Product Datasheet

Histone H3 [Asym-dimethyl Arg8] Antibody - BSA Free NB21-1062

Unit Size: 0.05 mg

Store at 4C short term. Aliquot and store at -20C long term. Avoid freeze-thaw cycles.

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NB21-1062

Application Notes

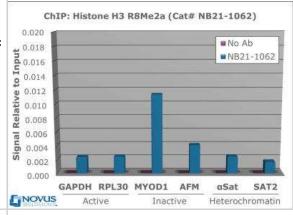
Histone H3 [Asym-dimethyl Arg8] Antibody - BSA Free	
Product Information	
Unit Size	0.05 mg
Concentration	0.84 mg/ml
Storage	Store at 4C short term. Aliquot and store at -20C long term. Avoid freeze-thaw cycles.
Clonality	Polyclonal
Preservative	0.05% Sodium Azide
Isotype	IgG
Purity	Immunogen affinity purified
Buffer	PBS and 30% Glycerol
Target Molecular Weight	15 kDa
Product Description	
Host	Rabbit
Gene ID	126961
Gene Symbol	H3C14
Species	Human, Mouse, C. elegans
Reactivity Notes	Predicted to react with many species including rat, chicken, Xenopus, Drosophila, and plant based on 100% sequence homology.
Immunogen	This Histone H3 [Asym-dimethyl Arg8] antibody was raised against synthetic asymmetric dimethylated peptide surrounding Arginine 8 of human Histone H3.2 [Swiss Prot# Q71DI3].
Notes	Epi-Plus antibody production in collaboration with Rockland Immunochemicals Inc.
Product Application Details	
Applications	Western Blot, Immunocytochemistry/ Immunofluorescence, Immunohistochemistry, Immunohistochemistry-Paraffin, Chromatin Immunoprecipitation (ChIP)
Recommended Dilutions	Western Blot 1:500, Immunohistochemistry, Immunocytochemistry/ Immunofluorescence 1:200, Immunohistochemistry-Paraffin reported in scientific literature (PMID 30382083), Chromatin Immunoprecipitation (ChIP) 2-5 ug per million cells



In Western Blot, a band is seen ~15 kDa in Hela histone prep, NIH 3T3 histone prep, and C. elegans embryo lysate.

Images

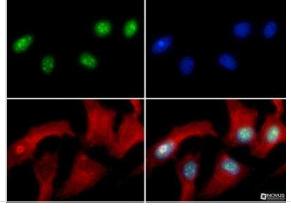
Chromatin Immunoprecipitation: Histone H3 [Asym-dimethyl Arg8] Antibody [NB21-1062] - Chromatin from one million formaldehyde cross-linked HeLa cells was used with 2ug of NB21-1062 and 20ul of magnetic IgG beads per immunoprecipitation. A no antibody (No Ab) control was also used. Immunoprecipitated DNA was quantified using quantitative real-time PCR and SYBR green dye, then normalized to the non-precipitated input chromatin, which is equal to one.



Western Blot: Histone H3 [Asym-dimethyl Arg8] Antibody [NB21-1062] - Analysis of Histone H3R8me2 in: (1) HeLa Histone Prep Lysate, (2) NIH 3T3 Histone Prep Lysate, and (3) C. elegans Embryo Lysate. Observed molecular weight is ~15 kDa.



Immunocytochemistry/Immunofluorescence: Histone H3 [Asym-dimethyl Arg8] Antibody [NB21-1062] - Immunofluorescence: [NB21-1062] - Histone H3.2 R8me2a antibody was tested in HeLa cells with FITC (green). Nuclei and alpha-tubulin were counterstained with DAPI (blue) and DyLight 550 (red).



Publications

Azhar M, Xu C, Jiang X et al. The arginine methyltransferase Prmt1 coordinates the germline arginine methylome essential for spermatogonial homeostasis and male fertility Nucleic acids research 2023-09-22 [PMID: 37739418]

Ka NL, Lim GY, Kim SS et al. Type I IFN stimulates IFI16-mediated aromatase expression in adipocytes that promotes E2-dependent growth of ER-positive breast cancer Cellular and molecular life sciences: CMLS 2022-05-20 [PMID: 35593921]

Details:

Fig. 3

Yang R, Dong S, Zhang J et al. Downregulation of PRMT5 by AMI-1 enhances therapeutic efficacy of compound kushen injection in lung carcinoma in vitro and in vivo Molecular and cellular biochemistry 2022-10-10 [PMID: 36214894]

Dong F, Li Q, Yang C et al. PRMT2 links histone H3R8 asymmetric dimethylation to oncogenic activation and tumorigenesis of glioblastoma. Nat Commun. 2018-10-31 [PMID: 30382083] (IHC-P, ICC/IF, WB, Chemotaxis, Human)

Zhang X, Zhu G, Su X et al. Nucleolar localization signal and histone methylation reader function is required for SPIN1 to promote rRNA gene expression. Biochem. Biophys. Res. Commun. 2018-09-21 [PMID: 30249398] (Chemotaxis, Human)

Zhang B, Dong S, Li Z et al. Targeting protein arginine methyltransferase 5 inhibits human hepatocellular carcinoma growth via the downregulation of beta-catenin. J Transl Med. 2015-11-06 [PMID: 26541651] (WB, Human)

Details:

H3R8me2s / Histone H3 [Asym-dimethyl Arg8] antibody was used for WB analysis of whole-cell lysates that were derived from human hepatocellular carcinoma HepG2 cells transfected with si-NC or si-PRMT5 (Fig. 2f).

Fishman J, Homon A, Lamsa E et al. The Complete Chemical Synthesis of Histones H3 and H4 Containing Epigenetic Modifications and Their Use in Characterizing Arginine Methylated Histone Antibodies. Poster presented at ASCB 21st Century Biochemicals, Marlborough, MA 01752, Novus Biologicals, LLC, 8100 Southpark Way, Littleton, CO 80120.



Procedures

Western Blot protocol for Histone H3 Antibody (NB21-1062)

Histone H3 [Asym-dimethyl Arg8] Antibody:

Western Blot Protocol

- 1. Perform SDS-PAGE on samples to be analyzed, loading 10 ug of histone preps per lane.
- 2. Transfer proteins to membrane according to the instructions provided by the manufacturer of the membrane and transfer apparatus.
- 3. Stain according to standard Ponceau S procedure (or similar product) to assess transfer success, and mark molecular weight standards where appropriate.
- 4. Rinse the blot.
- 5. Block the membrane using standard blocking buffer for at least 1 hour.
- 6. Wash the membrane in wash buffer three times for 10 minutes each.
- 7. Dilute primary antibody in blocking buffer and incubate 1 hour at room temperature.
- 8. Wash the membrane in wash buffer three times for 10 minutes each.
- 9. Apply the diluted HRP conjugated secondary antibody in blocking buffer (as per manufacturers instructions) and incubate 1 hour at room temperature.
- 10. Wash the blot in wash buffer three times for 10 minutes each (this step can be repeated as required to reduce background).
- 11. Apply the detection reagent of choice in accordance with the manufacturers instructions.

Note: Tween-20 can be added to the blocking or antibody dilution buffer at a final concentration of 0.05-0.2%.

Immunocytochemistry Protocol

Culture cells to appropriate density in 35 mm culture dishes or 6-well plates.

- 1. Remove culture medium and add 10% formalin to the dish. Fix at room temperature for 30 minutes.
- 2. Remove the formalin and add ice cold methanol. Incubate for 5-10 minutes.
- 3. Remove methanol and add washing solution (i.e. PBS). Be sure to not let the specimen dry out. Wash three times for 10 minutes.
- 4. To block nonspecific antibody binding incubate in 10% normal goat serum from 1 hour to overnight at room temperature.
- 5. Add primary antibody at appropriate dilution and incubate at room temperature from 2 hours to overnight at room temperature.
- 6. Remove primary antibody and replace with washing solution. Wash three times for 10 minutes.
- 7. Add secondary antibody at appropriate dilution. Incubate for 1 hour at room temperature.
- 8. Remove antibody and replace with wash solution, then wash for 10 minutes. Add Hoechst 33258 to wash solution at 1:25,0000 and incubate for 10 minutes. Wash a third time for 10 minutes.
- 9. Cells can be viewed directly after washing. The plates can also be stored in PBS containing Azide covered in Parafilm (TM). Cells can also be cover-slipped using Fluoromount, with appropriate sealing.

Chromatin Immunoprecipitation Protocol

Cell Fixation and Preparation

- 1. Begin with a cell culture that has reached 80% confluency.
- 2. Add formaldehyde to a final concentration of 1% in growth media and incubate for 10 minutes at room temperature.
- 3. Add glycine to reach a final concentration of 125 mM in the media. Incubate for 5 minutes at room temperature.
- Remove all media and wash twice with 20 mL of ice cold PBS.
- 5. Add 2 mL of ice cold PBS with protease inhibitors*. Scrape cells into microcentrifuge tube.
- 6. Spin cells at 4 C for 5 minutes at 800 x g.
- 7. Remove supernatant and resuspend cells in 750 ul of RIPA lysis buffer containing protease inhibitors* per 10,000,000 cells (enough for 10 IPs). Incubate at 4 C for 15 minutes.
- 8. Spin cells at 4 C for 5 minutes at 800 x g.

DNA Shearing by Sonication



- 1. Sonicate crosslinked DNA to fragments sizes of 200-1000 base pairs. Keep samples ice cold to prevent denaturing of chromatin. Conditions for fragmenting must be empirically derived, and vary depending on equipment, cell type, cell density, and cross-linking efficiency.
- 2. Centrifuge samples to remove debris at 4 C for 10 minutes at 12,500 x g. Remove supernatant and transfer to a new tube. Discard pellet. Set aside 75 ul of sample for input fraction, which will not go through the subsequent IP steps. The remaining sample can be moved into 75 ul aliquots, each of which is sufficient for a single IP. Although it is preferable to proceed directly to the following steps, sheared chromatin can now be frozen at -80 C for up to 1 month. a. Optional: Test the efficiency of the shearing by running 5-10 ul of sample on a 2% agarose gel after reversal of

Chromatin Immunoprecipitation

Recommended controls include: No antibody negative control OR normal IgG negative control, positive control antibody.

crosslinking, RNase treatment (0.5 mg/ml) and proteinase K treatment (0.1 ug/ul) as described below.

- 1. Dilute each IP sample 1:10 by adding 75 ul sheared chromatin to 675 ul dilution buffer, along with appropriate protease inhibitors*. Save undiluted input fraction for step 10.
- 2. Add 25 ul of fully suspended protein A/G magnetic bead slurry. Do not allow the beads to dry.
- 3. Add antibody of interest to the diluted sample. For best results, incubate tubes with rotation at 4 C overnight. Alternatively, incubate at room temperature for 1-2 hours.
- 4. Pellet magnetic beads with a magnetic separator and remove the supernatant.
- 5. Add 750 ul cold low salt buffer and wash for 5 minutes with rotation. Pellet beads with separator and discard supernatant.
- 6. Add 750 ul cold high salt buffer and wash for 5 minutes with rotation. Pellet beads with separator and discard supernatant.
- 7. Add 750 ul cold LiCl buffer and wash for 5 minutes with rotation. Pellet beads with separator and discard supernatant.
- 8. Add 750 ul cold TE buffer and wash for 5 minutes with rotation. Pellet beads with separator and discard supernatant.
- 9. Elute complex by adding 200 ul elution buffer and agitate at RT for 15 minutes. Pellet beads with separator and discard beads. Keep the supernatant.
- 10. Add 8 ul of 5M NaCl and 2 ul of proteinase K (10 ug/ul) to each sample to reverse cross-linking. For the input fraction, add 125 ul of elution buffer along with NaCl and proteinase K. Incubate at 62 C for 4 hours or overnight. Incubate at 95 C for 10 minutes to deactivate proteinase K.
- a. Optional: To decrease time, this step may also be performed at 95 C for 20 minutes without proteinase K treatment.

DNA Purification and Amplification

- 1. DNA can now be purified with either a commercially available column kit or by phenol/chloroform extraction.
- 2. Perform real-time PCR with 2 ul of purified DNA and primers (catalog numbers NBP1-71650, NBP1-71651, NBP1-71652, NBP1-71653, NBP1-71654 and NBP1-71655) per reaction. Dilute input fraction to 1% before PCR. Normalize all IPs and no antibody control IP to adjusted input fraction.
- a. Ex. Raw input CT=30, adjusted input: 30 6.44 = 23.4
- b. Antibody CT=28
- c. Normalized signal relative to input: 2 ^ (23.4-28) = 0.04

Buffers

Dilution Buffer: 0.01% SDS, 1.1% Triton X-100, 1.2 mM EDTA, 16.7 mM Tris-HCL (pH8.1), 167 mM NaCl

Low Salt Wash Buffer: 0.1% SDS, 1% Triton X-100, 2mM EDTA, 20mM Tris-HCl, pH 8.1, 150mM NaCl.

High Salt Wash Buffer: 0.1% SDS, 1% Triton X-100, 2mM EDTA, 20mM Tris-HCl, pH 8.1, 500mM NaCl.

LiCl Wash Buffer: 250mM LiCl, 1% NP-40, 1% Na-deoxycholate, 1mM EDTA, 10mM Tris, pH 8.1.



TE Buffer: 10mM Tris-HCL pH 8.1, 1 mM EDTA

Elution buffer: 1% SDS, 100mM NaHCO3

* Please note that protease inhibitors have variable half-lives and should be freshly prepared as applicable.

**The above information is only intended as a guide. The researcher should determine what protocol best meets their needs. Please follow safe laboratory procedures.





Novus Biologicals USA

10730 E. Briarwood Avenue Centennial, CO 80112

USA

Phone: 303.730.1950 Toll Free: 1.888.506.6887

Fax: 303.730.1966

nb-customerservice@bio-techne.com

Bio-Techne Canada

21 Canmotor Ave Toronto, ON M8Z 4E6

Canada

Phone: 905.827.6400 Toll Free: 855.668.8722 Fax: 905.827.6402

canada.inquires@bio-techne.com

Bio-Techne Ltd

19 Barton Lane Abingdon Science Park Abingdon, OX14 3NB, United Kingdom Phone: (44) (0) 1235 529449

Free Phone: 0800 37 34 15 Fax: (44) (0) 1235 533420 info.EMEA@bio-techne.com

General Contact Information

www.novusbio.com

Technical Support: nb-technical@bio-

techne.com

Orders: nb-customerservice@bio-techne.com

General: novus@novusbio.com

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NB21-1062B Histone H3 [Asym-dimethyl Arg8] Antibody [Biotin]

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