# **Product Datasheet**

# Histone H2AX [p Ser139] Antibody - BSA Free NB100-384

Unit Size: 0.1 ml

Store at 4C. Do not freeze.

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### NB100-384

Histone H2AX [p Ser139] Antibody - BSA Free

Product Information		
Unit Size	0.1 ml	
Concentration	1.0 mg/ml	
Storage	Store at 4C. Do not freeze.	
Clonality	Polyclonal	
Preservative	0.09% Sodium Azide	
Isotype	IgG	
Purity	Immunogen affinity purified	
Buffer	Tris-Citrate/Phosphate (pH 7.0 - 8.0)	
Target Molecular Weight	15 kDa	
Product Description		
Host	Rabbit	
Gene ID	3014	
Gene Symbol	H2AX	
Species	Human, Mouse, Rat, Canine	
Reactivity Notes	Rat reactivity reported in scientific literature (PMID: 27102221), Canine reactivity reported in scientific literature (PMID: 23365434).	
Marker	DNA Double-strand break marker	
Specificity/Sensitivity	The epitope maps to a region surrounding phosphorylated serine 139 of human histone H2AX.	
Immunogen	This Histone H2AX [p Ser139] Antibody was developed against to a region surrounding phosphorylated serine 139 of human histone H2AX [Swiss-Prot entry P16104] (GeneID 3014).	
Notes	Licensed to Novus Biologicals LLC under U.S. Patent Nos. 6,362,317 and 6,884,873.	
Product Application Details		
Applications	Western Blot, Simple Western, Flow Cytometry, Immunocytochemistry/ Immunofluorescence, Immunohistochemistry, Immunohistochemistry-Frozen, Immunohistochemistry-Paraffin, Chromatin Immunoprecipitation (ChIP), Knockout Validated	
Recommended Dilutions	Western Blot 1:10000-1:25000, Simple Western 5 ug/ml, Flow Cytometry 5 ug per 1 million cells, Immunohistochemistry 1:2000 - 1:10000, Immunocytochemistry/ Immunofluorescence 1:500 to 1:5000, Immunohistochemistry-Paraffin 1:2000 - 1:10000, Immunohistochemistry-Frozen 1:1000 - 1:5000, Chromatin Immunoprecipitation (ChIP), Knockout Validated	

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Application Notes	For IHC, epitope retrieval with citrate buffer pH6.0 is recommended for FFPE tissue sections. Formaldehyde fixation is recommended. Permeabilization with Triton-X 100 is recommended for formaldehydefixed cells. Immunoprecipitation is not recommended.
	In Simple Western only 10 - 15 uL of the recommended dilution is used per data point. See <u>Simple Western Antibody Database</u> for Simple Western validation: Tested in Jurkat lysate, separated by Size, antibody dilution of 5 ug/mL, apparent MW was 29 kDa. Separated by Size-Wes, Sally Sue/Peggy Sue. Use in chromatin immunoprecipitation reported in scientific literature (PMID: 30049290).

#### Images





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were treated for 3 hrs in 5ug/ml etoposide, fixed in 1.5% PFA, and



C1:16, C1:17

40 MW (kDa)

anti-KLH or anti-H2AX NB100-384 and secondary FITC-conjugated goat 400 anti-rabbit (in a 150ul reaction). Black- etosposide treated, anti-KLH; Red- untreated, anti-Histone H2AX [p Ser139]; Blue- etoposide treated, 000,1 anti-Histone H2AX [p Ser139]. **Due** 205 FITC-H FFPE section of human ovarian carcinoma. Antibody: Affinity purified rabbit Histone H2AX [p Ser139] antibody used at a dilution of 1:5,000 (0.2 ug/ml). Detection: DAB. Electropherogram image(s) of corresponding Simple Western lane view. Histone H2AX [p Ser139] antibody was used at 5 ug/ml dilution on Jurkat lysate(s).



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Immunohistochemistry: Rabbit Polyclonal Histone H2AX [p Ser139] Antibody - Histone H2AX Antibody on mouse cancer tissue. H2AX (Gray) and H2BGFP(Green). Primary antibody dilution: 1:1000 in a 10um slice.	YHDAX
mTORC1/2 activity prevents Cisplatin-induced cell death in MCF-10A cells. (A) Western blot displaying effects on mTOR signaling during a dose escalation of PP242 treatment in MCF-10A cells; (B) Western blot displaying effects of mTOR signaling on a dose escalation of cisplatin treatment in MCF-10A cells; (C) Western blot displaying effects on mTOR signaling and cell death during non-treated, Cisplatin, PP242, and Cisplatin + PP242-treated MCF-10A cells.	B C S C S C C S C C C C C C C C C C C C
Western Blot: Histone H2AX [p Ser139] Antibody [NB100-384] - ATM & NF- $\kappa$ B activation are downregulated in Ercc1-/ $\Delta$ mice heterozygous for Atm. (A) Livers were collected at 12 weeks of age from WT, Ercc1-/ $\Delta$ & Ercc1-/ $\Delta$ Atm+/- mic (n=3 per genotype) & lysates analyzed by western blot for activation of ATM & its downstream effectors. (B) Same liver lysates were used to measure phosphorylation of p65 & I $\kappa$ Ba. (C) Western blot analysis of livers from 16-week-old WT, Ercc1-/ $\Delta$ & Ercc1-/ $\Delta$ Atm+/-mice (n=3 per genotype) probed for activation of ATM. GAPDH was used as a loading control. (D) Same liver lysates used to measure activation of NF- $\kappa$ B. (E) Fourteen-week-old livers from Ercc1-/ $\Delta$ & Ercc1-/ $\Delta$ P65+/- mice (n=3 per genotype) were analyzed by western blot for activation of ATM (F) & NF- $\kappa$ B. Image collected & cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/32201398), licensed under a CC-BY license. Not internally tested by Novus Biologicals.	M 12 weeks   WT Ercc1 <sup>-/Δ</sup> Ercc1 <sup>-/Δ</sup> Atm <sup>+/-</sup> p-ATM Image: Comparison of the second s
Immunocytochemistry/ Immunofluorescence: Histone H2AX [p Ser139] Antibody [NB100-384] - Immunostaining of γH2AX, WT1 & 5mC in patients with IgA nephropathy & controls. Examples of PAS staining & immunostaining with γH2AX (green) & WT1 (red), pATM & 5mC in glomeruli of IgA nephropathy & controls. (A) A control kidney sample of 44-year-old female, (B) 65-year-old male of IgA nephropathy without podocytopathic features & (C) 55-year-old male of IgA nephropathy with podocytopathic features. Arrows indicate γH2AX & WT1 double-positive cells. Scale bars: 50 µm. Image collected & cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/31937846), licensed under a CC-BY license. Not internally tested by Novus Biologicals.	A PAS staining WT1 (red)+7H2AX (green) pATM SmC





16 weeks Western Blot: Histone H2AX [p Ser139] Antibody [NB100-384] - ATM & С NF- $\kappa$ B activation are downregulated in Ercc1-/ $\Delta$  mice heterozygous for WT Ercc1-/A Ercc1-/Atm+/ Atm. (A) Livers were collected at 12 weeks of age from WT, Ercc1-/ $\Delta$  & p-ATM £  $Ercc1 - \Delta Atm + - mic$  (n=3 per genotype) & lysates analyzed by western blot for activation of ATM & its downstream effectors. (B) Same liver ATM lysates were used to measure phosphorylation of p65 &  $I\kappa B\alpha$ . (C) yH2AX Western blot analysis of livers from 16-week-old WT, Ercc1-/A & Ercc1  $-\Delta Atm+/-mice$  (n=3 per genotype) probed for activation of ATM. GAPDH p21 was used as a loading control. (D) Same liver lysates used to measure GAPDH activation of NF- $\kappa$ B. (E) Fourteen-week-old livers from Ercc1-/ $\Delta$  & Ercc1  $-\Delta p65+/-$  mice (n=3 per genotype) were analyzed by western blot for activation of ATM (F) & NF-kB. Image collected & cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/32201398), licensed under a CC-BY license. Not internally tested by Novus Biologicals. Western Blot: Histone H2AX [p Ser139] Antibody [NB100-384] -D KU Pharmacologic inhibition of ATM rescues oxidative stress-induced p-ATM senescence by suppressing ATM- & NEMO-mediated NF-kB activation. (A) Representative images of primary WT & Ercc1-/- MEFs were induced ATM to undergo senescence by serial passaging at 20% oxygen. At passage CL 5, MEFs were grown in the presence or absence of KU-55933 (10  $\mu$ M) PARP1 for 72 hrs. Senescence was determined by SA-βgal staining. Images were obtained at the magnification of 10x. (B) Quantitation of the percent SA- $\beta$ gal positive cells. Graph represents the mean +/- s.e.m. of three GAPDH independent experiments. Student's t-test, \*\*\*p <0.001, \*\*\*\*p <0.0001. p-KAP1 (C) Passage 5 Ercc1-/- MEFs treated with vehicle or KU-55933 (10 µM) for 72 hours were collected & levels of p21 & p16INK4a were determined by western blotting. (D) Passage 5 Ercc1-/- MEFs were treated with KU-KAP1 NE 55933 (10 µM) for 72 hours & whole cell lysate (CL) & nuclear extracts (NE) were analyzed by immunoblotting for expression of proteins  $\gamma$ H2AX involved in the DNA damage response. (E) Whole cell lysate (CL) & Lamin A/C nuclear extract (NE) were extracted from Ercc1-/- MEFs treated with 10 µM of KU-55933 for analysis of nuclear NEMO & p65. GAPDH was used as a loading control of total proteins & LaminA/C as a loading control of nuclear protein. (F) Passage 5 WT & Ercc1-/- MEFs transfected with a NF-kB-luciferase reporter construct were cultured in the presence or absence of KU-55933 (10 µM) & were collected for luciferase assays after 72 hours. (G) qRT-PCR analysis of mRNA expression in passage 5 WT & Ercc1-/- MEFs treated with or without of KU-55933 (10 µM) for 72 hrs. P values were determined using a Student's t-test. \*p<0.05, \*\*p<0.01, \*\*\*p <0.001. Image collected & cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/32201398), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



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Western Blot: Histone H2AX [p Ser139] Antibody [NB100-384] -Inhibition of DNA-PK prevents monoubiquitination of H2AX & H2A in CPT-treated guiescent WI38 hTERT cells. (A-E) Serum-starved cells were treated with DMSO or DNA-PKi (10 µM) for 1 h before the addition of DMSO (untreated) or CPT (25  $\mu$ M) for 1 h. (A) Western blot of  $\gamma$ H2AX. +Ub1 indicates yH2AX monoubiguitinated. The top panel shows quantification of Ub1-γH2AX normalized to αTubulin (means ± SEM, n = 4). \*\*P < 0.01. (B & C) Cells were pre-extracted with CSK buffer before co-staining for Ub-H2A (red) & 53BP1 phosphorylated on S1778 (p53BP1) (green). (B) Representative pictures. Images were merged to determine colocalization (yellow). The large Ub-H2AX focus at the periphery of the nuclei of untreated & CPT-treated cells may marks the inactive X chromosome as reported (91). (C) Percentages of nuclei with at least 5 Ub-H2A foci (means ± SEM, n = 3, 100 nuclei were analyzed for each treatment in each experiment). \*\*\*P < 0.001. (D & E) Cells were co-stained for ubiquitinated proteins (FK2, red) & yH2AX (green). (D) Representative pictures. Images were merged to determine colocalization (yellow). (E) Number of FK2 foci per nucleus from one representative experiment (76–111 nuclei were analyzed for each treatment) out of three. \*\*\*\*P < 0.0001. In the microscopic images, nuclear contours, identified by DAPI staining (blue in the merge images at bottom), are indicated by dashed lines. Bars: 10 µm. Image collected & cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/26578593), licensed under a CC-BY license. Not internally tested by Novus Biologicals.

Immunocytochemistry/ Immunofluorescence: Histone H2AX [p Ser139] Antibody [NB100-384] - The production of DSBs depends on Top1 degradation in CPT-treated guiescent cells. (A–C) Serum-starved WI38 hTERT cells were co-transfected with siRNAs against cullin 3 & cullin 4B or against a control sequence & then treated with DMSO (-CPT) or 25 µM CPT (+CPT) for 1 h. (A & B) Western blotting of the indicated proteins. aTubulin: loading control. (C) Number of yH2AX foci per nucleus from one representative experiment (246-348 nuclei were analyzed for each treatment) out of three. \*\*\*P < 0.001. (D & E) Serumstarved WI38 hTERT cells were treated with DMSO or MG132 (50 µM) for 1 h before exposure to 0.8 Gy IR. One hour post-irradiation, cells were co-stained for yH2AX (green) & 53BP1 (red). (D) Representative pictures. (E) Number of yH2AX foci per nucleus from one representative experiment (162–180 nuclei were analyzed for each treatment) out of three. Ns: not significant. (F & G) U2OS EV28 cells were treated with DMSO or MG132 (10 µM) for 1 h before the addition of ethanol (untreated) or 300 nM 4-hydroxitamoxifen (4OHT) for 4 h to express AsiSI in the nucleus (42). (F) Representative pictures of cells co-stained for yH2AX (green) & 53BP1 (red). (G) ChIP analysis using an anti-yH2AX antibody (black) or a non-immune antibody (IgG, gray). Enrichment was assessed by QPCR amplification using primers proximal to two AsiSI sites located inside two genes (Gene I: SFRS6, Gene II: CCD47) & primers distal to an AsiSI site (Control). Enrichment was normalized to the maximum recovery for each experiment (means ± SEM, n = 3). Ns: not significant; \*P < 0.05. In the microscopic images, nuclear contours, identified by DAPI staining (not shown), are indicated by dashed lines. Bars: 10 µm. Image collected & cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/26578593), licensed under a CC-BY license. Not internally tested by Novus Biologicals.

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B p-ATM

detection of p-p65 & total p65 in WT & Ercc1-/- MEFs at passage 5 after culturing in 20% oxygen. (G) Representative images of immunofluorescent detection of p65 & NEMO in passage 4 WT & Ercc1 -/- MEFs grown at 20% oxygen. Blue: DAPI staining; Green: p65 (top panel) or NEMO (bottom panel). Images were taken at the magnification of 60x. Image collected & cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/32201398), licensed under a CC-BY license. Not internally tested by Novus Biologicals.





Western Blot: Histone H2AX [p Ser139] Antibody [NB100-384] - DDR & NF-kB are activated concomitantly in senescent MEFs & aged tissues. (A) Immunoblot detection of p-p65 & total p65 in liver tissue from 16week-old WT (n=3) & Ercc1- $/\Delta$  (n=3) mice. (B) Immunoblot detection of phosphorylation of ATM & downstream targets γH2AX & p21 in liver from 16-week-old WT & Ercc1-/ mice. (C) Immunoblot detection of phosphorylation of NF-κB & IκBα in liver lysates from 3, 12 & 24 monthold WT mice. n=3 mice per group. (D) Immunoblot detection of p-ATM, ATM & p21 in the same liver lysates. (E) Immunoblot detection of DDR effectors in nuclear extracts from passage 5 WT & Ercc1-/- MEFs, grown at 20% oxygen. (F) Level of NF-kB activation is higher in Ercc1-/- MEFs compared to WT MEFs at passage 5, as measured by Immunoblot detection of p-p65 & total p65 in WT & Ercc1-/- MEFs at passage 5 after culturing in 20% oxygen. (G) Representative images of immunofluorescent detection of p65 & NEMO in passage 4 WT & Ercc1 -/- MEFs grown at 20% oxygen. Blue: DAPI staining; Green: p65 (top panel) or NEMO (bottom panel). Images were taken at the magnification of 60x. Image collected & cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/32201398), licensed under a CC-BY license. Not internally tested by Novus Biologicals.

Western Blot: Histone H2AX [p Ser139] Antibody [NB100-384] -Induction of ubiquitin/proteasome-dependent DSBs in CPT-treated quiescent WI38 hTERT cells. (A-F) Serum-starved cells were treated with DMSO (1 h) or with MG132 (50  $\mu$ M, 1 h), lactacystin (10  $\mu$ M, 1 h), bortezomib (1 µM, 4 h), G5 (1.5 µM, 0.5 h) or Pyr-41 (9 µM, 0.5 h) before the addition of DMSO (untreated) or 25 µM CPT for 1 h & then costained for yH2AX (green) & 53BP1 (red) or analyzed by Western blot. '-' in panels C & F means cells treated with DMSO. (A & D) Representative pictures. Nuclear contours, identified by DAPI staining (not shown), are indicated by dashed lines. Bars: 10 µm. (B & E) Number of yH2AX foci per nucleus from two independent experiments (147–153 nuclei were analyzed for each treatment). \*\*\*\*P < 0.0001. (C & F) Western blot of yH2AX. αTubulin: loading control. Dashed lines indicate that intervening wells have been spliced out. The top panels show quantification of yH2AX normalized to  $\alpha$ Tubulin (means ± SEM, n = 4 in panel C, n = 3 in panel F). \*\*\*P < 0.001; \*\*P < 0.01. (G & H) Detection of DSBs by neutral Comet assays in serum-starved cells treated with DMSO or MG132 (25  $\mu$ M) for 1 h before the addition of DMSO (untreated) or CPT for 1 h (7.5  $\mu$ M for experiments (Exp) I & II; 5 & 7.5  $\mu$ M for Exp III). (G) Representative pictures of nuclei from Exp I. (H) Quantification of neutral Comet tail moments for three independent experiments (95–133 nuclei were analyzed for each treatment in each experiment). \*\*\*P < 0.001; \*\*\*\*P < 0.0001. The untreated & CPT data from Exp I are from the same experiment as that of Supplementary Figure S3D. Image collected & cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/26578593), licensed under a CC-BY license. Not internally tested by Novus Biologicals.





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#### **Publications**

Li Q, Yang G, Ren B, Liu X et AI. ZC3H14 facilitates backsplicing by binding to exon-intron boundary and 3' UTR Mol Cell 2024-10-26 [PMID: 39461343]

Herrera FG, Ronet C, Ochoa de Olza M, Barras D et Al. Low-Dose Radiotherapy Reverses Tumor Immune Desertification and Resistance to Immunotherapy Cancer Discov 2021-09-04 [PMID: 34479871]

Zeng F, Parker K, Zhan Y et al. Upregulated DNA Damage-Linked Biomarkers in Parkinson's Disease Model Mice ASN Neuro 2023-01-23 [PMID: 36683340]

S Min, HS Lee, JH Ji, Y Heo, Y Kim, S Chae, YW Choi, HC Kang, M Nakanishi, H Cho The chromatin remodeler RSF1 coordinates epigenetic marks for transcriptional repression and DSB repair Nucleic Acids Research, 2021-12-02;49(21):12268-12283. 2021-12-02 [PMID: 34850117]

Chen L, Gai X, Yu X. et Al. Pre-rRNA facilitates the recruitment of RAD51AP1 to DNA double-strand breaks J Biol Chem 2024-02-24 [PMID: 38403248]

Pignanelli C, Ma D, Noel M et al. Selective Targeting of Cancer Cells by Oxidative Vulnerabilities with Novel Curcumin Analogs. Sci Rep 2017-04-24 [PMID: 28439094]

DR Whelan, WTC Lee, F Marks, YT Kong, Y Yin, E Rothenberg Super-resolution visualization of distinct stalled and broken replication fork structures PloS Genetics, 2020-12-28;16(12):e1009256. 2020-12-28 [PMID: 33370257]

Fancy NN, Smith AM, Caramello A et Al. Characterisation of premature cell senescence in Alzheimer's disease using single nuclear transcriptomics Acta Neuropathol 2024-05-02 [PMID: 38695952]

Whelan DR, Rothenberg E., et Al. Super-resolution mapping of cellular double-strand break resection complexes during homologous recombination Proc Natl Acad Sci U S A 2021-03-16 [PMID: 33707212]

Idrees SM, Waite SL, Granados Aparici S et Al. Nicotine exposure is associated with targeted impairments in primordial follicle phenotype in cultured neonatal mouse ovaries Ecotoxicol Environ Saf 2024-11-14 [PMID: 39546863]

Fried W, Tyagi M, Minakhin L et Al. Discovery of a small-molecule inhibitor that traps Pol? on DNA and synergizes with PARP inhibitors Nat Commun 2024-04-05 [PMID: 38580648]

Yu L, Wu D., et Al. SMARCA2 and SMARCA4 Participate in DNA Damage Repair Front Biosci (Landmark Ed) 2024-07-31 [PMID: 39082357]

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#### Products Related to NB100-384

NBP2-24891	Rabbit IgG Isotype Control
NB7160	Goat anti-Rabbit IgG (H+L) Secondary Antibody [HRP]
HAF008	Goat anti-Rabbit IgG Secondary Antibody [HRP]
NBL1-11424	Histone H2AX Overexpression Lysate

#### Limitations

This product is for research use only and is not approved for use in humans or in clinical diagnosis. Primary Antibodies are guaranteed for 1 year from date of receipt.

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