Product Datasheet

beta-Actin Antibody (AC-15) NB600-501

Unit Size: 0.1 ml

Store at 4C short term. Aliquot and store at -20C long term. Avoid freeze-thaw cycles.



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NB600-501

beta-Actin Antibody (AC-15)

Product Information		
Unit Size	0.1 ml	
Concentration	This product is unpurified. The exact concentration of antibody is not quantifiable.	
Storage	Store at 4C short term. Aliquot and store at -20C long term. Avoid freeze-thaw cycles.	
Clonality	Monoclonal	
Clone	AC-15	
Preservative	0.9% Sodium Azide	
Isotype	IgG1	
Purity	Unpurified	
Buffer	Ascites.	
Target Molecular Weight	42 kDa	
Product Description		
Host	Mouse	
Gene ID	60	
Gene Symbol	ACTB	
Species	Human, Mouse, Rat, Porcine, Bovine, Canine, Chicken, Feline, Fish, Guinea Pig, Hamster, Leech, Mammal, Primate, Rabbit, Sheep, Squirrel, Turkey, Zebrafish, Drosophila (Negative)	
Reactivity Notes	The antibody cross reacts with beta-actin expressing cells in carp, leech tissues (Hirudo medicinalis), ground squirrel. Does not cross react with adult cardiac, skeletal muscle, drosophila or amoeba beta actin. Mammal reactivity and Hamster reactivity reported in scientific literature (PMID: 25130694 and 24478435 respectively). Please note that this antibody is reactive to Mouse and derived from the same host, Mouse. Additional Mouse on Mouse blocking steps may be required for IHC and ICC experiments. Please contact Technical Support for more information. Human reactivity reported in scientific literature (PMID:32938681) Use in Turkey reported in scientific literature (PMID:27816932).	
Specificity/Sensitivity	In staining of chicken gizzard ultrathin tissue cryosections, the antibody labels the dense bodies and longitudinal channels linking consecutive dense bodies that are also occupied by desmin and the membrane-associated dense plaque. It does not stain adult cardiac and skeletal muscles except for traces due to contaminations of the sample with non-muscle cells, or if embryonic tissue is being used. The epitope recognized by the antibody is resistant to formalin-fixing and paraffin-embedding.	
Immunogen	This beta-Actin Antibody (AC-15) was made to a slightly modified Beta- cytoplasmic actin N-terminal peptide, Ac-Asp-Asp-Asp-Ile-Ala-Ala-Leu-Val-Ile- Asp-Asn-Gly-Ser-Gly-Lys, conjugated to KLH.	
Product Application Details		
Applications	Western Blot, Simple Western, ELISA, Flow Cytometry, Immunocytochemistry/ Immunofluorescence, Immunohistochemistry, Immunohistochemistry-Frozen, Immunohistochemistry-Paraffin, Immunoprecipitation, Chromatin Immunoprecipitation (ChIP), Knockdown Validated, Knockout Validated	



Recommended Dilutions	Western Blot 1:5000 - 1:10000, Simple W 1:100 - 1:2000, Immunohistochemistry 1: Immunofluorescence 1:1000 - 1:2000, Im Immunohistochemistry-Paraffin 1:10 - 1:5 1:500, Chromatin Immunoprecipitation (C Validated	Vestern 1:25, Flow Cytometry, ELISA 10 - 1:500, Immunocytochemistry/ mmunoprecipitation 1:10 - 1:500, 500, Immunohistochemistry-Frozen ChIP), Knockout Validated, Knockdown
Application Notes	See <u>Simple Western Antibody Database</u> for Simple Western validation: tested in HeLa lysate (0.1 mg/ml); antibody dilution of 1:25; separated by size; detects a band at 54 kDa	
Images		
Western Blot: beta-Actin Antibody (AC-15) [NB600-501] - MCDK cells		Increasing Gene Induction







Immunocytochemistry/Immunofluorescence: beta-Actin Antibody (AC-15) [NB600-501] - Analysis of beta-Actin antibody on mouse fibroblasts. Image from verified customer review.





Western Blot: beta-Actin Antibody (AC-15) [NB600-501] - Analysis of beta-Actin in bovine whole skeletal muscle lysate. Image from verified customer review.

Immunohistochemistry: beta-Actin Antibody (AC-15) [NB600-501] -

1:1000 Monoclonal Anti-beta-Actin Clone: AC-15.

Western Blot: beta-Actin Antibody (AC-15) [NB600-501] - Whole cell extract of human fibroblasts was separated on SDS-PAGE and blotted with Monoclonal Anti-beta-Actin. The antibody was developed with Goat Anti-Mouse IgG, Peroxidase conjugate and AEC substrate. Lanes A: Antibody dilution 1:5,000 B: Negative control (only secondary antibody).





















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Vps34 activity is required for maintaining LC3B-ii levels at P10 & mTOR D inhibition increases p-Beclin-1 & LC3B-ii levels at P28. (A) Representative Western blot images for actin, DARPP32 & LC3B-i & -ii in striatal slices obtained from P10 or P28 mice, treated with SAR405 (1 μM) or vehicle (Veh; DMSO, 0.1%). (B,C) Quantification of LC3B-ii relative to actin, normalized to vehicle condition at each age. (B) P10: unpaired, two-tailed t-test, t11 = 2.985, p = 0.0124; P28: unpaired, twotailed t-test, t24 = 1.807, p = 0.0922. P10: Veh: n = 6 slices, SAR405 n = 7 slices from 3 to 4 mice/age. (C) P28: Veh: n = 9 slices, Baf n = 7 slices from 3 to 4 mice/age. *p < 0.05. (D) Representative Western blot images for actin, DARPP32, p-Beclin-1 Ser14 & LC3B-i & -ii in striatal slices from P28 mice, treated with Torin-1 (5 μ M) or vehicle (Veh; DMSO, 0.1%). Quantification of (E) LC3B-ii & (F) p-Beclin S14 relative to actin, normalized to vehicle. Data analyzed with unpaired, two-tailed t-test; (E) LC3B-ii/actin, t24 = 2.853, p = 0.0088. Veh: n = 19 slices, Baf n = 7 slices from 5 to 7 mice. *p < 0.05. (F) p-Beclin S14/actin, t22 = 3.337, p = 0.0030. **p < 0.01, Veh: n = 18 slices, Torin-1 n = 6 slices from 3 to 5 mice. Image collected & cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/32296308), licensed under a CC-BY license. Not internally tested by Novus Biologicals. The hypoxia-activated pro-drug Tirapazamine (TPZ) reduces stem-like е Aldefluorbright population in vitro & in liver metastases(a) Human colonospheres cultured in hypoxia (0.1%) & normoxia (21%) for 24 hrs in absence or presence of TPZ for 4 hrs. Cells processed for IF analysis of yH2AX (green). DAPI (blue) used to visualize cell nuclei. (b) Bar graphs showing quantification of vH2AX foci observed in (a), shown as % of mean yH2AX intensity at normoxia (n=3). (c) Representative confocal pictures of comet assay of colonospheres cultured as in (a). (d) Bar graphs showing quantification of DNA damage observed with the comet assay (Extend tail moment) (n=77). (e) Experiment was performed as in (a), but cells were either treated or untreated for 16 hrs. Cells were lysed & analyzed by WB for the indicated markers (cropped). (f) Colonospheres cultured in hypoxia & treated with TPZ for 16 hrs. The cells were FACS sorted into Aldefluorbright & Aldefluordim populations & analyzed by WB for the indicated markers. (g) Human colonospheres cultured in hypoxia (0.1%) for 24 hrs in absence or presence of TPZ for 4 hrs & FACS sorted into Aldefluorbright & Aldefluordim populations. Bar graph shows the fold change Aldefluordim & Aldefluorbright cells upon TPZ treatment (n=3). (h) C26 tumor cells were injected into the liver parenchyma of immune-deficient mice. Following tumor initiation, tumorbearing liver lobes were subjected to a vascular clamping protocol [24] to induce hypoxia & treated with saline (control) or TPZ (treatment) for 10 days. At endpoint, livers were excised & expression of ALDH1 was examined by IHC. Magnification 20x, T=tumor. (i) Bar graphs showing quantification of ALDH1 (n=28) observed in (h) (n=14). * = significant (p>0.05). ns = not significant. Image collected & cropped by CiteAb from the following publication (https://www.oncotarget.com/lookup/doi/10.18632/oncotarget.21145), licensed under a CC-BY license. Not internally tested by Novus

P28 Veh Torin LC3B-i LC3B-ii p-Beclin Ser14 Actin DARPP32 Normoxia Hypoxia TP2 Control Control TPZ yH2aX RIF1 RAD51 53BP1 KU70 Cleaved Caspase 3 **B**-actin

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Western Blot: beta-Actin Antibody (AC-15) [NB600-501] - Effect of IFN-y 🗃 pretreatment on carfilzomib sensitivity of H727 (a) & H23 (b) cells. IFN-y (150 U□ml-1) pretreatment for 24 h led to increased expression (left panel) & activity (middle panel) of proteasome immuno-subunits, & sensitized H727 cells to Cfz, while desensitizing H23 cells towards Cfz. The IC50 values displayed a statistically significant difference between IFN-y-pretreated cells & vehicle control (P value < 0.01, n = 3, Student's t-test comparing the log transformed IC50 values obtained from three independent runs). Image collected & cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/30858500), licensed under a CC-BY license. Not internally tested by Novus Biologicals.

Western Blot: beta-Actin Antibody (AC-15) [NB600-501] - RARA overexpression increases chemosensitivity to AM80(A, B) Overexpression of either RARAwt or RARAR394Q increases chemosensitivity of (A) HuT78 & (B) Karpas 299 cells to AM80. The presence of the R394Q mutation does not reduce chemosensitivity significantly. (C) RARAwt overexpression increases retinoic acid response element (RARE) activity (indicated by luciferase) in HuT78 cells treated with AM80. (D) RARAwt overexpression increases the degree of CDK2, CDK4 & CDK6 inhibition caused by AM80. wt, wildtype; CDK, cyclin-dependent kinase. Image collected & cropped by CiteAb from the following publication

(https://www.oncotarget.com/lookup/doi/10.18632/oncotarget.15441), licensed under a CC-BY license. Not internally tested by Novus Biologicals.

Western Blot: beta-Actin Antibody (AC-15) [NB600-501] - RARA drives expression of cyclin-dependent kinases(A) Knockdown of RARA causes G1 cell cycle arrest (P = 0.004) in Mac-1 cells. (B) Expression of the regulators of cell cycle progression, CDK6, CDK4, & to a lesser extent, CDK2, is inhibited by RARA knockdown in Mac-1 cells. (C) Expression of CDK6, CDK4, & CDK2 is increased following overexpression of RARAwt & RARAR394Q in HuT78 cells. RARA, retinoic acid receptor alpha; wt, wild-type; siRNA, small interfering RNA; CDK, cyclin-dependent kinase. Image collected & cropped by CiteAb from the following publication (https://www.oncotarget.com/lookup/doi/10.18632/oncotarget.15441), licensed under a CC-BY license. Not internally tested by Novus Biologicals.

Western Blot: beta-Actin Antibody (AC-15) [NB600-501] - (A) Protein expression level of CCL2 with WARS silencing under control & IFNy stimulation conditions, performed on human primary PBMC-derived macrophages. *p<0.05, Student's t-test(B) Western blot analysis showing the protein levels of phosphorylation of STAT1 with WARS silencing under control & IFNy stimulation conditions, performed on human primary PBMC-derived macrophages. (C) Relative mRNA & protein expression levels for CCL2, TNFa, STAT1, JAK2 & CD36 with GBP1 silencing under control & IFNy stimulation conditions, performed on THP-1-differentiated macrophage-like cells. (D) Western blot analysis showing the protein levels of JAK2 & phospho-STAT1 with GBP1 silencing in THP-1 macrophage-like cells at 3 & 6 hr after IFNy stimulation. Image collected & cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/30303482), licensed under a CC-BY license. Not internally tested by Novus Biologicals.









Western Blot: beta-Actin Antibody (AC-15) [NB600-501] - 16:4(n-3) stimulation increases cPLA2 activity in splenic macrophages. A) Schematic overview of hydrolysis sites of PC to yield metabolites, such as LPC. Splenic macrophages were harvested from murine BALB/c spleens by magnetic bead sorting using F4/80 Abs. Macrophages were incubated with either vehicle or 16:4(n-3) (25 nM) for indicated times. B, C) Fluorescent-based analysis of PLA1 & cPLA2 enzymatic activity revealed a decrease in PLA1 activity (B) & a significant time-dependent increase in cPLA2 activity (C) upon stimulation with 16:4(n-3). Preincubation of splenic macrophages with the GPR120-specific antagonist AH-7614 did not affect 16:4(n-3)-mediated changes in PLA1 activity but was able to prevent the 16:4(n-3)-induced increase in cPLA2 activity. D) In addition, splenic macrophage cell lysates were analyzed by Western blot for phospho-cPLA2. Actin was used as a loading control. E) Treatment of tumor-bearing BALB/c mice with cisplatin (cis) & sCM derived from splenocytes that were incubated with 16:4(n-3) & AACOCF3 did not induce chemoresistance, whereas cotreatment with cisplatin & sCM+ 16:4(n-3) did. Panel B shows grouped results of 2 independent experiments with similar outcomes. Panels C & D show results of 4 independent experiments. Panel E shows grouped results of 2 independent experiments with similar outcomes (n = 8 per group). Data are presented as means ± sem. Statistical significance was determined by 2-tailed Student's t test (B, C) or 1-way ANOVA (E); all compared with vehicle control (B, C) or with cisplatin alone (E) unless indicated otherwise. *P < 0.05; **P < 0.01. Image collected & cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/28183801), licensed under a CC-BY

license. Not internally tested by Novus Biologicals. Western Blot: beta-Actin Antibody (AC-15) [NB600-501] - WEGL reduces 9 serum LPS & TLR4-related signalling pathways in HFD mice. Effects of WEGL treatment on serum endotoxin (a) TLR4 protein production (b,c) JNK phosphorylation (d,e) & $I\kappa B - \alpha$ production (f,q) were examined in the liver & epididymal adipose tissues of chow- & HFD-fed mice as described in Fig. 1. Serum endotoxin (EU ml-1) was determined as mean±s.e.m. using the limulus amebocyte lysate assay kit. Representative immunoblots for target proteins in b-g are shown. Molecular weight markers were indicated as kilodaltons (kDa). Protein levels were normalized to internal controls (β-actin or total JNK, T-JNK) & the relative ratio to the Chow group was labelled on the top of immunoblots. Graph bars in a with different letters on top represent statistically significant results (P<0.05) based on Newman–Keuls post hoc one-way ANOVA analysis, whereas bars labelled with the same letter correspond to results that show no statistically significant differences. Where two letters are present on top of the bar, each letter should be compared separately with the letters of other bars to determine whether the results show statistically significant differences. Image collected & cropped by CiteAb from the following publication (https://www.nature.com/articles/ncomms8489), licensed under a CC-BY



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Western Blot: beta-Actin Antibody (AC-15) [NB600-501] - Liver injury induces intra-nuclear metabolic changes & histone hyper-acetylation. Nuclear (A) acetyl-CoA, (B) lactate, & (C) nHDAC activity in livers harvested 6 h after the injection of either saline or CD95-Ab (n = 3 per group). (D) Western blotting for acetylated histone H3 (H3K9) & H4 (H4K5) on livers harvested 6 h after the injections of either saline or CD95-Ab. Histone H3 & β-actin were used as loading controls. Each line corresponds to an independent mouse. (E) Affinity purification-liquid chromatography tandem-mass spectrometry of liver proteins enriched by immunoprecipitation with acetyl lysine-specific antibody revealed 13 proteins with fold-change >2 in CD95-Ab vs. saline liver samples (n = 5per group). (F) Venn diagram including the 751 genes upregulated in livers of mice injected with CD95-Ab & the H3K9Ac ChIP-seg data on eight-week-old mouse livers in ENCODE showing a high degree of overlap (Hypergeometric test; p value: 9.643501e-58). (G) Chart generated using custom annotation scripts shows that response to external stimulus including inflammatory & defense response processes. & cytokine-cytokine receptor interaction pathways were enriched among the 467 genes overlapping between the two groups according to KEGG pathways. Means ± SDs are shown; t test; *p <0.05. Ab, antibody; nHDAC, nuclear histone deacetylase. (This figure appears in colour on the web.) Image collected & cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/29580866), licensed under a CC-BY license. Not internally tested by Novus Biologicals.

Western Blot: beta-Actin Antibody (AC-15) [NB600-501] - In vivo inhibition of histone acetylation by garcinol improves survival & reduces injury in acute liver failure. (A) Kaplan-Meier survival curves of mice receiving 20 mg/kg/day of garcinol (n = 8) or saline (n = 5) for 5 days prior to the injection of the CD95-Ab. Means ± SDs are shown. Long-rank test p = 0.0473. (B) Western blotting for histone H3 (H3) & acetylated histone H3 (H3K9) on livers harvested 6 h after the administrations of CD95-Ab. β -actin was used as a loading control. (C) Densitometric quantifications of H3K9 over H3 from the Western blotting shown in B; t test; *p <0.05. (D) Representative TUNEL (upper panels) & H&E (lower panels) staining of livers harvested 6 h after the administrations of CD95-Ab or saline. (E) Quantifications of TUNEL-positive cells & (F) Necrotic areas on livers of at least three sections from each mouse (at least n = 4 mice per group). Means ± SDs are shown. Likelihood ratio test for generalized linear model: * $p = 2.2 \times 10-16$. ANOVA plus Tukey's post hoc test; for necrotic area: *p = $2.45 \times 10-8$. Enzyme activities of PDHC (G) & LDH (H) in nuclear fractions (nPDHC & nLDH) of livers harvested 6 h after CD95-Ab injections in mice treated with garcinol (n = 5) or saline (n = 4). Means ± SDs are shown. ANOVA plus Tukey's post hoc test: p = 0.11 for nPDHC; p = 0.1 for nLDH. Ab, antibody; A.U., arbitrary units; nLDH, nuclear lactate dehydrogenase; nPDHC, nuclear pyruvate dehydrogenase complex; NS, not statistically significant; TUNEL, terminal deoxynucleotidyl transferase dUTP nick end labeling. (This figure appears in colour on the web.) Image collected & cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/29580866), licensed under a CC-BY license. Not internally tested by Novus Biologicals.

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Western Blot: beta-Actin Antibody (AC-15) [NB600-501] - Activation of the mTOR signaling pathway during spheroid formation on chitosan film. (a, Left panel) After 48 hr of seeding on monolayer or with chitosan coating (Spheroid), cell lysates were analyzed using the Human Phospho-MAPK Array kit. (a, Right panel) the spots of phospho-mTOR (S2448) were analyzed by an image analysis system. (b & c) Whole-cell lysates were analyzed by western blotting with specific antibodies against indicated molecules. β-tubulin & actin were used as the loading control. (d) Immunostaining of p-mTOR (S2448), p-S6 (S235), & p-4EBP1 in 3D spheres. DAPI merged images are shown in the low panel. Scale bar = 50 μ M. The results are expressed as the mean ± standard deviation of three independent experiments, which is representative of MSCs from two individuals. **p < 0.01 compared with monolayer cells. Image collected & cropped by CiteAb from the following publication (https://www.nature.com/articles/s41598-017-10708-0), licensed under a CC-BY license. Not internally tested by Novus Biologicals.

Western Blot: beta-Actin Antibody (AC-15) [NB600-501] - Triple AAV Vectors Drive Full-Length Protein Expression in the Pig Retina(A) Western blot (WB) analysis of lysates from Large White pig retina 2 months following subretinal injection of either single or triple AAV2/8 vectors encoding for ED under the control of the PR-specific promoter IRBP. The arrow indicates the full-length protein. (B) WB analysis of truncated products in lysates from Large White pig retina 2 months following subretinal injection of the combinations of triple ED-AAVs. Lysates of HEK293 cells infected with an ED-AAV 1 + 2 + 3 are shown on the left as positive control. The arrows indicate the following products: #A, ED full-length protein; #E, probably from AAV 3. α -3xflag, WB with anti-3xflag antibodies; α - β -Actin, WB with anti- β -Actin antibodies, used as loading control. Neg, lysates of retinas following injection with PBS. The molecular weight ladder is depicted on the left; 200 µg of proteins were loaded. Image collected & cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/29292161), licensed under a CC-BY license. Not internally tested by Novus Biologicals.

Western Blot: beta-Actin Antibody (AC-15) [NB600-501] - (a) Immunoblots of proteasome catalytic subunits in H727 cells transfected with siRNA targeting β 5, β 2 or β 5i. (b) The catalytic activity of β 5 subunit was decreased in H727 cells transfected with siRNA targeting
ß5 compared with H727 cells transfected with scrambled siRNA. (c) Effects of siRNA knockdown of \beta5, \beta5i, or \beta2 on Cfz sensitivity (IC50 values) in H23 & H727 cells. Data are shown as mean ± SD derived from a single non-linear regression based on n = 3 replicates per compound per concentration. (d) Heat map showing proteasome catalytic subunit activity profiles of 6 PI-naïve patient MM samples purchased from Conversant Biologics & AllCells (left). The numbers represent hydrolysis rates of respective substrates (RFU/min, mean values derived from three technical replicates) & were converted to color format & clustered by using the program "R" (http://www.R-project.org) (left panel). Carfilzomib (Cfz) cell viability of the same 6 patient MM cells was measured via CellTiter-Glo Luminescent Cell Viability Assay (right panel). Image collected & cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/30858500), licensed under a CC-BY license. Not internally tested by Novus Biologicals.







Western Blot: beta-Actin Antibody (AC-15) [NB600-501] - 14-3-37 suppresses hedgehog signalling to control the expression of p27Kip1.(a) Measurement of relative luciferase activity from various lengths of the Cdkn1b promoter in control or 14-3-32-depleted NIH-3T3 cells (n=7-8 per group, *P<0.05 when compared with control cells as assessed by Student's t-test). (b) Transcription factor binding sites within the -939-554 promoter region of Cdkn1b. (c) Hedgehog signalling (Gli-dependent) transcriptional activity in control or si14-3-3ζ-transfected Shh-light2 cells treated with 100 nM SAG. (n=4 per group, *P<0.05 when compared to control cells, as assessed by Student's t-test). (d) Expression levels of various components of the hedgehog signalling pathway in differentiating control & 14-3-3ζ-depleted 3T3-L1 cells, as measured by RNA-seq (n=4 per group, *P<0.05 when compared to control cells, as assessed by Student's t-test). (e) Immunoblotting of full length (□170 kDa) & processed (□80 kDa) Gli3 in 14-3-3ζ-depleted 3T3-L1 cells treated with MDI (representative of n=3 experiments). (f,g) Co-immunoprecipitation of endogenous (f) 14-3-3ζ or (g) Gli3 from lysates of 3T3-L1 cells treated with MDI for 24 h (representative of n=3 experiments) (h) ChIP-qPCR analysis of Gli3 occupancy on the Cdkn1b promoter in 3T3-L1 cells treated with MDI for 24 h (n=4 per group, *P<0.05, as assessed by Student's t-test). (i,j) Co-transfection of siRNA against 14-3-37 & Gli3 was used to examine whether knockdown of both proteins could restore differentiation, as determined by Ppary expression (i) or Oil Red-O staining (j) (n=4 independent experiments). Error bars represent s.e.m. Image collected & cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/26220403), licensed under a CC-BY license. Not internally tested by Novus Biologicals.

Western Blot: beta-Actin Antibody (AC-15) [NB600-501] - Silencing MFN1 or MFN2 reveals their distinct roles in DENV infection.(A & B) RTqPCR (A) & western blot (B) validation of A549 cells stably expressing shRNA targeting LacZ. MFN1 or MFN2. Data in panels A & C-F are mean ± SD (n = 3 per group) & were compared by two-tailed Student's t test. Quantification in panel B is the relative ratio of the indicated protein to actin expression. (C) LDH release assay of shLacZ-, shMFN1- or shMFN2-expressing A549 cells infected with DENV serotype 2 (moi 10) for the indicated time. (D) DENV plaque-forming assay of shLacZ-, shMFN1- or shMFN2-expressing A549 cells infected with DENV (moi 0.1) for 24 or 48 h. (E) Analysis of antiviral activity in culture media from indicated cells with DENV infection (moi 5, 48 h) against dSinF-Luc/2A virus. (F) RT-qPCR analysis of IFN β mRNA level in DENV infected (moi 10) A549-shLacZ, -shMFN1, & -shMFN2 cells for the indicated time. (G) Western blot analysis of DENV infected (moi 10) A549 cells expressing shLacZ, shMFN1 or shMFN2. The relative ratios of band intensity were quantified as indicated. (H) Flow cytometry with JC-1 staining of shLacZ-, shMFN1 or shMFN2-expressing A549 cells infected with DENV (moi 5) for 30 h. Decreased red/green fluorescence ratio of JC-1 represents disrupted MMP. CCCP, carbonyl cyanide m-chlorophenyl hydrazone, an ionophore that disrupts MMP. (I) Plague forming assay of vesicular stomatitis virus (VSV) by using A549 cells expressing shLacZ, shMFN1 or shMFN2 as indicated. pfu, plague-forming unit. Image collected & cropped by CiteAb from the following publication (https://dx.plos.org/10.1371/journal.ppat.1005350), licensed under a CC-BY license. Not internally tested by Novus Biologicals.

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Western Blot: beta-Actin Antibody (AC-15) [NB600-501] - Sortilin forms homodimers in the extracellular & intracellular domains with intermolecular disulfide bonds in HEK293 cells.A, schematic of FLAGsortilin Full, ECD+TMD, & ICD+TMD. SP, signal peptide; PP, propeptide. B & C, protein expression of FLAG-sortilin Full, ECD+TMD, & ICD+TMD was validated in reducing (B) & non-reducing (C) Western blotting using anti-FLAG antibody. FLAG-sortilin Full & ECD+TMD form homodimers & multimers. Empty vector was used as a control. D, HEK293 cells transiently overexpressing FLAG-sortilin Full or ECD+TMD were treated with a cross-linker, BS3, & the cell lysates were used for reducing Western blotting with anti-FLAG antibody, showing dimerization of sortilin Full & ECD+TMD (n = 3). E, HEK293 cells stably overexpressing FLAGsortilin ICD+TMD (FLAG-sortilin ICD+TMD HEK293 cells) were incubated with DMSO (Control), 20 µmol/liter MG-132 (MG) or 10 µmol/liter chloroquine (Chlo) for 7 h, & then reducing Western blotting was performed using anti-sortilin antibody. MG-132 increased the protein expression of FLAG-sortilin ICD+TMD, but chloroquine did not (n = 3). F. FLAG-sortilin ICD+TMD HEK293 cells were incubated with DMSO or MG-132 (2–20 µmol/liter) for 7 or 24 h. MG-132 increased FLAG-sortilin ICD+TMD in a time- & concentration-dependent manner (n = 3). G, following 16-h incubation of HEK293 cells (Control) or FLAG-sortilin ICD +TMD HEK293 cells (ICD+TMD) with MG-132 (5 µmol/liter) & immunoprecipitation with anti-FLAG antibody, non-reducing Western blotting showed dimerization of sortilin ICD+TMD using anti-sortilin antibody (n = 3). Monomers, homodimers, & multimers are abbreviated as MO, D, & MU, respectively. IB, immunoblotting. Image collected & cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/29382723), licensed under a CC-BY license. Not internally tested by Novus Biologicals. g Western Blot: beta-Actin Antibody (AC-15) [NB600-501] - Defects in adipogenesis due to 14-3-37 depletion are due to cell cycle arrest & upregulation of Cdkn1b/p27Kip1.(a–d) Analysis by flow cytometry (a) of control or si14-3-3ζ-transfected 3T3-L1 cells at different stages of the cell cycle at 0 (b), 24 (c) & 48 (d) hours following differentiation (n=4 independent experiments; *P<0.05 when compared with siContransfected cells, assessed by Student's t-test). (e) Analysis of various cell cycle regulatory genes by RNA-Seg from siCon- or si14-3-32 transfected 3T3-L1 adipocytes (n=4 per goup, *P<0.05 when compared with siCon cells at the same time point, assessed by Student's t-test). (f) Immunoblotting of p27Kip1 from lysates of differentiating of siCon- or si14-3-3ζ-3T3-L1 adipocytes (n=4 experiments). (g,h) Co-transfection of siRNA against 14-3-3ζ & p27Kip1 was used to examine whether knockdown of both proteins could restore differentiation, as determined by Ppary expression (g) or Oil Red-O staining (h). (n=4 independent experiments). Error bars represent s.e.m. Image collected & cropped by CiteAb from the following publication

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Western Blot: beta-Actin Antibody (AC-15) [NB600-501] - Effects of HSM d on IL-1 β gene expression & secretion in human macrophages.Cells were pre-treated with either 1 or 2% of HSM extract for 20 h, followed by treatment with LPS (0.5 µg/ml) for 3 h & with ATP (5 mM) for 1 h. (a) The mRNA expression levels of IL-1 β were determined by RT-PCR analysis. (b) IL-1 β mRNAs were quantified using real-time PCR. β -actin gene expression was used for normalization. The results are expressed as fold changes, considering one as the value of untreated cells. (c) The amount of IL-1 β in cell culture supernatants was detected by ELISA. (d) The presence of IL-1 β in cell lysates & cell culture supernatants were analyzed by Western blot analysis. Data are presented as means ± SE of three experiments performed in duplicate. #P < 0.01 versus untreated cells. *P < 0.01 versus HSM-untreated (ethanol-treated) control cells. †P < 0.05 versus HSM (1%) treated cells. Image collected & cropped by CiteAb from the following publication

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Western Blot: beta-Actin Antibody (AC-15) [NB600-501] - (A) shows western blots of nuclear extracts of mCCD cells treated with 100 µmol/L CoCl2. Treatment with CoCl2 increased stabilization of HIF1a in the nucleus of the cells. GAPDH levels are shown as loading controls. (B) shows western blots of whole cell extracts of mCCD cells treated with 100 µmol/L CoCl2 for 4 h. As shown in the images, treatment with CoCl2 increased the EPO production by cells. Beta actin levels are shown as loading controls. (C) shows western blots of whole cell extracts of mCCD cells treated with 100 µmol/L CoCl2 for 4 h & chronic fluid flow. As shown in the images, treatments decreased ZO 1 production by cells independently of fluid flow stimulation. Beta actin levels are shown as loading controls. (D) shows western blots of whole cell extracts of mCCD cells treated with 100 µmol/L CoCl2 for 4 h & chronic fluid flow. As shown in the images, treatments decreased NaKATPase production by cells independently of fluid flow stimulation. Beta actin levels are shown as loading controls. (E) Densitometric analysis of Western blots presented in (A–D). HIF1 α level: P = 0.004 (N = 3), t test, showing significant nuclear stabilization of HIF1a. EPO level: increases slightly though not statistically significant (N = 3, P > 0.05, One \Box way ANOVA). ZO \Box 1 level: P < 0.0001 (N = 4), One \Box way ANOVA, showing significant decrease in ZO 1 levels, independent of fluid flow. NaKATPase α1 subunit level: P = 0.001 (N = 3), One □ way ANOVA, showing significant decrease in NaKATPase a1 subunit levels, largely independent of fluid flow. Image collected & cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/29263117), licensed under a CC-BY license. Not internally tested by Novus Biologicals.







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Western Blot: beta-Actin Antibody (AC-15) [NB600-501] - Proteasome inhibitors restored the expression of MFN2 & induced apoptosis under hypoxic conditions. HPASMCs were exposed to hypoxic conditions & were treated with bortezomib & marizomib for 24 h. The expression of MFN2, pro-PARP1, & cleaved PARP1 were detected by Western blotting. The expression of actin was used as a loading control. Bortezomib (A) & marizomib (B) restored the expression of MFN2inhibited by hypoxia, inhibited the levels of pro-PARP1, & induced the level of cleaved PARP1. One-way ANOVA was used to determine the differences between the experimental & vehicle control (DMSO) groups & the Newman–Keuls Multiple Comparison Test was used as a post hoc test following ANOVA. The data represent means ±standard error of three independent experiments. * p < 0.05 versus normoxic group. # p < 0.050.05 versus hypoxic control group. Image collected & cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/35453623), licensed under a CC-BY license. Not internally tested by Novus Biologicals. Western Blot: beta-Actin Antibody (AC-15) [NB600-501] - Ad.5-TCTV 5-TCT С 5-CT induces ER stress & apoptosis in cancer cells. (A) Breast cancer cells (MDA-MB-231 & SUM-159) treated with 25 pfu of Ad.5-E1A or the indicated pfu of Ad.5-CTV or Ad.5-TCTV for 72 h. Cells were then 50 25 50 collected & Western blotting analysis was performed for signaling MDA-7/IL-24 molecules using specific antibodies & β-actin served as a loading E1A control. (B) Breast cancer cells were cultured in 8-well chamber slides & Luciferase infected with 25 pfu of Ad.5-E1A, Ad.5-CTV, or Ad.5-TCTV for 72 h. PARP Cells were fixed & TUNEL assays were performed. Data presented as TUNEL positive cells in a defined microscopic field as compared with un-Bcl-2 treated control cells. (C) Prostate cancer cells (DU-145 & PC-3) were GRP-78 treated as described in A, cells were then collected & Western blotting GADD153 analysis was performed for signaling molecules using specific antibodies β-Actin & β-actin served as a loading control. (D) Prostate cancer cells were DU-145 treated as described in B & cells were fixed & TUNEL assays were PC-3 performed. Data presented as TUNEL positive cells in a defined microscopic field as compared with un-treated control cells. Scale bar: 200 µM. Image collected & cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/33670594), licensed under a CC-BY license. Not internally tested by Novus Biologicals. Α Western Blot: beta-Actin Antibody (AC-15) [NB600-501] - Pyruvate dehydrogenase kinase 1 (PDK 1) regulates DRP1 mediated mitochondrial fission & glycolytic phenotype in BRAFV600E cells. (A, B) Western blot & representative confocal micrographs (2.5x zoom, 60x original magnification) of si-PDK1 silenced BRAFV600E CRC cells, showing a reduction in both pDRP1S616 levels as well as mitochondrial fission after PDK1 knockdown, 30-40 cells were guantified (n=3, mean± HT 29 Colo 205 SEM), red= mitochondria, CMX Ros stained & blue= Nuclei, DAPI K Da stained, statistical analysis was done by two way ANOVA, followed by 75 pDRP1^{S616} Bonferroni post-tests comparing replicate means by column with corresponding p values shown. (C, D) Genetic & pharmacologic knockdown of PDK1 in BRAFV600E CRC cells leads to a reduction in 75 DRP1 cell proliferation (n= 4, mean± SEM) & clonogenic rates in soft agar (n=3, mean± SEM) respectively; statistical analysis for proliferation was done by 2-way ANOVA followed by Bonferroni post-tests comparing replicate means by row with corresponding p values shown, & statistical analysis PDK1 for soft agar clonogenic assay was done by unpaired t-test followed by 50 Welch's correction assuming unequal variance in means, corresponding p values shown. Image collected & cropped by CiteAb from the following **B-Actin** publication (https://pubmed.ncbi.nlm.nih.gov/33738242), licensed under

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Western Blot: beta-Actin Antibody (AC-15) [NB600-501] - Lack of HIF-P4H-1 increases the amount of p53. (a) Western blot analysis of p53 in wt & Hif-p4h-1-/- MEFs cultured in 21% or 1% O2 for 24 h. (b,c) Western blot analysis of p53 in Hif-p4h-1 & scrambled (Scrm) siRNA transfected HEK293 cells (b) & wt MEFs treated with 50 µM FG4497 for 24 h (c). (d) gPCR analysis of p53 mRNA in wt & Hif-p4h-1-/- MEFs cultured in 21% or 1% O2 for 24 h. (e) Analysis of p53 protein turnover rate. Hif-p4h-1-/- & wt MEFs were treated with 200 µg/ml of cycloheximide for the indicated time points & whole cell lysates were blotted for p53. (f) Western blot analysis of MDM2 in wt & Hif-p4h-1-/-MEFs cultured in 21% or 1% O2 for 24 h. (g) Western blot analysis of p53 & MDM2 in wt MEFs treated with 10 µM nutlin-3a for 24 h with or without overexpression of human HIF-P4H-1 (OE). (h,i) Western blot analysis of HIF1α in wt & Hif-p4h-1-/- MEFs cultured in 21% or 1% O2 for 24 h (h) & in scrambled & Hif-p4h-1 siRNA transfected HEK293 cells (i). (j) Western blot analysis of ubiquitination of p53 in Hif-p4h-1 & scrambled siRNA transfected HEK293 cells. The cells were transfected with HA-ubiquitin & endogenous p53 was immunoprecipitated followed by Western blotting with anti-HA & anti-P53 antibodies. (k) Western blot analysis of p53 in Hif-p4h-1-/- & wt MEFs were treated with or without 10 µM MG132 for 5 h. Data are presented as representative Western blots & as mean ± s.d., n = at least 3 individual MEF isolates or experiments. *P < 0.05, **P < 0.01 & ***P < 0.001, two-tailed Student's ttest. Unprocessed original scans of blots are shown in Supplementary Fig. 5. Image collected & cropped by CiteAb from the following publication (https://www.nature.com/articles/s41598-017-17376-0), licensed under a CC-BY license. Not internally tested by Novus Biologicals.

Western Blot: beta-Actin Antibody (AC-15) [NB600-501] - Lack of HIF-P4H-1 suppresses NF-kB activity. (a) Western blot analysis of Rel-A in wt & Hif-p4h-1-/- MEFs subjected to 21% or 1% O2 for 24 h. (b-d) Analysis of NF-kB luciferase reporter activity in wt & Hif-p4h-1-/- MEFs (b), in Hif-p4h-1 & scrambled (Scrm) siRNA transfected HEK293 cells (c) & in MDA-MB-231 cells treated with 50 µM FG4497 for 6 h (d). Cells were harvested 48 h after the NF-kB luciferase reporter plasmid transfection. siRNA transfection was performed 24 h before the NF-κB luciferase reporter plasmid transfection. FG4497 was added 6 h before the cell harvest. (e) Western blot analysis of Rel-A, p-Rel-A, IkBa, p105 & p50 in wt & Hif-p4h-1-/- MEFs treated with or without 200 ng/ml LPS for 12 h. (f,g) Analysis of ReI-A & p-ReI-A by Western blotting (f) & NF-κB luciferase reporter activity (g) in wt MEFs treated with or without 200 ng/ml LPS & 50 µM FG4497 for 6 h. (h,i) Analysis of Rel-A & p-Rel-A by Western blotting (h) & NF-kB luciferase reporter activity (i) in Hif-p4h-1-/ - MEFs transfected with empty vector (EV) or a vector encoding V5tagged human recombinant HIF-P4H-1 (OE) & treated with or without 200 ng/ml LPS for 12 h. (j) qPCR analysis of Rel-A mRNA in Hif-p4h-1-/ – & wt MEFs treated with 200 ng/ml of LPS for 12 h. (k) Western blot analysis of ReI-A & p-ReI-A in cytosolic & nuclear fractions of Hif-p4h-1-/ - & wt MEFs. (I) NF-κB luciferase reporter activity in HIF1A-/- HCT116 cells treated with FG4497 or transfected with HIF-P4H-1 siRNA. Data are presented as representative Western blots & as mean ± s.d., n = 3-4 individual MEF isolates or experiments. *P < 0.05, **P < 0.01 & ***P < 0.001, two-tailed Student's t-test. Unprocessed original scans of blots are shown in Supplementary Fig. 5. Image collected & cropped by CiteAb from the following publication

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Western Blot: beta-Actin Antibody (AC-15) [NB600-501] - CD95-induced d senescence is mediated by low-level canonical caspase signaling. (a) CRC29 colonospheres were exposed once (cycle 1) or chronically exposed to CD95L (10 ng/ml) for 2 weeks (cycle 2), & collected at the indicated time points. Cells were collected & analyzed for the presence of activated (cleaved) caspase-3 by western blotting. (b-c) CRC29 colonospheres were exposed to CD95L (5 ng/ml) in the presence or absence of zVAD (25 µM) for 1 or 7 days & were then processed for imunofluorescence analysis of caspase-3 activation (b) & DNA damage (y-H2AX) (c) An overview of the significance of all comparisons is provided in Supplementary Table S1. (d) Colonspheres were exposed to CD95L (10 ng/ml) for 24 h in the presence or absence of zVAD (25 µM) followed by western blot analysis of caspase cleavage & iCAD processing. (e & f) CRC29 colonospheres were exposed to increasing concentrations of CD95L as indicated in the presence or absence of zVAD (25 μ M), after which SA-βGAL (e) & clone-forming capacity (f) were assessed at 7 or 14 days, respectively. The asterisks indicate significant differences (ordinary one-way ANOVA) (****P<0.00001; **P<0.001). An overview of the significance of all comparisons is provided in Supplementary Table S1 Image collected & cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/28300842). licensed under a CC-BY license. Not internally tested by Novus Biologicals.

Western Blot: beta-Actin Antibody (AC-15) [NB600-501] - mRNA & protein expression in Subject 1 & control lymphoblast cells. A. Scatter graph of mRNA expression of EXOC6, RPJB and HES1in lymphoblasts. Values for EXOC6, RPJB and HES1 mRNA expression were normalized to the corresponding ß-actin mRNA levels. The results derived from at least three sets of samples & the mean level of each sample is represented in the graph, horizontal bar is an average value between three controls. The difference between the study sample & the average of the controls has been evaluated by student t-test (*, p < 0.05; **, P < 0.01). B. RBPJ protein expression. C. HES1 protein expression. C1-C5 indicate control samples from the lymphoblasts, S1 indicate study sample from Subject1. Cell lysates were analyzed by SDS-PAGE & immunoblotting with membrane probed for RBPJ, HES1 & B-actin, Values for RBPJ & HES1 expression were normalized to the corresponding ß-actin levels. The results derived from at least three sets of samples & the mean value of each sample is represented in the bar graph. The difference between Subject1 & the average of three controls has been evaluated by student t-test (*, p < 0.05). Image collected & cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/23837398), licensed under a CC-BY license. Not internally tested by Novus Biologicals.





Western Blot: beta-Actin Antibody (AC-15) [NB600-501] - Ketogenic diet induces anti-inflammatory IL-4 & Arginase-1 expression in D2 retina & Keto D2 Unt D2 D2G ON. a Bar graph showing II-4 mRNA normalized to Hprt mRNA by gRT-PCR in untreated (Unt) & keto D2 retina, *p = 0.0264; n = 8 samples per group. b, c IL-4 protein levels in the retina (b), **p = 0.0014, & ON (c), *p Arginase-1 → = 0.0121, analyzed by ELISA; n = 8 samples per group. dArg1 mRNA 40 kDa levels in retina normalized by Hprt mRNA levels, *p = 0.0368; n = 8 samples per group. e Immunohistochemical analysis of Arginase-1 Actin → (green) in the untreated & keto D2 retina & ON. Iba1 (red) labels 45 kDa microglia & DAPI stains nuclei. f, g Percentage of mean fluorescence of Arginase-1 in inner retina (f), ***p = 0.0001, & in the ROI of the proximal ON (g), ***p = 0.0001; five retinas & ON from each group were analyzed, with five sections from each individual retina & ON. h Western blot analysis of Arginase-1 protein in the retina. i Quantification by densitometry of Arginase-1 levels normalized to β -actin levels, ***p = 0.0001; n = 3 blots per group, each with independent samples. j Arginase-1 protein in the ON normalized to β -actin levels, as determined by capillary electrophoresis, **p = 0.0006; n = 3 ON per group. All bar graphs are presented as the mean ± SEM, analyzed by two-tailed unpaired t test. Scale bar, 20 µm Image collected & cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/30424795), licensed under a CC-BY license. Not internally tested by Novus Biologicals. Western Blot: beta-Actin Antibody (AC-15) [NB600-501] - Substituting the transmembrane domain of sortilin with the corresponding domain of CD43 does not decrease the dimeric form of sortilin.A. schematic of FLAG-sortilin FLAG-sortilin wildtype (WT) & FLAG-sortilin CD43-TMD. The transmembrane domain of sortilin was replaced with that of CD43. SP, CD43signal peptide; PP, propeptide. B, FLAG-sortilin WT & FLAG-sortilin Control WT TMD CD43-TMD were transiently overexpressed in HEK293 cells, & nonreducing Western blotting was carried out using cell lysate with antikDa FLAG antibody (n = 3). Monomers, homodimers, & multimers are MU abbreviated as MO, D, & MU, respectively. C & D, His6-sortilin WT or 250 His6-sortilin CD43-TMD was transiently overexpressed in HEK293 cells stably overexpressing FLAG-sortilin, & immunoprecipitation was performed using anti-FLAG M2 antibody. Western blotting was carried 150 out using whole-cell lysates (C) & immunoprecipitants (D). His6-sortilin CD43-TMD coprecipitated with FLAG-sortilin as well as His6-sortilin WT. Arrows, sortilin wildtype or sortilin CD43-TMD (n = 3). E, in FLAG-sortilin 100 HEK293 cells or HEK293 cells, His6-sortilin CD43-TMD was MO overexpressed. The cells were subjected to TR-FRET assay. Change of FRET signal by expression of His6-sortilin WT or CD43-TMD is indicated 75 by percent change (mean \pm S.D., n = 4, one independent experiment). Error bars represent S.D. *, p < 0.05; **, p < 0.01 by t test. F & G, in FLAG-sortilin HEK293 cells, His6-sortilin WT or His6-sortilin CD43-TMD 50 were overexpressed. The cell lysates were subjected to non-reducing Western blotting with anti-FLAG antibody (F) & anti-His6 antibody (G), **IB: FLAG** demonstrating that substituting the transmembrane domain of sortilin with that of CD43 did not decrease dimerization (n = 3). IB, 50 immunoblotting. Image collected & cropped by CiteAb from the following 37 publication (https://pubmed.ncbi.nlm.nih.gov/29382723), licensed under a CC-BY license. Not internally tested by Novus Biologicals. IB: β-actin



D



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Western Blot: beta-Actin Antibody (AC-15) [NB600-501] - Sortilin S316E & sortilin wp increase dimerization in HEK293 cells, & the addition of propeptide decreases dimerization in the extracellular vesicles of FLAGsortilin HEK293 cells.A, schematic of FLAG-sortilin WT, S316E, & wp. Serine 316 was replaced by glutamic acid in FLAG-sortilin S316E. Propeptide was removed in FLAG-sortilin wp. SP, signal peptide; PP, propeptide. B, S316E increased dimerization of sortilin in HEK293 cells (n = 3). C, removal of propertide increased dimerization of sortilin in HEK293 cells (n = 3). D & E, addition of propeptide (100 nmol/liter) decreased dimerization of sortilin in the extracellular vesicles of FLAGsortilin HEK293 cells (E), whereas a decrease in the cells was not observed (D) (n = 2). Monomers & homodimers of high & low molecular weight are abbreviated as MO, D(HMW), & D(LMW), respectively. Vec, vector; IB, immunoblotting. Image collected & cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/29382723), licensed under a CC-BY license. Not internally tested by Novus Biologicals.

Western Blot: beta-Actin Antibody (AC-15) [NB600-501] - Chronic CD95 stimulation reduces colony-forming capacity in human colonospheres. (a) Human colonosphere cultures isolated from the tumors of nine distinct colon cancer patients were pretreated with FC control or FC-CD95L (10 ng/ml) for 2 weeks, seeded as single cells in Matrigel & cultured continuously in the absence or presence of FC-CD95L. Colony formation was scored after 2 weeks. The experiments were performed multiple times with similar results (L145: n=4, L146: n=4, L167: n=3, CRC47 n=4, CRC29 n=3, L193 n=2, CRC26 n=1, L169 n=4, CR16 n=3). One representative experiment is shown (triplicate values). (b) The expression of CD95 was measured with FACS (CH11, left panel) & with western blotting (middle panel; asterisks indicate non-specific bands). The FACS results are plotted as the percentage CH11-positive cells over control for each cell line. Averaged FACS values were then plotted against the percentage growth inhibition (right panel). The Pearson correlation & the accompanying P-value are shown in the inset. (c) CRC29 cells were cultured in the presence of FC control or FC-CD95L for 2 weeks & were then processed for FACS analysis of DNA content using PI. (d) CRC29 cells were cultured in the presence of control or CD95L for 2 weeks, labeled with CFSE, & subsequently plated in Matrigel as single cells. Cells were collected from the Matrigel & analyzed for maintenance of the fluorescent signal by FACS at days 2, 4, & 7 after plating. (e) The experiment was performed as in a, comparing a 2-day treatment to chronic stimulation with FC-CD95L during colony formation Image collected & cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/28300842), licensed under a CC-BY license. Not internally tested by Novus Biologicals.









Western Blot: beta-Actin Antibody (AC-15) [NB600-501] - Lack of HIF-P4H-1 increases the amount of p53. (a) Western blot analysis of p53 in wt & Hif-p4h-1-/- MEFs cultured in 21% or 1% O2 for 24 h. (b,c) Western blot analysis of p53 in Hif-p4h-1 & scrambled (Scrm) siRNA transfected HEK293 cells (b) & wt MEFs treated with 50 µM FG4497 for 24 h (c). (d) qPCR analysis of p53 mRNA in wt & Hif-p4h-1-/- MEFs cultured in 21% or 1% O2 for 24 h. (e) Analysis of p53 protein turnover rate. Hif-p4h-1-/- & wt MEFs were treated with 200 μ g/ml of cycloheximide for the indicated time points & whole cell lysates were blotted for p53. (f) Western blot analysis of MDM2 in wt & Hif-p4h-1-/-MEFs cultured in 21% or 1% O2 for 24 h. (g) Western blot analysis of p53 & MDM2 in wt MEFs treated with 10 µM nutlin-3a for 24 h with or without overexpression of human HIF-P4H-1 (OE). (h,i) Western blot analysis of HIF1 α in wt & Hif-p4h-1-/- MEFs cultured in 21% or 1% O2 for 24 h (h) & in scrambled & Hif-p4h-1 siRNA transfected HEK293 cells (i). (j) Western blot analysis of ubiquitination of p53 in Hif-p4h-1 & scrambled siRNA transfected HEK293 cells. The cells were transfected with HA-ubiquitin & endogenous p53 was immunoprecipitated followed by Western blotting with anti-HA & anti-P53 antibodies. (k) Western blot analysis of p53 in Hif-p4h-1-/- & wt MEFs were treated with or without 10 µM MG132 for 5 h. Data are presented as representative Western blots & as mean ± s.d., n = at least 3 individual MEF isolates or experiments. *P < 0.05, **P < 0.01 & ***P < 0.001, two-tailed Student's ttest. Unprocessed original scans of blots are shown in Supplementary Fig. 5. Image collected & cropped by CiteAb from the following publication (https://www.nature.com/articles/s41598-017-17376-0), licensed under a CC-BY license. Not internally tested by Novus Biologicals.

Western Blot: beta-Actin Antibody (AC-15) [NB600-501] - Celecoxib induced cell cycle arrest at the G2 phase in CGCCA cells.CGCCA cells were treated with the indicated concentration of celecoxib for 1 day, & the cell cycle distribution was analysed. (A & B) After 1 day of celecoxib treatment, CGCCA cells were analysed by flow cytometry to determine cell cycle distribution. (C) Treated CGCCA cells were stained with PI to show condensed chromatin (white arrow) in M-phase cells. In the right lower panel, the (+) & (-) are examples of cells with & without condensed chromatin, respectively. (D) Ratio of CGCCA cells at M-phase. (E) Expression levels of Cdc25C & CDK1 in CGCCA cells after 1 day of treatment with the indicated concentrations of celecoxib as measured using western blot. Results are presented as % of the control. Each value is a mean ± SD of 3 to 5 determinations. *p<0.05, **p<0.001 (versus control). Image collected & cropped by CiteAb from the following publication (https://dx.plos.org/10.1371/journal.pone.0069928), licensed under a CC-BY license. Not internally tested by Novus Biologicals.





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#P < 0.01 versus untreated cells. *P < 0.01 versus HSM-untreated control (ethanol) cells. †P < 0.05 versus HSM (1%) treated cells. Image collected & cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/23459183), licensed under a CC-BY license. Not internally tested by Novus Biologicals.







Western Blot: beta-Actin Antibody (AC-15) [NB600-501] - Sortilin forms homodimers in the extracellular & intracellular domains with intermolecular disulfide bonds in HEK293 cells.A, schematic of FLAGsortilin Full, ECD+TMD, & ICD+TMD. SP, signal peptide; PP, propeptide. B & C, protein expression of FLAG-sortilin Full, ECD+TMD, & ICD+TMD was validated in reducing (B) & non-reducing (C) Western blotting using anti-FLAG antibody. FLAG-sortilin Full & ECD+TMD form homodimers & multimers. Empty vector was used as a control. D, HEK293 cells transiently overexpressing FLAG-sortilin Full or ECD+TMD were treated with a cross-linker, BS3, & the cell lysates were used for reducing Western blotting with anti-FLAG antibody, showing dimerization of sortilin Full & ECD+TMD (n = 3). E, HEK293 cells stably overexpressing FLAGsortilin ICD+TMD (FLAG-sortilin ICD+TMD HEK293 cells) were incubated with DMSO (Control), 20 µmol/liter MG-132 (MG) or 10 µmol/liter chloroquine (Chlo) for 7 h, & then reducing Western blotting was performed using anti-sortilin antibody. MG-132 increased the protein expression of FLAG-sortilin ICD+TMD, but chloroquine did not (n = 3). F. FLAG-sortilin ICD+TMD HEK293 cells were incubated with DMSO or MG-132 (2–20 µmol/liter) for 7 or 24 h. MG-132 increased FLAG-sortilin ICD+TMD in a time- & concentration-dependent manner (n = 3). G, following 16-h incubation of HEK293 cells (Control) or FLAG-sortilin ICD +TMD HEK293 cells (ICD+TMD) with MG-132 (5 µmol/liter) & immunoprecipitation with anti-FLAG antibody, non-reducing Western blotting showed dimerization of sortilin ICD+TMD using anti-sortilin antibody (n = 3). Monomers, homodimers, & multimers are abbreviated as MO, D, & MU, respectively. IB, immunoblotting. Image collected & cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/29382723), licensed under a CC-BY license. Not internally tested by Novus Biologicals.

Western Blot: beta-Actin Antibody (AC-15) [NB600-501] - RAF1/ERK signalling is preferentially required for PANC1 cell survival in the absence of exogenous insulin. (A) Effects of different small molecule inhibitors on propidium iodide (PI) incorporation (PI) in PANC1 cells were tracked & expressed as the fold change in the percent of PI & Hoechst co-positive cells over total Hoechst positive cells at that hour relative to t =0 hour. Kinetic data were analyzed relative to serum-free control by two-way ANOVA (n = 3) Data points that have been shaded solid black represent statistical significance when compared to non-treated conditions at that time point. # Indicates statistical significance in cells treated with Akti1/2 when compared to control at that time point. (B) Average number of PI positive cells over time of each treated group in Figure 5A is shown as a histogram expressed in arbitrary units (AU). GW5074 exhibited statistical significance, where as other treatments did not yield significance. U0126 p = 0.38, GW5074 *p = 0.0005, Akti-1/2 p =0.395, Wort. p = 0.292 (n = 3). (C) The effect of 24 hours treatment with inhibitors on cleaved caspase 3 protein levels in PANC1 cells. This is a representative immunoblot of three independent biological replicates (n =3). (D-E) PANC1 cells were serum starved & treated with either DMSO. 10 µM GW5074, 10 µM U0126, 200 nM Akti-1/2 & 1 mM wortmannin (wort.) for 24 hours & 120 hours (n =4-5). Cell viability of PANC1 cells was expressed as the fold change of the treated relative to control. (C-E) One-way ANOVA analysis with Bonferroni post-test was performed. *Represents statistical significance of p < 0.05 where treated groups are compared to control (-) in the post-hoc test. Image collected & cropped by CiteAb from the following publication

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Western Blot: beta-Actin Antibody (AC-15) [NB600-501] - Nifedipine & the expression p-AMPK, SREBP-1/2(nuclear), PPAR-α (nuclear) & lipin-1 (nuclear) in 48 h. (a) The AMPK/actin was upregulated to 107.1%, 112.6%, & 116.4%, significantly at doses of 7.5, 15, & 30 µM (p < 0.05), the ratio p-AMPK/AMPK was downregulated to 82.1%, 58.7%, & 49.9%, respectively. (b,c) SREBP-1 was activated in control. The SREBP-1/HDAC was upregulated to 109.9% & 132.0% by nifedipine at doses of 7.5 & 30 μ M, respectively (p < 0.01); The SREBP-2 were elevated to 127.5%, 146.7% & 161.0% by nifedipine at doses of 7.5, 15 & 30 µM, respectively (p < 0.05 in 7.5 μ M, p < 0.01 in 15 & 30 μ M) (d) nifedipine downregulated PPAR-a. The PPARa/Lamin B was suppressed from 80.2%, 85.6%, to 96.4% compared to control (100%) upon administration of nifedipine at doses 7.5, 15 & 30 μ M, respectively. (p < 0.05 in 15 & 30 µM) (e) Lipin-1 was activated in control. The lipin-1/lamin B was upregulated after nifedipine treatment to 102.5%, 115.8% & 121.1%, respectively, by nifedipine at doses of 7.5, 15, & 30 µM, respectively (p < 0.05 in 15 & 30 μ M). p-values ≤ 0.05 (marked as *) were considered statistically significant. In addition, p-values \leq 0.01 are marked as **. Image collected & cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/30934807), licensed under a CC-BY license. Not internally tested by Novus Biologicals.

Western Blot: beta-Actin Antibody (AC-15) [NB600-501] - JEV NS5 interacts with mitochondrial trifunctional protein (MTP), the enzyme complex involved in LCFA β-oxidation.(A) Western blot analysis of protein levels of the indicated proteins in A549 cells with JEV infection (MOI = 10) or NS5-Flag overexpression. (B) Western blot analysis of V5tag, Flag-tag, & actin in HEK293T cells co-transfected with HADHα-V5-His plus the indicated plasmids expressing Flag-tagged JEV viral proteins for 24 h, then immunoprecipitated with anti-Flag affinity gel. WCL, whole-cell lysates. (C) Immunoprecipitation (IP) analysis with control IgG or anti-Flag affinity gel in A549, GFP-A549 & NS5-Flag-A549 🖁 cells. The protein bands identified as HADH α & HADH β are indicated by arrows. (D) IP with anti-Flag affinity gel & Western blot analysis with the indicated antibodies in HEK293T cells co-transfected with vector control or NS5-Flag plus HADH α -V5-His or HADH β -HA for 24 h. (E) IP analysis with nickel beads & Western blot analysis with the indicated antibodies in HEK293T cells adsorbed with JEV for 3 h, then transfected with vector control, HADHα-V5-His or HADHβ-V5-His for 24 h. Image collected & cropped by CiteAb from the following publication (https://dx.plos.org/10.1371/journal.ppat.1004750), licensed under a CC-

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Western Blot: beta-Actin Antibody (AC-15) [NB600-501] - The effect of AM1241, SB203580, & PDTC on the thermal threshold & the expression of P2Y12 & P2Y13 receptors. Data are presented as TWL. All values represent mean ± standard deviation. a Comparison of thermal stimulation pain threshold in the ipsilateral hind paw of rats (n = 8). Spinal ADPbetaS injection reduced TWL from days 3 to 7 after ADPbetaS injection. *P < 0.05 compared w/ control group. Pretreated w/ AM1241 (100 PM, 20 min), SB203580 (50 μM), or PDTC (5 μg/10 μl) significantly inhibited the reduced w/drawal threshold days 3 to 7 in ADPbetaS-treated rats, respectively (compared w/ ADPbetaS-treated rats, AM1241: P < 0.05; SB203580: P < 0.05; PDTC: P < 0.05). b RT-PCR results show the expression of P2Y12 mRNA expression in the dorsal spinal cord of the rats in every groups (n = 8 per group). *P < 0.05 means comparison w/ the control group; +P < 0.05 means comparison w/ the ADPbetaS-treated rats. c RT-PCR results show the expression of P2Y13 mRNA expression in the dorsal spinal cord of the rats in every group (n = 8 per group). P < 0.05 means comparison w/ the control group; +P < 0.05 means comparison w/ the ADPbetaS-treated rats. d Western blotting image of P2Y12R expression. The top panel the target band, P2Y12R, & the bottom one for the loading control beta-actin. e Western blotting quantitative analysis of the P2Y12R expression in the dorsal spinal cord of the rats. P < 0.05 means comparison w/ the control; +P < 0.05 means comparison w/ the ADPbetaS-treated rats. f Western blotting quantitative analysis of the P2Y13R expression in the dorsal spinal cord of the rats. *P < 0.05 means comparison w/ the control: +P < 0.05 means comparison w/ the ADPbetaS-treated rats Image collected & cropped by CiteAb from the following publication (http://jneuroinflammation.biomedcentral.com/articles/10.1186/s12974-017-0960-0), licensed under a CC-BY license. Not internally tested by Novus Biologicals.

Western Blot: beta-Actin Antibody (AC-15) [NB600-501] - Effects of insulin on AKT & ERK phosphorylation & cell viability in primary human pancreatic duct cells. Phosphorylated AKT & ERK were measured in primary pancreatic exocrine cultures treated with the indicated concentrations of insulin for 5 minutes (A, B) & 24 hours (C, D) (n = 3-4) Fold refers to the fold change of sample relative to control at the same time point. (E) Quantification of automated cell-counting studies employing live-cell imaging of Hoechst-labeled cell cultures over 60 hours. (n = 3). (F) Quantification of proliferation by BrdU staining of treated relative to untreated over 3 days (n =4). (G) Quantification of the average number of dying/dead treated cells, propidium iodide (PI) labeled, over 60 hours relative to non-treated cells. (n =3). (H) Human exocrine cells were exposed to 0, 0.2, 2, 20, 200 nM insulin for 3 days. Bright-field images are representative of 3 cultures. (I) Effects of inhibition of RAF1/ERK signalling on PI incorporation with 10 µM GW5074 or AKT signalling with 100 nM Akti1/2 on human primary pancreatic exocrine cell viability (n = 3). SF denotes serum free. Repeated Measures ANOVA analyses with Bonferroni's post-test were performed. *Represents statistical significance of p < 0.05 when compared to DMSO control. Image collected & cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/25373319), licensed under a CC-BY license. Not internally tested by Novus Biologicals.







Western Blot: beta-Actin Antibody (AC-15) [NB600-501] - Intrathecal injection of SB203580 (50 μM) or PDTC (5 μg/10 ml) suppressed the increased expression of P2Y12 & P2Y13R in the dorsal spinal cord of CCI rats. a Comparison of thermal stimulation pain threshold in the ipsilateral hind paw of the rats (n = 8). Data are presented as TWL. CCI reduced TWL from days 1 to 7 after nerve injury. *P < 0.05 compared with control group. Pretreated with SB203580 (50 µM) or PDTC (5 µg/10 ml) significantly inhibited the reduced withdrawal threshold days 1 to 7 in CCI rats, respectively (compared with vehicle-treated CCI group, SB203580: +P < 0.05; PDTC: +P < 0.05). b RT-PCR results show the expression of P2Y12 mRNA expression in the rat dorsal spinal cord (n = 8 per group). *P < 0.05 means comparison with the control group; +P < 0.05 means comparison with the vehicle-treated CCI group. c RT-PCR results show the expression of P2Y13 mRNA expression in the rat dorsal spinal cord (n = 8 per group). P < 0.05 means comparison with the control group; +P < 0.05 means comparison with the vehicle-treated CCI group. d Western blotting image of P2Y12R & P2Y13R expression. The top panel was the target band, & the bottom one was for the loading control beta-actin. e Western blotting quantitative analysis of the P2Y12R expression in the rat dorsal spinal cord. *P < 0.05 means comparison with the sham group; +P < 0.05 means comparison with the vehicletreated CCI group. f Western blotting quantitative analysis of the P2Y13R expression in the rat dorsal spinal cord. *P < 0.05 means comparison with the sham group; +P < 0.05 means comparison with the vehicletreated CCI group Image collected & cropped by CiteAb from the following publication

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Western Blot: beta-Actin Antibody (AC-15) [NB600-501] - Effects of HSM a on the expression of P2X4R & P2X7R & ROS production in human macrophages. Cells were pretreated with various concentrations (1 or 2%) of HSM extract for 20 h, followed by treatment with LPS (0.5 µg/ml) for 3 h & ATP (5 mM) for 1 h. (a) The mRNA expression levels of P2X4R & P2X7R were determined by RT-PCR, using β -actin as the internal control. (b) P2X4R & P2X7R mRNAs were quantified using real-time PCR. β-actin gene expression was used for normalization. The results are expressed as fold changes, considering one as the value of untreated cells. (c) Cell lysates were analyzed by Western blot analysis used specific anti-P2X4R & anti-P2X7R antibodies. (d) ROS production was measured with the total ROS detection kit, using a fluorescence microplate reader. Pyocyanin (200 µM), a ROS inducer, was used as a positive control for ROS formation. Data are presented as means ± SE of three experiments preformed in duplicate. #P < 0.01 versus untreated cells. versus HSM-untreated control (ethanol) cells. *P < 0.01 versus HSM-untreated control (ethanol) cells. †P < 0.05 versus HSM (1%) treated cells. Image collected & cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/23459183), licensed under a CC-BY license. Not internally tested by Novus Biologicals.







Western Blot: beta-Actin Antibody (AC-15) [NB600-501] - Zinc-finger motifs of ZAP are required for JEV RNA binding & its antiviral activity.(A) Schematic representation of human ZAP isoforms (ZAP-L, 902 a.a. & ZAP-S, 699 a.a.). The four tandem CCCH-type zinc-finger (ZF) motifs within the N-terminus of ZAP are shown as solid black boxes. Deletion of the four ZFs (a.a. 73–193) are indicated with a dashed line. (B) 293T/17 cells infected with JEV (MOI = 5) for 16 h were transfected with plasmids expressing EGFP, WT or ZF-deleted ZAP-L-V5 & ZAP-S-V5 for additional 24 h. The viral RNA bound with V5-tagged proteins was pulled down by anti-V5 agarose affinity gel & amplified by RT-PCR with JEV 3'-UTR specific primers (middle panel). RT-PCR of input viral RNA in JEV-infected cells (lower panel). Western blot analysis of the immunoprecipitated ZAP-L & ZAP-S (WT & del4ZFs) (upper panel). (C-E) The indicated cells were infected with JEV (MOI = 5) for 10 h. Cell lysates, total RNA, & culture supernatants were determined for the indicated proteins by western blot (C), viral RNA by RT-PCR (D), & viral titer by plague assay (E). Representative data from three independent experiments shown as mean \pm SD (n = 3) were analyzed by two-tailed Student's t test. *** P≤0.001. Image collected & cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/30016363), licensed under a CC-BY license. Not internally tested by Novus Biologicals.

Western Blot: beta-Actin Antibody (AC-15) [NB600-501] - Effects of HSM a on IL-1ß gene expression & secretion in human macrophages. Cells were pre-treated with either 1 or 2% of HSM extract for 20 h, followed by treatment with LPS (0.5 µg/ml) for 3 h & with ATP (5 mM) for 1 h. (a) The mRNA expression levels of IL-1 β were determined by RT-PCR analysis. (b) IL-1 β mRNAs were quantified using real-time PCR. β -actin gene expression was used for normalization. The results are expressed as fold changes, considering one as the value of untreated cells. (c) The amount of IL-16 in cell culture supernatants was detected by ELISA. (d) The presence of IL-1β in cell lysates & cell culture supernatants were analyzed by Western blot analysis. Data are presented as means ± SE of three experiments performed in duplicate. #P < 0.01 versus untreated cells. *P < 0.01 versus HSM-untreated (ethanol-treated) control cells. *P < 0.05 versus HSM (1%) treated cells. Image collected & cropped by CiteAb from the following publication

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Western Blot: beta-Actin Antibody (AC-15) [NB600-501] - Intrathecal injection of AM1241 (100pM) attenuated CCI-induced hyperalgesia & increased expression of p-p38MAPK & NF-kappaBp65 in the rat dorsal spinal cord (n = 8 per group). a Western blotting image of p-p38MAPK expression. The top panel was the target band, p-p38MAPK, & the bottom one was for the loading control beta-actin. b Western blotting quantitative analysis of the p-p38MAPK expression in the rat dorsal spinal cord. *P < 0.05 means comparison with the sham group; +P < 0.05 means comparison with the vehicle-treated CCI group. c Western blotting image of NF-kappaBp65 expression. The top panel was the target band, NF-kappaBp65, & the bottom one was for the loading control beta-actin. d Western blotting quantitative analysis of the NFkappaBp65 expression in the dorsal spinal cord of the rats. *P < 0.05means comparison with the sham group; +P < 0.05 means comparison with the vehicle-treated CCI group Image collected & cropped by CiteAb from the following publication

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sham 3d 7d 3d 7d CCI+AM1241 CCI+0.005%DMSO



Western Blot: beta-Actin Antibody (AC-15) [NB600-501] - Intralipid enhances the cytotoxicity of Abraxane in tumors. (A) The box-andwhisker plot represents the amount of paclitaxel in tumors determined by LC/MS. The number above the box denotes the median amount of remaining paclitaxel in the tumor tissues. (B) Western blot analysis of the apoptosis signal, cleaved caspase-3, in the tumors at 24 & 48 hrs after Abraxane treatment. β-actin served as a loading control. The blots of caspase-3 & β-actin are cropped from different parts of the same gel. The intensity of each band of the cleaved caspase-3 was measured by ImageJ software (NIH) & normalized to its own signal of β-actin. The values summarized in the box-and-whisker plot denote the relative intensities compared with those in the control group. (C,D) The immunohistochemistry of cleaved caspase-3 (C) & the TUNEL assay (D) are analyzed in 48 hrs tumor sections. Original: 200x magnification; Inserts; 400x magnification. Image collected & cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/32071352), licensed under a CC-BY license. Not internally tested by Novus Biologicals.

Western Blot: beta-Actin Antibody (AC-15) [NB600-501] - The effect of minocycline on the expression of P2Y12 & P2Y13 receptors. All values represent mean ± standard deviation. a RT-PCR results show the expression of P2Y12 mRNA expression in the rat dorsal spinal cord (n = 8 per group). *P < 0.05 means comparison with the sham group; +P < 0.05 means comparison with the CCI rats. b RT-PCR results show the expression of P2Y13 mRNA expression in the rat dorsal spinal cord (n = 8 per group). *P < 0.05 means comparison with the sham group; +P < 0.05 means comparison with the CCI rats. c Western blotting quantitative analysis of the P2Y12R expression in the rat dorsal spinal cord. *P < 0.05 means comparison with the control; +P < 0.05 means comparison with the ADPbetaS-treated rats. d Western blotting quantitative analysis of the P2Y13R expression in the rat dorsal spinal cord. *P < 0.05 means comparison with the control; +P < 0.05 means comparison with the ADPbetaS-treated rats. e Western blotting image of P2Y12R expression. The top panel was the target band, P2Y12R, & the bottom one was for the loading control beta-actin. f Western blotting image of P2Y13R expression. The top panel was the target band, P2Y13R, & the bottom one was for the loading control beta-actin Image collected & cropped by CiteAb from the following publication (http://jneuroinflammation.biomedcentral.com/articles/10.1186/s12974-017-0960-0), licensed under a CC-BY license. Not internally tested by Novus Biologicals.





Western Blot: beta-Actin Antibody (AC-15) [NB600-501] - Effects of HSM a on inflammasome components & caspase-4 activation in human macrophages. Cells were pretreated with various concentrations (1 or 2%) of HSM extract for 20 h, followed by treatment with LPS ($0.5 \mu g/ml$) for 3 h & ATP (5 mM) for 1 h. (a) The mRNA expression levels of ASC, NLRP1, NLRP3 & caspase-4 were determined by RT-PCR, using β -actin as the internal control. (b) ASC, NLRP1, NLRP3 & caspase-4 mRNAs were quantified using real-time PCR. β -actin gene expression was used for normalization. The results are expressed as fold changes, considering one as the value of untreated cells. (c) Cell lysates were analyzed by Western blot analysis using specific anti-ASC, anti-NLRP1 & anti-NLRP3 antibodies. (d) Cell lysates were analyzed for protein levels of caspase-4 by Western blot analysis. β -actin was used as an internal control. Data are presented as means ± SE of three experiments preformed in duplicate. #P < 0.01 versus untreated cells. versus HSMuntreated control (ethanol) cells. *P < 0.01 versus HSM-untreated control (ethanol) cells. Image collected & cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/23459183), licensed under a CC-BY license. Not internally tested by Novus Biologicals.

Western Blot: beta-Actin Antibody (AC-15) [NB600-501] - The cleaved MFN fragments were further degraded by host proteasome machinery. (A) Western blot analysis of A549+on/MFN1 (lane 1-6) & /MFN2 (lane 7–12) cells with Dox treatment (all lanes) & DENV infection (lane 3–6 & 9-12). The cells were infected with DENV serotype 2 (moi 5) for 24 h, then incubated in culture media containing Dox (1 µg/ml) with (lanes 5–6 & 11–12) or without MG132 (0.5 μM) for 16 h. (B) Western blot analysis of four different serotypes DENV-infected A549 cells incubated with MG132-containing medium. (C) Western blot analysis of endogenous MFN1 & 2 cleavage in A549 stable cells overexpressing WT or proteasedead mutant (S135A) of DENV NS2B3 with or without 18-h treatment with MG132 (0.5 µM). Filled arrow: full-length; open arrow: cleaved product; star: non-specific band. The longer exposure images for endogenous MFN1 & 2 signals are also shown in panels B & C. Image collected & cropped by CiteAb from the following publication (https://dx.plos.org/10.1371/journal.ppat.1005350), licensed under a CC-BY license. Not internally tested by Novus Biologicals.

Western Blot: beta-Actin Antibody (AC-15) [NB600-501] - ENERGI-F704 decreases LPS-induced NO, iNOS, COX-2, & PGE2 production in BV2 cells. BV2 cells were stimulated with 200 ng/mL LPS & subsequently treated with either ENERGI-F704 or AICAR for 24 h. (a) Levels of NO in the culture of BV2 cell were determined. (b) After treatments, cell lysates were used to determine the level of iNOS & COX-2 using Western blotting. (c) LPS-treated BV2 cells were treated with ENERGI-F704 in cotreatment with or without compound C for 24 h. Cell lysates were used to determine the level of iNOS & COX-2 using Western blotting. (d) LPSinduced BV2 cells were treated with ENERGI-F704 in cotreatment with or without compound C for 48 h, & the level of PGE2 in the culture medium was determined. Data are presented as the mean \pm SEM of three independent experiments (two-way ANOVA; *, **, & ***, P < 0.05, P < 0.01, & P < 0.01; specific comparison to LPS-treated control). Representative images of three independent experiments are shown. Image collected & cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/25025067), licensed under a CC-BY license. Not internally tested by Novus Biologicals.





(b)

Simple Western: beta-Actin Antibody (AC-15) [NB600-501] - iPSC-RPE from all donors express RPE marker proteins. (A) As a loading control, each iPSC-RPE line (ARB patient, the patient's parents, & three unrelated, unaffected controls) as well as iPSCs (negative control) were blotted for ß-actin. (B) Unlike iPSCs, all iPSC-RPE lines showed robust expression of CRALBP. (C) Just like CRALBP, iPSC-RPE from the patient, the patient's mom, the patient's dad, & the three unrelated, unaffected controls all expressed RPE65. The negative control (iPSCs) had no detectable RPE65 protein expression. (D) Each of the iPSC-RPE lines but not iPSCs displayed strong expression of the RPE marker Mertk. (E) The relative expression levels for each of these three markers is summarized for iPSCs & all of the iPSC-RPE lines. Graphs show the summary of expression for each marker from three separate blotting experiments. Variability between each of the controls as well as between the different iPSC-RPE lines was observed for each of the RPE markers. Control 4 displayed the highest levels of CRALBP, Control 138 had the highest levels of RPE65, & Control 1 showed the highest levels of Mertk. Image collected & cropped by CiteAb from the following publication (https://www.nature.com/articles/s41598-018-21651-z), licensed under a CC-BY license. Not internally tested by Novus Biologicals.

Western Blot: beta-Actin Antibody (AC-15) [NB600-501] - Effects of MFN1 or MFN2 overexpression on DENV infection.(A) Western blot analysis of A549+on/HA-MFN1 cells induced with Dox (1 µg/ml) or not for 18 h, then infected with DENV serotype 2 for 24 or 48 h by the indicated moi. The relative ratios of band intensity were quantified by ImageJ. p.i. (h): hours post infection. (B) DENV plague-forming assay of culture supernatants from A549+on/HA-MFN1 cells cultured with or without Dox (1 µg/ml) for 18 h, then infected with DENV (moi 0.1 or 10) as indicated. Data in panels B-D & F-G are mean ± SD (n = 3 per group) & were compared by two-tailed Student's t test. (C) RT-qPCR analysis of IFNβ mRNA expression at indicated time point in DENV-infected (moi 10) A549+on/HA-MFN1 cells with or without an 18 h-Dox-pretreatment. (D) Analysis of antiviral activity against dSinF-Luc/2A virus in culture media from A549+on/HA-MFN1 cells with DENV infection (moi 5, 48 h) with or without 18-h Dox pretreatment. (E) Western blot analysis of A549+on/HA-MFN2 cells induced with Dox (1 µg/ml, 18 h) or not, then infected with DENV (moi 10) for the indicated time. (F & G) Analysis of the culture supernatants derived from panel E for DENV titer (F) & LDH release (G). (H) Flow cytometry of A549+on/HA-MFN2 cells double stained with JC-1 & annexin V. Cells were infected with DENV (moi 5) for 48 h with or without 18-h Dox pre-treatment. Decreased red/green fluorescence ratio of JC-1 represents disrupted MMP. Image collected & cropped by CiteAb from the following publication (https://dx.plos.org/10.1371/journal.ppat.1005350), licensed under a CC-

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Procedures

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Immunohistochemistry Protocol for Beta Actin Antibody (NB600-501)
(NB600-501):
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IHC-FFPE sections:

I. Deparaffinization:

A. Treat slides with Xylene: 3 changes for 5 minutes each. Drain slides for 10 seconds between changes.
 B. Treat slides with 100% Reagent Alcohol: 3 changes for 5 minutes each. Drain slides for 10 seconds between changes.

II. Quench Endogenous Peroxidase:

A. Place slides in peroxidase quenching solution: 15-30 minutes.

To Prepare 200 ml of Quenching Solution:

Add 3 ml of 30% Hydrogen Peroxide to 200 ml of Methanol. Use within 4 hours of preparation

B. Place slides in distilled water: 2 changes for 2 minutes each.

III. Retrieve Epitopes:

A. Preheat Citrate Buffer. Place 200 ml of Citrate Buffer Working Solution into container, cover and place into steamer. Heat to 90-96 degrees Celcius.

B. Place rack of slides into hot Citrate Buffer for 20 minutes. Cover.

- C. Carefully remove container with slides from steamer and cool on bench, uncovered, for 20 minutes.
- D. Slowly add distilled water to further cool for 5 minutes.
- E. Rinse slides with distilled water. 2 changes for 2 minutes each.
- IV. Immunostaining Procedure:

A. Remove each slide from rack and circle tissue section with a hydrophobic barrier pen (e.g. Liquid Blocker-Super Pap Pen).

- B. Flood slide with Wash Solution. Do not allow tissue sections to dry for the rest of the procedure.
- C. Drain wash solution and apply 4 drops of Blocking Reagent to each slide and incubate for 15 minutes.

D. Drain Blocking Reagent (do not wash off the Blocking Reagent), apply 200 ul of Primary Antibody solution to each slide, and incubate for 1 hour.

E. Wash slides with Wash Solution: 3 changes for 5 minutes each.

- F. Drain wash solution, apply 4 drops of Secondary antibody to each slide and incubate for 1 hour.
- G. Wash slides with Wash Solution: 3 changes for 5 minutes each.
- H. Drain wash solution, apply 4 drops of DAB Substrate to each slide and develop for 5-10 minutes. Check development with microscope.
- I. Wash slides with Wash Solution: 3 changes for 5 minutes each.

J. Drain wash solution, apply 4 drops of Hematoxylin to each slide and stain for 1-3 minutes. Increase time if darker counterstaining is desired.

- K. Wash slides with Wash Solution: 2-3 changes for 2 minutes each.
- L. Drain wash solution and apply 4 drops of Bluing Solution to each slide for 1-2 minutes.
- M. Rinse slides in distilled water.
- N. Soak slides in 70% reagent alcohol: 3 minutes with intermittent agitation.

O. Soak slides in 95% reagent alcohol: 2 changes for 3 minutes each with intermittent agitation.

P. Soak slides in 100% reagent alcohol: 3 changes for 3 minutes each with intermittent agitation. Drain slides for 10 seconds between each change.

Q. Soak slides in Xylene: 3 changes for 3 minutes each with intermittent agitation. Drain slides for 10 seconds between each change.

R. Apply 2-3 drops of non-aqueous mounting media to each slide and mount coverslip.

S. Lay slides on a flat surface to dry prior to viewing under microscope.

NOTES:



Use treated slides (e.g. HistoBond) to assure adherence of FFPE sections to slide.

Prior to deparaffinization, heat slides overnight in a 60 degrees Celcius oven.

All steps in which Xylene is used should be performed in a fume hood.

For Epitope Retrieval, a microwave or pressure cooker may be substituted for the steamer method. Adjust times as necessary depending on conditions.

For the initial IHC run with a new primary antibody, test tissues with and without Epitope Retrieval. In some instances, Epitope Retrieval may not be necessary.

200 ul is the recommended maximum volume to apply to a slide for full coverage. Using more than 200 ul may allow solutions to wick off the slide and create drying artifacts. For small tissue sections less than 200 ul may be used. 5 minutes of development with DAB Substrate should be sufficient. Do not develop for more than 10 minutes. If 5 minutes of development causes background staining, further dilution of the primary antibody may be necessary. Hematoxylin should produce a light nuclear counterstain so as not to obscure the DAB staining. Counterstain for 1-1.5 minutes for nuclear antigens. Counterstain for 2-3 minutes for cytoplasmic and membranous antigens. If darker counterstaining is desired increase time (up to 10 minutes).





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