biotechne

Activation and Expansion of Human T Cells

With Bead-Bound GMP CD3 and CD28 Antibodies

Introduction

Adoptive T cell therapy is a rapidly growing segment of **cell and gene therapy**. This protocol outlines how to activate and expand purified Human T cells using R&D Systems CD3 and CD28 antibodies conjugated to beads. T cells are stimulated with GMP CD3 and CD28 antibodies for 2 days followed by 7 days of expansion in Bio-Techne's Xeno-free GMP T cell media. Robust CD3+ T cell expansion and high cell viabilities are achieved in the G-Rex® bioreactor.

ABBREVIATIONS

BSC: Biological Safety Cabinet GMP: Good Manufacturing Practices PBMCs: Peripheral Blood Mononuclear Cells

Materials Required

Material	Catalog Number		
Human GMP T Cell Media	CCM038-GMP		
Human IL-7*	BT-007-GMP / BT-007-AFL		
Human IL-15*	BT-015-GMP / BT-015-AFL		
Human CD3 Antibody	MAB11411-GMP		
Human CD28 Antibody	MAB11412-GMP		
G-Rex® 6 Well Plate M-Series	80240M		
Magnetic Beads Coupled to Streptavidin			
Human AB Serum			
15 and 50 mL Centrifuge Tubes	Multiple vendors		
Biotin Labeling Kit			
Cell Counter			

^{*}Note that IL-7 and IL-15 are available Animal-Free and GMP in lyophilized and liquid formulations.

Timeline

Activation	tivation Dilution	
Day 0	Day 2	Day 6 (optional)
 Prepare media Isolate/thaw T cells Stimulate with CD3/CD28 activator 	 Prepare media to dilute the culture Mix and count cells Fill G-Rex bioreactor 	Media exchangeCheck cell growth
 Incubate (2 days) 	Incubate (7 days)	Day 9
		Harvest cells for desired application

General Guidelines

- · When handling bio-hazardous materials such as human cells, safe laboratory procedures should be followed.
- · Maintain sterile technique, wearing gloves, using nuclease-free reagents, and sterile pipette filter tips.
- All reagents should be stored according to the manufacturer's recommendations.
- If using the recommended Bio-Techne media and G-Rex bioreactor, cells will be confluent after 9 days of culture. If using a different media or reaction vessel, follow manufacturer's protocol to determine optimal culture time.

Protocol

ANTIBODY PREPARATION

PRIOR TO CELL CULTURE

Biotinylation and Immobilization of the Antibodies on Magnetic Beads

- 1. Conjugate CD3 and CD28 antibodies to biotin.
 - a. Follow manufacturer instructions from desired kit.
- 2. Immobilize biotin-conjugated CD3 and CD28 antibodies to magnetic beads coupled to streptavidin.
 - a. Following manufacturer instructions from kits of choice.
 - b. NOTE: For the representative data at the end of this protocol, biotinylated CD3 and CD28 antibodies were immobilized on Dynabeads M-270 Streptavidin (Catalog # 65305) at a 1:3 CD3/CD28 ratio.
- 3. Store at 4 °C until use.

ACTIVATION

DAY 0 OF TOTAL CULTURE TIME

Prepare Media

- 1. Mix and sterile filter complete media.
 - a. Human GMP T Cell Media
 - b. 5% Human AB Serum
 - c. 10 ng/mL IL-7
 - d. 10 ng/mL IL-15
- 2. NOTE: Complete media can be stored at 2 8 °C, protected from light for 2 weeks.

Thaw T Cells

- 3. Either:
 - a. Isolate T cells from human PBMCs following desired protocol or
 - b. Thaw purified T cells and wash with complete media.
- 4. Aspirate supernatant and resuspend cell pellet in complete media.
- 5. Count resuspended cells.
- 6. Seed 0.5 x 10⁶ cells/cm² in 1 mL/cm² of complete media per well in G-Rex plate.
 - a. NOTE: When adding 1 mL/cm² complete media, take into account the volume of activation beads that will be added to the bioreactor.
 - b. Refer to the G-Rex Plating Reference for seeding cell number and vessel sizes.

Wash CD3/CD28 Beads Prior to Use

- 7. Take the desired volume of pre-made CD3/CD28 antibodies conjugated to magnetic beads and place in an appropriately sized tube.
 - a. NOTE: 3 beads per cell are needed for activation.
 - b. EXAMPLE: Prepare beads to activate 5 x 10⁶ cells.
 - I. CD3/CD28 antibody conjugated beads are at 3 x 10⁷ beads per mL.
 - II. 15 x 106 beads are needed for 5 x 106 cells.
 - III. Remove 0.5 mL of CD3/CD28 antibody conjugated beads from stock.
- 8. Place the tube on a magnet for 1 minute.
- 9. Without removing the tube from the magnet, carefully remove the supernatant.
- 10. Remove the tube from the magnet and resuspend in the same volume of complete media as the initial volume taken from the stock.

Stimulate With CD3/CD28 Activator

- 11. Add CD3 and CD28 activation reagent to cells.
- 12. Gently pipet mix to evenly distribute the activators in the culture.
- 13. Transfer G-Rex bioreactor to a humidified incubator (37 °C, 5% CO₂) and incubate for 2 days.

DILUTION

DAY 2 OF TOTAL CULTURE TIME

Prepare Media

1. Pre-warm volume of complete media needed to fill the G-Rex bioreactor.

Fill G-Rex Bioreactor

- 1. Gently mix the activated cell complexes to break them apart, approximately 10-15 times.
 - a. If desired, cell counts can be performed at this step.
 - b. NOTE: We have observed disrupting cell complexes at this time improves cell expansion.
- 2. Fill G-Rex to 4 mL/cm² per well in the non-M series or 10 mL/cm² per well in the M series with pre-warmed complete media.
 - a. Refer to the G-Rex Plating Reference table for fill volumes of other vessel sizes.
- 3. Place G-Rex bioreactor into incubator at 37 °C, 5% CO₂.

EXPANSION

OPTIONAL: DAY 6 OF TOTAL CULTURE TIME

Prepare Media and Perform 1/2 Media Exchange

- 1. Pre-warm volume of complete media needed to complete ½ media exchange in a G-Rex 6 well bioreactor.
 - a. NOTE: If using G-Rex 6M, 100M or 500M, cells and media do not need handling and can be left untouched until day 9 of total culture time.
- 2. Transfer G-Rex bioreactor to BSC.
- 3. Carefully volume reduce wells of G-Rex to ~2 mL/cm².
- 4. Add pre-warmed complete media to wells to fill G-Rex bioreactor.
- 5. Proceed to check cell growth, or place G-Rex bioreactor into incubator at 37 °C, 5% CO₂.

Check Cell Growth (Optional)

- 1. Cell counts and other analysis can be performed at this time.
- 2. Mix cells and sample each well for cell counts and desired analysis.
- 3. Place G-Rex bioreactor into incubator at 37 °C, 5% CO₂ until day 9 of total culture.

DAY 9 OF TOTAL CULTURE TIME

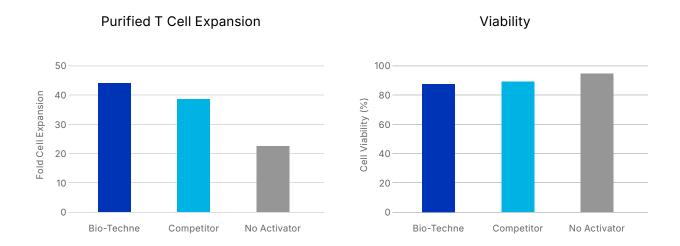
Harvest Cells

- 1. Reduce the volume of the cell culture in G-Rex 6-well to ~2 mL/cm² (about 1/2 of the volume capacity).
- 2. Mix the cells and sample each well for final cell counts and for desired flow cytometry applications for phenotype characterization.
 - a. NOTE: It is recommended to remove the activation beads prior to cell phenotyping, flow cytometry, functional assays and cryopreservation.
- 3. Cryopreserve remaining cells or use directly for desired applications.

G-Rex Plating Reference for Purified T Cells

G-Rex Format	cm²	Cell # for Activation	Activation Volume	Volume Post Activation	Confluency
G-Rex 24 Well Plate	2 cm ²	1 x 10 ⁶	2 mL	8 mL	60-80 x 10 ⁶
G-Rex 6 Well Plate	10 cm ²	5 x 10 ⁶	10 mL	40 mL	350-400 x 10 ⁶
G-Rex 6M Well Plate	10 cm ²	5 x 10 ⁶	10 mL	100 mL	350-400 x 10 ⁶
G-Rex 100M	100 cm ²	50 x 10 ⁶	100 mL	1,000 mL	3.5-4 x 10 ⁹
G-Rex 500M	500 cm ²	250 x 10 ⁶	500 mL	5,000 mL	15-20 x 10 ⁹

Representative Data: CD3/CD28 Antibodies Bound to Beads



BIO-TECHNE'S CD3 AND CD28 ANTIBODIES BOUND TO MAGNETIC BEADS PROMOTE EXPANSION OF PURIFIED T CELLS.

Purified T cells from two independent donors were thawed and activated using streptavidin magnetic beads conjugated with CD3 (Catalog # MAB11411-GMP) and CD28 (Catalog # MAB11412-GMP) antibodies. The cells were cultured for 9 days in 6 well G-Rex bioreactors (Catalog # 80240M) using GMP T Cell Media (Catalog # CCM038-GMP) supplemented with 10 ng/mL liquid GMP IL-7 (Catalog # BT-007-GMP) and IL-15 cytokines (Catalog # BT-015-GMP). Bars represent the average of two donors.

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