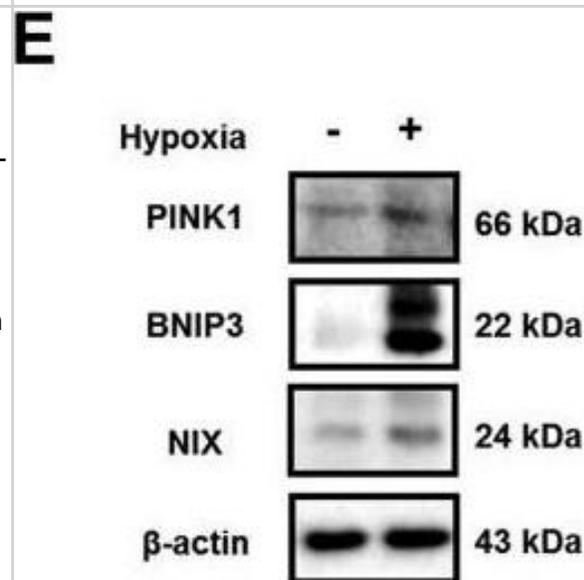
Western Blot: BNIP3L Antibody [NBP1-88558] - GR-dependent downregulation of NIX expression via PGC1 α . a, b Nontargeting (NT) or GR siRNA was transfected to hippocampal neurons & SH-SY5Y cells for 24 h prior to corticosterone & cortisol for 24 h, respectively. NIX expression was detected in western blot where β -actin was used as a loading control. n = 5. c SH-SY5Y cells were incubated with cortisol for 6 h. The mRNA expression levels of genes associated with nuclear receptors & coregulators were assessed by RT2 Profiler PCR array. Heat maps with hierarchical clustering were acquired by using the GeneGlobe Data analysis Center on Qiagen website. n = 3. d A thousand base pair upstream of the first codon of the PPARGC1A was described & the putative GRE binding sequence was emphasized with yellow labeling. e SH-SY5Y cells were incubated with cortisol for 6 h. DNA was immunoprecipitated with IgG, RNA polymerase (RNAPol), & glucocorticoid receptor (GR) antibody. The immunoprecipitation & input samples were amplified with primers of GAPDH & PPARGC1A gene. n = 5. All blots are representative. n = 3 or 5 from independent experiments with two technical replicates each, respectively. Quantitative data are presented as a mean \pm S.E.M. Two-sided two-way ANOVA was conducted in Fig. 6a, b. Two-sided unpaired student's t test was conducted in Fig. 6e. ** indicates $p < 0.01$ versus control. ## indicates $p < 0.01$ versus corticosterone in hippocampal neurons & cortisol in SH-SY5Y, respectively. Data are provided as a Source data file. Image collected & cropped by CiteAb from the following publication (<https://pubmed.ncbi.nlm.nih.gov/33473105>), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



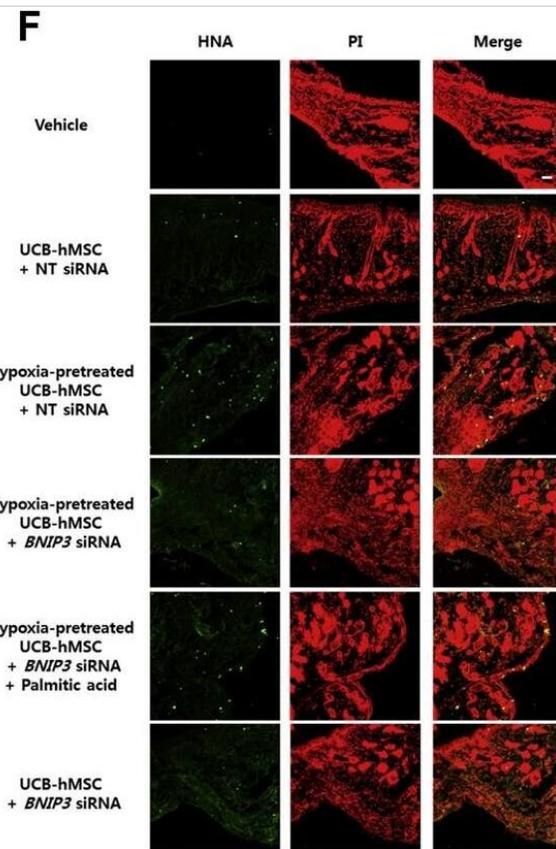
Western Blot: BNIP3L Antibody [NBP1-88558] - GR-dependent downregulation of NIX expression via PGC1 α . a, b Nontargeting (NT) or GR siRNA was transfected to hippocampal neurons & SH-SY5Y cells for 24 h prior to corticosterone & cortisol for 24 h, respectively. NIX expression was detected in western blot where β -actin was used as a loading control. n = 5. c SH-SY5Y cells were incubated with cortisol for 6 h. The mRNA expression levels of genes associated with nuclear receptors & coregulators were assessed by RT2 Profiler PCR array. Heat maps with hierarchical clustering were acquired by using the GeneGlobe Data analysis Center on Qiagen website. n = 3. d A thousand base pair upstream of the first codon of the PPARGC1A was described & the putative GRE binding sequence was emphasized with yellow labeling. e SH-SY5Y cells were incubated with cortisol for 6 h. DNA was immunoprecipitated with IgG, RNA polymerase (RNAPol), & glucocorticoid receptor (GR) antibody. The immunoprecipitation & input samples were amplified with primers of GAPDH & PPARGC1A gene. n = 5. All blots are representative. n = 3 or 5 from independent experiments with two technical replicates each, respectively. Quantitative data are presented as a mean \pm S.E.M. Two-sided two-way ANOVA was conducted in Fig. 6a, b. Two-sided unpaired student's t test was conducted in Fig. 6e. ** indicates $p < 0.01$ versus control. ### indicates $p < 0.01$ versus corticosterone in hippocampal neurons & cortisol in SH-SY5Y, respectively. Data are provided as a Source data file. Image collected & cropped by CiteAb from the following publication (<https://pubmed.ncbi.nlm.nih.gov/33473105>), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



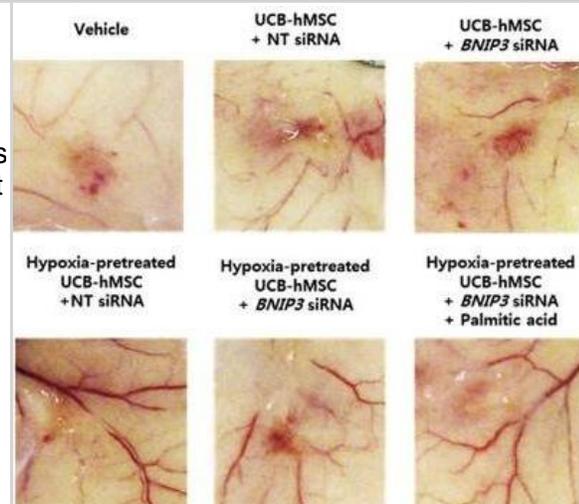
Western Blot: BNIP3L Antibody [NBP1-88558] - Effects of hypoxia on mitophagy regulator expressions & mitophagy in UCB-hMSCs. (A) UCB-hMSCs incubated w/ various times of hypoxia (0–48 h). Cells stained w/ MitotrackerTM. n = 6 (magnification, $\times 1,200$). Scale bars, 50 μ m. (B) The expressions of COX4 & β -actin detected by western blot. n = 4. (C) UCB-hMSCs exposed to 24 h of normoxia or hypoxia. Cells immuno-stained w/ COX4 & LC3B-specific antibodies (magnification, $\times 600$). Scale bars, 37.5 μ m. (D) The mRNA expressions of PINK1, BNIP3, NIX & FUNDC1 analyzed by quantitative real-time PCR (qPCR). n = 5. (E) The protein expressions of PINK1, BNIP3, NIX & β -actin assessed by western blot. n = 4. (F, G) siRNAs of PINK1, BNIP3, NIX or non-targeting (NT) transfected to UCB-hMSCs prior to hypoxia treatment for 24 h. COX4 & β -actin expressions assessed by western blot. n = 4 (F). Cells immunostained w/ COX4 & PI. n = 3 (magnification, $\times 400$). All scale bars, 50 μ m. COX4 fluorescence intensity analyzed by luminometer. n = 5 (G). (H) BNIP3 siRNA transfected to UCB-hMSCs prior to hypoxia treatment for 24 h. Cells stained w/ MitotrackerTM. n = 6 (magnification, $\times 1,200$). Scale bars, 50 μ m. (I) BNIP3, β -tubulin & COX4 w/ cytosol & mitochondrial fractionized samples detected by western blot. (J) Cells incubated w/ hypoxia or normoxia for 24 h. Cells immuno-stained w/ BNIP3 & LC3B-specific antibodies. (magnification, $\times 600$). Scale bars, 37.5 μ m. Western blot data normalized by β -actin, & qPCR data normalized by ACTB mRNA expression level. Quantitative data are presented as a mean \pm S.E.M. All blot & confocal images are representative. * $p < 0.05$ versus control, # $p < 0.05$ versus hypoxia. Image collected & cropped by CiteAb from the following publication (<https://linkinghub.elsevier.com/retrieve/pii/S2213231717303804>), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



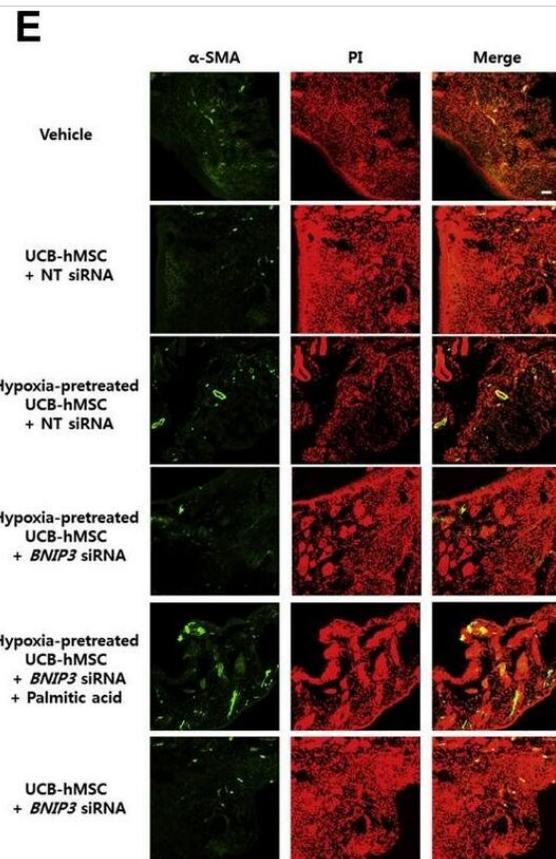
Immunocytochemistry/ Immunofluorescence: BNIP3L Antibody [NBP1-88558] - The role of PA in BNIP3 silenced UCB-hMSC survival in the mouse skin wound healing model. (A) Mouse skin wound surgery with UCB-hMSC transplantation was performed as described in Section 2. Representative gross wound images were acquired at post injection days 0, 4, 8, 12. Skin wound sizes at day 8 were compared with wound size at day 0. $n = 5$. (B) Tissue slide samples were stained with hematoxylin & eosin. Low & high magnified histological gross images are shown in the left & right panels, respectively. Scale bars, 260 μm (magnification, $\times 40$) & 100 μm (magnification, $\times 100$). $n = 5$. (C) Representative images of blood vessels in skin wounds on day 12 (top panel). Vessel density was analyzed by using ImageJ program (bottom panel). $n = 5$. (D-F) Histological tissue samples were immuno-stained with CD31, α -SMA, & HNA-specific antibodies & PI for counterstaining. α -SMA & HNA-positive cells were visualized by confocal microscopy. The number of CD31 & α -SMA-positive cells in high power field (HPF), & the percentage of HNA-positive cells in total cells were analyzed by using Metamorph software. Scale bars, 100 μm (magnification, $\times 100$). $n = 5$. Data are presented as a mean \pm S.E.M. $\$p < 0.05$ versus vehicle group, $*p < 0.05$ versus UCB-hMSC group given NT siRNA, $\#p < 0.05$ versus UCB-hMSC group given NT siRNA with hypoxia pretreatment, $@p < 0.05$ versus UCB-hMSC group given BNIP3 siRNA with hypoxia pretreatment. Image collected & cropped by CiteAb from the following publication (<https://linkinghub.elsevier.com/retrieve/pii/S2213231717303804>), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



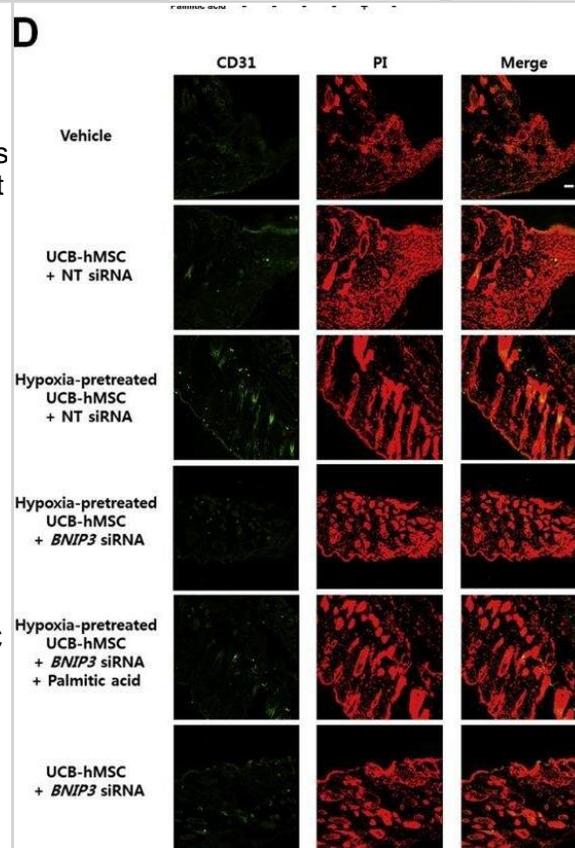
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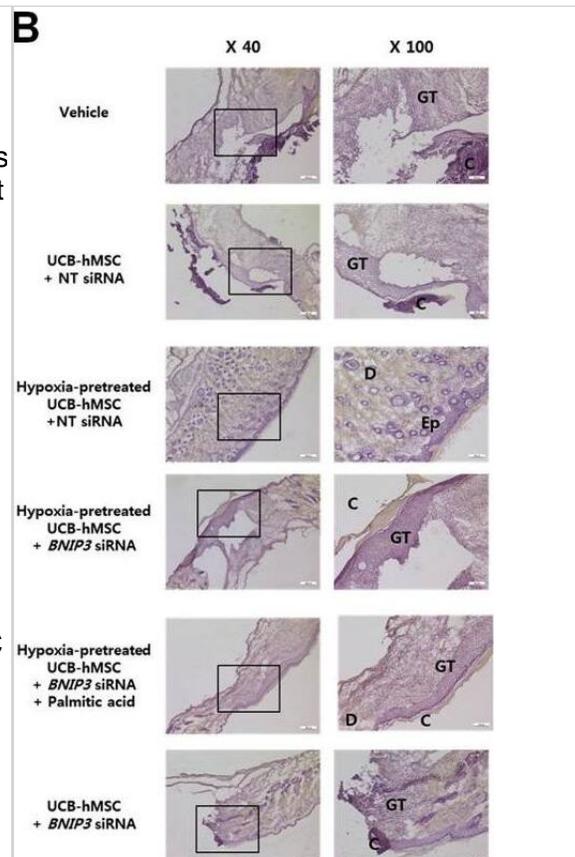
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Publications

Kim MJ, Choi GE, Chae CW et al. Melatonin-mediated FKBP4 downregulation protects against stress-induced neuronal mitochondria dysfunctions by blocking nuclear translocation of GR Cell death & disease 2023-02-21 [PMID: 36810730] (WB, Human)

Choi GE, Lee HJ, Chae CW, et al. BNIP3L/NIX-mediated mitophagy protects against glucocorticoid-induced synapse defects Nature communications 2021-01-20 [PMID: 33473105] (WB, Mouse)

Lee H, Jung Y, Choi G et al. BNIP3 induction by hypoxia stimulates FASN-dependent free fatty acid production enhancing therapeutic potential of umbilical cord blood-derived human mesenchymal stem cells Redox Biol 2017-07-04 [PMID: 28704726] (WB, Human)

Li W, Zhang X, Zhuang H et al. MicroRNA-137 Is a Novel Hypoxia-responsive MicroRNA That Inhibits Mitophagy via Regulation of Two Mitophagy Receptors FUNDC1 and NIX. J Biol Chem 2014-04-11 [PMID: 24573672] (ICC/IF, Mouse)

Stadler C, Rexhepaj E, Singan VR et al. Immunofluorescence and fluorescent-protein tagging show high correlation for protein localization in mammalian cells. Nat Methods 2013-04-01 [PMID: 23435261]



Novus Biologicals USA

10730 E. Briarwood Avenue
Centennial, CO 80112
USA
Phone: 303.730.1950
Toll Free: 1.888.506.6887
Fax: 303.730.1966
nb-customerservice@bio-techne.com

Bio-Techne Canada

21 Canmotor Ave
Toronto, ON M8Z 4E6
Canada
Phone: 905.827.6400
Toll Free: 855.668.8722
Fax: 905.827.6402
canada.inquires@bio-techne.com

Bio-Techne Ltd

19 Barton Lane
Abingdon Science Park
Abingdon, OX14 3NB, United Kingdom
Phone: (44) (0) 1235 529449
Free Phone: 0800 37 34 15
Fax: (44) (0) 1235 533420
info.EMEA@bio-techne.com

General Contact Information

www.novusbio.com
Technical Support: nb-technical@bio-techne.com
Orders: nb-customerservice@bio-techne.com
General: novus@novusbio.com

Products Related to NBP1-88558

| | |
|---------------|---|
| NBP1-88558PEP | BNIP3L Recombinant Protein Antigen |
| HAF008 | Goat anti-Rabbit IgG Secondary Antibody [HRP] |
| NB7160 | Goat anti-Rabbit IgG (H+L) Secondary Antibody [HRP] |
| NBP2-24891 | Rabbit IgG Isotype Control |

Limitations

This product is for research use only and is not approved for use in humans or in clinical diagnosis. Primary Antibodies are guaranteed for 1 year from date of receipt.

For more information on our 100% guarantee, please visit www.novusbio.com/guarantee

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