



PRODUCT INFORMATION & MANUAL

Human Thrombopoietin (Tpo) Valukine™ ELISA

VAL149

For the quantitative determination of natural and recombinant
Human Tpo concentrations

For research use only.
Not for diagnostic or therapeutic procedures.

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Please refer to the kit label for expiry date.
Novus kits are guaranteed for 3 months from date of receipt

Version202306.2

TABLE OF CONTENTS

I. BACKGROUND	2
II. OVERVIEW	3
III. ADVANTAGES	4
IV. EXPERIMENT	6
V. KIT COMPONENTS AND STORAGE	7
VI. PREPARATION	10
VII. ASSAY PROCEDURE	12
VIII. REFERENCES	14

I. BACKGROUND

Human thrombopoietin (Tpo), also known as megakaryocyte growth and development factor (MGDF), Mpl-ligand, and megakaryocyte colony stimulating factor (MK-CSF), is a 60-70 kDa, 332 amino acid (aa) residue glycosylated polypeptide that is believed to play a key role in the development of megakaryocytes (1-4). Secreted principally by hepatocytes (5) and bone marrow stromal cells (6), the Tpo molecule can be structurally and functionally divided into N-and C-terminal halves. The N-terminal half of Tpo is structurally analogous to the erythropoietin (Epo) molecule, showing approximately 25% aa residue identity and conservation of a number of cysteines. Functionally, this area is suggested to be responsible for receptor binding (1, 2, 7). The C-terminal region, on the other hand, is structurally characterized by a high percentage of serine and proline residues and contains a number of potential N-linked glycosylation sites. The C-terminus is believed to contribute to overall molecule stability and circulating half-life (1, 2, 8). Across species, there is marked conservation of aa residues, with human Tpo demonstrating 75%, 73% and 76% aa identity with rat (9), mouse (2) and dog (2) Tpo, respectively. The conservation of aa structure accounts for the cross-species bioactivity noted on human and mouse cells.

Functionally, Tpo seems to increase the absolute number of megakaryocytes (10), to increase megakaryocyte viability (11), and to shorten the time necessary for cell cycling (10). Tpo does not seem to contribute to pro-platelet formation (11, 12). Circulating Tpo has been measured in both normal and pathophysiological conditions, including idiopathic thrombocytopenia purpura (ITP) (13-15), essential thrombocythemia (ET) (16), liver cirrhosis (17), amegakaryocytic thrombocytopenia (AMT) (18), and aplastic anemia (AA) (13-16). While Tpo levels may be regulated by megakaryocyte counts, and platelets may play a role in regulating plasma Tpo levels, there does not seem to be a direct correlation between platelet count and blood Tpo levels (13, 15, 17-19).

II. OVERVIEW

A. PRINCIPLE OF THE ASSAY

This assay employs the quantitative sandwich enzyme immunoassay technique. An antibody specific for human Tpo has been pre-coated onto a microplate. Standards and samples are pipetted into the wells and any Tpo present is bound by the immobilized antibody. After washing away any unbound substances, a biotin-linked detect antibody specific for human Tpo are pipetted into the wells. After washing away any unbound substances, streptavidin-HRP are pipetted into the wells. Following a wash to remove any unbound reagent, a substrate solution is added to the wells and color develops in proportion to the amount of Tpo bound in the initial step. The color development is stopped, and the intensity of the color is measured.

B. LIMITATIONS OF THE PROCEDURE

- **FOR RESEARCH USE ONLY NOT FOR USE IN DIAGNOSTIC PROCEDURES.**
- This kit is suitable for cell culture supernate and human serum.
- The kit should not be used beyond the expiration date on the kit label.
- Do not mix or substitute reagents with those from other lots or sources.
- If samples generate values higher than the highest standard, dilute the samples with Calibrator Diluent and repeat the assay.
- Any variation in operator, pipetting technique, washing technique, incubation time or temperature, and kit age can cause variation in binding.

III. ADVANTAGES

A. PRECISION

Intra-assay Precision (Precision within an assay)

Three samples were tested twenty times on one plate to assess intra-assay precision.

Inter-assay Precision (Precision between assays)

Three samples were tested in twenty separate assays to assess inter-assay precision.

	Intra-assay Precision			Inter-assay Precision		
Sample	1	2	3	1	2	3
Mean (pg/mL)	226.5	1006.8	5015.6	221.7	1003.7	5070.9
Standard Deviation	8.6	39.2	80.6	11.0	44.6	201.8
CV%	3.8	3.9	1.6	5.0	4.4	4.0

B. RECOVERY

The recovery of human Tpo spiked to different levels throughout the range of the assay in cell culture media was evaluated. The recovery ranged from 96.8-109.4% with an average of 105.5%.

The recovery of human Tpo spiked to different levels throughout the range of the assay in human serum was evaluated. The recovery ranged from 95.3-111.1% with an average of 105.1%.

C. SENSITIVITY

The minimum detectable dose (MDD) of human Tpo is typically less than 5.394 pg/mL.

The MDD was determined by adding two standard deviations to the mean optical density value of twenty zero standard replicates and calculating the corresponding concentration.

D. CALIBRATION

This immunoassay is calibrated against a highly purified Sf21-expressed recombinant human Tpo produced at R&D Systems.

E. LINEARITY

To assess the linearity of the assay, different samples were containing or spiked with high concentrations of human Tpo and diluted with Calibrator Diluent (1×) to produce samples with values within the dynamic range of the assay.

Dilution	Average % of Expected	Range (%)
1:2	94.4	86.8-96.6
1:4	94.9	92.7-101.8
1:8	98.4	86.7-104.1
1:16	88.7	80.9-113.8

F. SAMPLE VALUES

Serum - Five human serum samples were evaluated for the presence of human Tpo in this assay. All samples measured less than the lowest human Tpo standard, 125 pg/mL.

G. SPECIFICITY

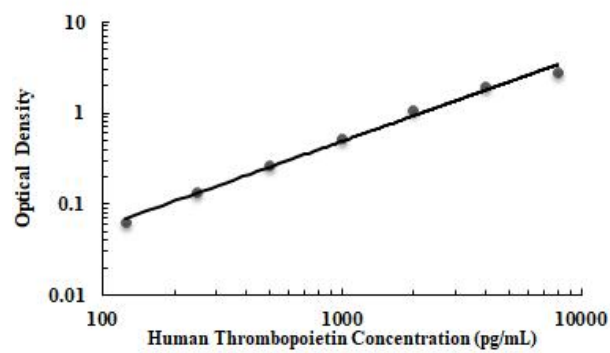
The following factors prepared at 50 ng/mL were assayed and exhibited no cross-reactivity or interference.

Recombinant human	Recombinant mouse
Epo	Epo
Epo R/Fc Chimera	Tpo
Tpo R/Fc Chimera	

IV. EXPERIMENT

EXAMPLE STANDARD

The standard curve is provided for demonstration only. A standard curve should be generated for each set of samples assayed.



pg/mL	OD	Average	Corrected
0	0.079 0.067	0.073	-
125.0	0.134 0.136	0.135	0.062
250.0	0.210 0.207	0.209	0.136
500.0	0.332 0.334	0.333	0.260
1000.0	0.600 0.603	0.602	0.529
2000.0	1.112 1.156	1.134	1.061
4000.0	2.096 1.930	2.013	1.940
8000.0	2.887 2.846	2.867	2.794

V. KIT COMPONENTS AND STORAGE

A. MATERIALS PROVIDED

Parts	Description	Size
Human Tpo Microplate	96 well polystyrene microplate (12 strips of 8 wells) coated with a Mouse antibody against Human Tpo.	1 plate
Human Tpo Standard	Recombinant Human Tpo in a buffered protein base; lyophilized. Refer to the vial label for reconstitution volume.	2 vials
Human Tpo Detection Antibody	Biotinylated Tpo antibody, lyophilized. Refer to the vial label for reconstitution volume.	1 vial
Calibrator Diluent (2×)	Concentrated buffered diluent used to dilute standard and samples.	1 vial
Detection Antibody Diluent (2×)	Concentrated buffered diluent used to dilute Detection Antibody.	1 vial
Streptavidin-HRP A (200×)	200× Streptavidin conjugated to horseradish peroxidase.	1 vial
Reagent Diluent (10×)	A 10× concentrated buffered protein base used to dilute Detection Antibody and HRP.	1 vial
Wash Buffer Concentrate (25×)	A 25× concentrated solution of buffered surfactant with preservatives.	1 vial
TMB Substrate	TMB ELISA Substrate Solution	1 vial
Stop Solution	2 N sulfuric acid.	1 vial
Plate Sealers	Adhesive strip.	3 strips

B. STORAGE

Unopened Kit	Store at 2-8 °C. Do not use past kit expiration date.	
Opened/ Reconstituted Reagents	Streptavidin-HRP A	May be stored for up to 1 month at 2-8 °C.*
	Diluted Wash Solution	
	TMB Substrate	
	Stop Solution	
	Standard	Prepare fresh for each assay.
	Detection Antibody	Aliquot and store for up to 1 month at -20 °C in a manual defrost freezer. *
	Reagent Diluent (10×)	May be stored for up to 1 month at 2-8 °C.* Use and discard diluted Reagent Diluent (1×). Prepare fresh for each assay.
	Calibrator Diluent (2×)	May be stored for up to 1 month at 2-8 °C.* Use and discard diluted Calibrator Diluent (1×). Prepare fresh for each assay.
	Detection Antibody Diluent (2×)	May be stored for up to 1 month at 2-8 °C.* Use and discard diluted Detection antibody Diluent (1×). Prepare fresh for each assay.
	Microplate Wells	Return unused wells to the foil pouch containing the desiccant pack, reseal along entire edge of zip-seal. May be stored for up to 1 month at 2-8 °C.*

* Provided this is within the expiration date of the kit.

C. OTHER SUPPLIES REQUIRED

- ♦ Microplate reader capable of measuring absorbance at 450 nm, with the correction wavelength set at 540 nm or 570 nm.
- ♦ Pipettes and pipette tips.
- ♦ Deionized or distilled water.
- ♦ Squirt bottle, manifold dispenser, or automated microplate washer.
- ♦ Horizontal orbital microplate shaker capable of maintaining a speed of 500±50 rpm.
- ♦ 500 mL graduated cylinder.

D. PRECAUTION

- ♦ Some components in this kit contain a preservative which may cause an allergic skin reaction. Avoid breathing mist.
- ♦ The Stop Solution provided with this kit is an acid solution. Wear protective gloves, clothing, eye, and face protection. Wash hands thoroughly after handling.

VI. PREPARATION

A. SAMPLE COLLECTION AND STORAGE

Cell Culture Supernates - Remove particulates by centrifugation and assay immediately or aliquot and store samples at $\leq -20^{\circ}\text{C}$. Avoid repeated freeze-thaw cycles. Samples may require dilution with Calibrator Diluent (1 $\times$).

Serum - Use a serum separator tube (SST) and allow samples to clot for 30 minutes at room temperature before centrifugation for 15 minutes at 1000 x g. Remove serum and assay immediately or aliquot and store samples at $\leq -20^{\circ}\text{C}$. Avoid repeated freeze-thaw cycles.

B. SAMPLE PREPARATION

Cell Culture Supernates samples require a 2-fold dilution. A suggested 2-fold dilution is 100 μL of sample + 100 μL of Calibrator Diluent (1 $\times$).

Serum samples require a 2-fold dilution. A suggested 2-fold dilution is 100 μL of sample + 100 μL of Calibrator Diluent (1 $\times$).

C. REAGENT PREPARATION

Note: *Bring all reagents to room temperature before use.*

Wash Buffer - If crystals have formed in the concentrate, warm to room temperature and mix gently until the crystals have completely dissolved. Dilute 20 mL of Wash Buffer Concentrate (25 $\times$) into deionized or distilled water to prepare 500 mL of Wash Buffer.

Calibrator Diluent (1 $\times$) - Use deionized or distilled water to prepare Calibrator Diluent (1 $\times$).

Reagent Diluent (1 $\times$) - Use deionized or distilled water to prepare Reagent Diluent (1 $\times$).

Detection Antibody Diluent (1 $\times$) - Use deionized or distilled water to prepare Detection Antibody Diluent (1 $\times$).

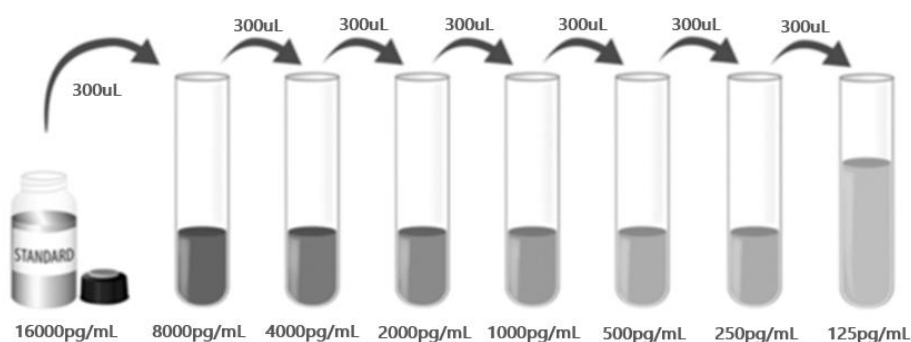
Detection Antibody (1 $\times$) - **Centrifuge briefly before opening. Reconstitution volume refer to vial label to prepare Detection Antibody (100 $\times$).** Allow the Detection Antibody to sit for a minimum of 15 minutes with gentle agitation prior to making dilutions. Aliquot and store if needed. Dilute to Detection Antibody (1 $\times$) with Detection Antibody Diluent (1 $\times$). Prepare at least 15 minutes prior to use.

Streptavidin-HRP A (1 $\times$) - **Centrifuge briefly before opening.** Dilute to the working concentration specified on the vial label using Reagent Diluent (1 $\times$).

Human Tpo Standard - Centrifuge briefly before opening. Refer to the vial label for the reconstitution volume*. This reconstitution produces a stock solution of 16000 pg/mL. Allow the standard to sit for a minimum of 15 minutes with gentle agitation prior to making dilutions.

*if you have any question, please seek help from our Technical Support.

Pipette 300 μ L of the appropriate Calibrator Diluent (1 $\times$) into each tube. Use the stock solution to produce a dilution series (below). Mix each tube thoroughly before the next transfer. The 8000 pg/mL standard serves as the high standard. The Calibrator Diluent (1 $\times$) serves as the zero standard (0 pg/mL).



D. TECHNICAL HINTS

- When mixing or reconstituting protein solutions, always avoid foaming.
- To avoid cross-contamination, change pipette tips between additions of each standard level, between sample additions, and between reagent additions. Also, use separate reservoirs for each reagent.
- It is recommended that the samples be pipetted within 15 minutes.
- To ensure accurate results, proper adhesion of plate sealers during incubation steps is necessary.
- TMB Substrate should remain colorless until added to the plate. Keep Substrate Solution protected from light. Substrate Solution should change from colorless to gradations of blue.
- Stop Solution should be added to the plate in the same order as the Substrate Solution. The color developed in the wells will turn from blue to yellow upon addition of the Stop Solution. Wells that are green in color indicate that the Stop Solution has not mixed thoroughly with the Substrate Solution.

VII. ASSAY PROCEDURE

Note: *Bring all reagents and samples to room temperature before use. It is recommended that all samples and standards be assayed in duplicate.*

1. Prepare all reagents and working standards as directed in the previous sections.
2. Remove excess microplate strips from the plate frame, return them to the foil pouch containing the desiccant pack, and reseal.
3. Add 100 μ L of standard, or prepared sample per well. Cover with the adhesive strip provided. Incubate for 2 hours at room temperature **on a horizontal orbital microplate shaker set at 500 ± 50 rpm**. A plate layout is provided for a record of standards and samples assayed. (Samples may require dilution. See Sample Preparation section.)
4. Aspirate each well and wash, repeating the process three times for a total of four washes. Wash by filling each well with Wash Solution (400 μ L) using a squirt bottle, manifold dispenser, or autowasher. Complete removal of liquid at each step is essential to good performance. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels.
5. Add 100 μ L of the Detection Antibody (1 $\times$) diluted in Detection Antibody Diluent, to each well. Cover with a new adhesive strip and incubate 2 hours at room temperature **on a horizontal orbital microplate shaker set at 500 ± 50 rpm**.
6. Repeat the aspiration/wash as in step 4.
7. Add 100 μ L of the working dilution of Streptavidin-HRP A to each well. Cover the plate and incubate for 30 minutes at room temperature **on a horizontal orbital microplate shaker set at 500 ± 50 rpm. Avoid placing the plate in direct light.**
8. Repeat the aspiration/wash as in step 4.
9. Add 100 μ L of TMB Substrate to each well. Incubate for 20 minutes at room temperature **on a horizontal orbital microplate shaker set at 500 ± 50 rpm. Avoid placing the plate in direct light.**
10. Add 50 μ L of Stop Solution to each well. Gently tap the plate to ensure thorough mixing.
11. Determine the optical density of each well within 10 minutes, using a microplate

reader set to 450 nm. If wavelength correction is available, set to 540 nm or 570 nm. If wavelength correction is not available, subtract readings at 540 nm or 570 nm from the readings at 450 nm. This subtraction will correct for optical imperfections in the plate. Readings made directly at 450 nm without correction may be higher and less accurate.

12. CALCULATION OF RESULTS

Average the duplicate readings for each standard, control, and sample and subtract the average zero standard optical density (O.D.). Create a standard curve by reducing the data using computer software capable of generating a four parameter logistic (4-PL) curve-fit. As an alternative, construct a standard curve by plotting the mean absorbance for each standard on the y-axis against the concentration on the x-axis and draw a best fit curve through the points on the graph. The data may be linearized by plotting the log of the Tpo concentrations versus the log of the O.D. and the best fit line can be determined by regression analysis. This procedure will produce an adequate but less precise fit of the data. If samples have been diluted, the concentration read from the standard curve must be multiplied by the dilution factor.

VIII. REFERENCES

1. De Sauvage, F.J. *et al.* (1994) Nature 369:533.
2. Bartley, T.D. *et al.* (1994) Cell 77:1117.
3. Kaushansky, K. *et al.* (1994) Nature 369:568.
4. Wendling, F. *et al.* (1994) Nature 369:571.
5. Shimada, Y. *et al.* (1995) Exp. Hematol. 23:1388.
6. Nagahisa, H. *et al.* (1996) Blood 87:1309.
7. Sohma, Y. *et al.* (1994) FEBS Lett. 353:57.
8. Lok, S. *et al.* (1994) Nature 369:565.
9. Ogami, K. *et al.* (1995) Gene 158:309.
10. Angchaisuksiri, P. *et al.* (1996) Br. J. Haematol. 93:13.
11. Ito, T. *et al.* (1996) Br. J. Haematol. 94:387.
12. Choi, E.S. *et al.* (1996) Br. J. Haematol. 95:227.
13. Kosugi, S. *et al.* (1996) Br. J. Haematol. 93:704.
14. Kunishima, S. *et al.* (1996) Eur. J. Hematol. 57:68.
15. Ichikawa, N. *et al.* (1996) Thromb. Haemost. 76:156.
16. Tahara, T. *et al.* (1996) Br. J. Haematol. 93:783.
17. Shimodaira, S. *et al.* (1996) Thromb. Haemost. 76:545.
18. Mukai, H.Y. *et al.* (1996) Thromb. Haemost. 76:675.
19. Fielder, P.J. *et al.* (1996) Blood 87:2154.

PLATE LAYOUT

Use this plate layout to record standards and samples assayed.

	A	B	C	D	E	F	G	H
1								
2								
3								
4								
5								
6								
7								
8								
9								
10								
11								
12								



产品信息及操作手册

人 Thrombopoietin (Tpo) Valukine™ ELISA 试剂盒

目录号: **VAL149**

适用于定量检测天然和重组人 Tpo 的浓度

科研专用, 不可用于临床诊断

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目录

I. 背景	18
II. 概述	19
III. 优势	20
IV. 实验	22
V. 试剂盒组成及储存	23
VI. 实验前准备	25
VII. 操作步骤	27
VIII. 参考文献	28

I. 背景

人血小板生成素(Tpo), 又被称为巨核细胞生长和发育因子(MGDF), Mpl配体或巨核细胞集落刺激因子(MK-CSF), 是一种60-70 kDa大小, 含332个氨基酸 (aa)残基的糖基化多肽, 被认为在巨核细胞的发育中发挥关键作用(1-4)。Tpo分子主要由肝细胞 (5)和骨髓基质细胞(6)分泌, 在结构和功能上可以分为N端和C端两部分。Tpo的N端在结构上类似于促红细胞生成素(Epo)分子, 显示约25%的氨基酸残基同源性和部分半胱氨酸保守性。在功能上, 这个区域被认为是负责与受体结合(1, 2, 7)。另一方面, C-末端区域的结构特征为丝氨酸和脯氨酸残基的高百分比, 并包含许多潜在的N-连接的糖基化位点。C末端被认为有助于构建整体分子稳定性和延长循环半衰期(1, 2, 8)。在不同物种中, 氨基酸残基具有明显的保守性, 人Tpo与大鼠(9), 小鼠(2)和狗(2)Tpo分别具有75%, 73%和76%的氨基酸同源性。氨基酸结构上的保守性解释了人类和小鼠细胞的跨物种生物活性。

在功能上, Tpo似乎增加了巨核细胞的绝对数量(10), 增加了巨核细胞活力(11), 并缩短了细胞循环所需的时间(10)。Tpo似乎并不能促进前体血小板生成(11, 12)。已在正常和病理生理条件下测量了循环Tpo, 包括特发性血小板减少性紫癜(ITP) (13-15), 原发性血小板增多症(ET) (16), 肝硬化(17), 无巨核细胞血小板减少症(AMT) (18)和再生障碍性贫血(AA) (13-16)。尽管Tpo水平可能受巨核细胞数量的调节, 血小板可能在调节血浆Tpo水平中发挥作用, 但血小板计数与血液Tpo水平之间似乎没有直接的相关性(13, 15, 17-19)。

II. 概述

A. 检测原理

本实验采用双抗体夹心ELISA法。抗人Tpo抗体包被于微孔板上，样品和标准品中的人Tpo会与固定在板上的抗体结合，游离的成分被洗去；接着加入生物素化的抗人Tpo检测抗体进行孵育，洗涤去除未结合的物质后，加入链霉亲和素标记的辣根过氧化物酶(streptavidin-HRP) 孵育。洗涤未结合试剂后，加入TMB底物溶液。溶液颜色与结合的目标蛋白成正比；加入终止液；用酶标仪测定吸光度。

B. 检测局限

- ◆ 仅供科研使用，不可用于体外诊断；
- ◆ 该试剂盒适用于细胞培养上清样本和人血清样本；
- ◆ 请在试剂盒有效期内使用；
- ◆ 不同试剂盒及不同批号试剂盒的组分不能混用；
- ◆ 样本值若大于标准曲线的最高值，应将样本用标准品稀释液（1×）稀释后重新检测；
- ◆ 检测结果的不同可由多种因素引起，包括实验人员的操作、移液器的使用方式、洗板技术、反应时间或温度、试剂盒的储存等。

III. 优势

A. 精确度

板内精确度（同一板内不同孔间的精确度）

已知浓度的三个样本，在同一板内分别检测20次，以确定板内精确度。

板间精确度（不同板之间的精确度）

已知浓度的三个样本，在不同板中分别检测20次，以确定板间精确度。

	板内精确度			板间精确度		
样本	1	2	3	1	2	3
平均值 (pg/mL)	226.5	1006.8	5015.6	221.7	1003.7	5070.9
标准差	8.6	39.2	80.6	11.0	44.6	201.8
CV%	3.8	3.9	1.6	5.0	4.4	4.0

B. 回收率

在细胞上清样本中掺入检测范围内不同水平的人Tpo，测定其回收率。回收率范围在96.8-109.4%，平均回收率在105.5%。

在人血清样本中掺入检测范围内不同水平的人Tpo，测定其回收率。回收率范围在95.3-111.1%，平均回收率在105.1%。

C. 灵敏度

人Tpo的最低可测剂量（MDD）一般小于5.394pg/mL。

MDD是根据20个重复的零标准品孔的吸光度值的平均值加两倍标准差计算得到的相对应浓度。

D. 校正

此ELISA试剂盒经由R&D Systems生产的Sf21表达的高纯度重组人Tpo蛋白所校正。

E. 线性

不同的样本中含有或掺入高浓度的人Tpo，然后用标准品稀释液（1×）将样本稀释到检测范围内，测定其线性。

稀释倍数	平均值/期待值 (%)	范围 (%)
1:2	94.4	86.8-96.6
1:4	94.9	92.7-101.8
1:8	98.4	86.7-104.1
1:16	88.7	80.9-113.8

F. 样本预值

血清样本 - 使用本试剂盒检测了5份人血清样本中Tpo的水平。5份检测值均低于样本检测限125pg/mL。

G. 特异性

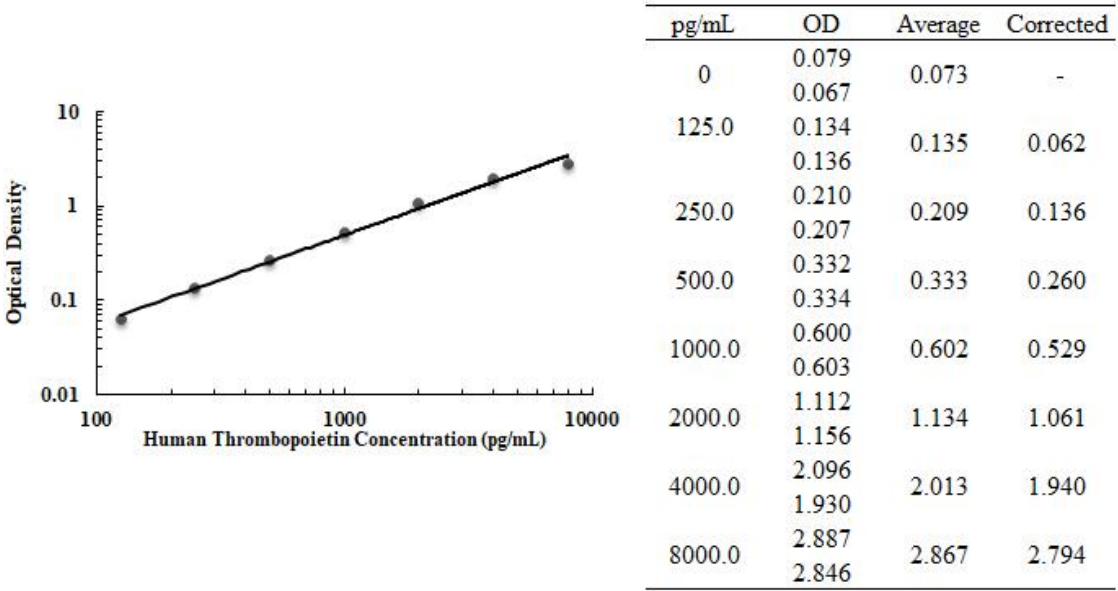
将以下因子配置成50ng/mL的浓度来检测没有观察到明显的交叉反应或干扰。

Recombinant human	Recombinant mouse
Epo	Epo
Epo R/Fc Chimera	Tpo
Tpo R/Fc Chimera	

IV. 实验

标准曲线实例

该标准曲线数据仅供参考，每次实验应绘制其对应的标准曲线。



V. 试剂盒组成及储存

A. 试剂盒组成

组成	描述	规格
Human Tpo Microplate	包被小鼠抗人 Tpo 抗体的 96 孔聚苯乙烯板，8 孔×12 条	1 块板
Human Tpo Standard	标准品（冻干粉），参考瓶身标签进行重溶	2 瓶
Human Tpo Detection antibody	生物素化的 Tpo 检测抗体，冻干粉，参考瓶身标签进行重溶	1 瓶
Calibrator Diluent (2×)	浓缩的标准品稀释液（2×）	1 瓶
Detection Antibody Diluent (2×)	浓缩的检测抗体稀释液（2×）	1 瓶
Streptavidin-HRP A (200×)	200×浓缩的链霉亲和素标记的 HRP	1 瓶
Reagent Diluent (10×)	浓缩的试剂稀释液（10×）	1 瓶
Wash Buffer Concentrate (25×)	浓缩洗涤缓冲液（25×）	1 瓶
TMB Substrate	TMB 底物溶液	1 瓶
Stop Solution	终止液	1 瓶
Plate Sealers	封板膜	3 张

B. 试剂盒储存

未开封试剂盒	2-8℃储存；请在试剂盒有效期内使用	
已打开，稀释 或重溶的试剂	链霉亲和素-HRP A	2-8℃储存，最多 30 天*
	洗涤缓冲液（1×）	
	TMB 底物溶液	
	终止液	
	标准品	使用时新鲜配制*
	检测抗体	分装， -20℃储存，最多 30 天*
	标准品稀释液（2×）	2-8℃储存，最多 30 天* 请每次使用新鲜配制的 1×标准品稀释液
	试剂稀释液（10×）	2-8℃储存，最多 30 天* 请每次使用新鲜配制的 1×试剂稀释液
	检测抗体稀释液（2×）	2-8℃储存，最多 30 天* 请每次使用新鲜配制的 1×检测抗体稀释液
	包被的微孔板条	将未用的板条放回带有干燥剂的铝箔袋内，密封：2-8℃储存，最多 30 天*

*必须在试剂盒有效期内

C. 实验所需自备试验器材

- ◆ 酶标仪（可测量450nm检测波长的吸收值及540nm或570nm校正波长的吸收值）
- ◆ 高精度加液器及一次性吸头
- ◆ 蒸馏水或去离子水
- ◆ 洗瓶（喷瓶）、多通道洗板器或自动洗板机
- ◆ 振荡器（速度可调至500±50rpm）
- ◆ 500mL量筒

D. 注意事项

- ◆ 试剂盒中的一些组分含有防腐剂，可能引起皮肤过敏反应，避免吸入。
- ◆ 试剂盒中的终止液是酸性溶液，使用时请做好眼睛、手、面部及衣服的防护。使用后请彻底洗手。

VI. 实验前准备

A. 样品收集及储存

细胞培养上清液：颗粒物应离心去除；立刻检测样本。样本收集后若不及时检测，需按一次使用量分装，冻存于-20℃冰箱内，避免反复冻融。样本可能需要用标准品稀释液（1×）稀释。

血清样本：用血清分离管(SST)分离血清。使血样室温凝集30分钟，然后1000 x g离心15分钟。吸取血清样本之后即刻用于检测，或者分装，-20℃贮存备用。避免反复冻融。

B. 样本准备工作

细胞上清样本需要用标准品稀释液（1×）2倍稀释后进行检测，取100 μL样本加到100 μL标准品稀释液（1×）中，充分混匀，即制备成2倍稀释的样本。

人血清样本需要用标准品稀释液（1×）2倍稀释后进行检测，取100 μL样本加到100 μL标准品稀释液（1×）中，充分混匀，即制备成2倍稀释的样本。

C. 检测前准备工作

使用前请将所有试剂放置于室温

洗涤液：从冰箱中取出的浓缩洗涤液可能有结晶，属于正常现象；放置室温，轻摇混匀，待结晶完全溶解后再配制洗涤液。可将20mL浓缩洗涤缓冲液（25×）用蒸馏水或去离子水稀释配置成500mL工作浓度的洗涤液。未用完的放回4℃。

标准品稀释液（1×）：使用蒸馏水或去离子水稀释配置成标准品稀释液（1×）。

试剂稀释液（1×）：使用蒸馏水或去离子水稀释配置成试剂稀释液（1×）。

检测抗体稀释液（1×）：使用蒸馏水或去离子水稀释配置成检测抗体稀释液（1×）。

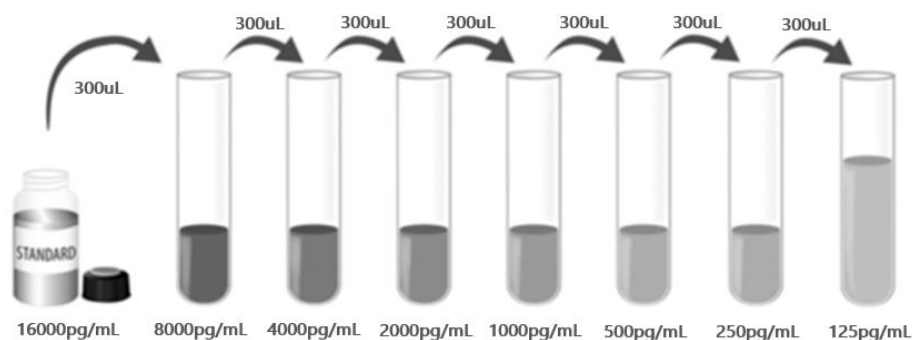
检测抗体（1×）：开盖前请瞬时离心。参考检测抗体瓶标签重溶冻干粉，制备检测抗体（100×）。轻轻震荡至少15分钟，其充分溶解。如有需要分装保存。用检测抗体稀释液（1×）稀释至检测抗体（1×），至少在使用前15分钟准备。

链霉亲和素- HRP A：开盖前请瞬时离心。用试剂稀释液（1×）将链霉亲和素- HRP A（200×）稀释至工作浓度链霉亲和素- HRP A（1×）。

Tpo标准品：开盖前请瞬时离心。冻干标准品的重溶体积请参考瓶身标签，得到浓度为16000pg/mL标准品母液。轻轻震荡至少15分钟，使其充分溶解。

*如有疑问，请咨询我们的技术支持。

向各稀释管中加入 300 μ L标准品稀释液（1 $\times$ ）。将标准品母液参照下图做系列稀释，每管须充分混匀后再移液到下一管。8000pg/mL管作标准曲线最高点，标准品稀释液（1 $\times$ ）可用作标准品零点（0 pg/mL）。



D. 技术小提示

- ◆ 当混合或重溶蛋白液时，尽量避免起沫；
- ◆ 为了避免交叉污染，配置不同浓度标准品、上样、加不同试剂都需要更换枪头。另外不同试剂请分别使用不同的移液槽；
- ◆ 建议15分钟内完成一块板的上样；
- ◆ 每次孵育时，正确使用封板胶纸可保证结果的准确性；
- ◆ TMB底物溶液在上板前应为无色，请避光保存；加入微孔板后，将由无色变成不同深度的蓝色；
- ◆ 终止液上板顺序应同TMB底物溶液上板顺序一致；加入终止液后，孔内颜色由蓝变黄； 若孔内有绿色，则表明孔内液体未混匀，请充分混合。

VII. 操作步骤

使用前请将所有试剂和样本放置于室温，建议所有的实验样本和标准品做复孔检测

1. 按照上一节的说明，准备好所有需要的试剂和标准品；
2. 从已平衡至室温的密封袋中取出微孔板，未用的板条请放回铝箔袋内，重新封口；
3. 分别将不同浓度标准品，实验样本或者质控品加入相应孔中，每孔100 μ L。用封板膜封住反应孔，**室温500 $\pm$ 50rpm水平振荡孵育2小时**。说明书提供了一张96孔模板图，可用于记录标准品和试验样本的板内位置；（样本需要稀释，详情参见样本制备部分。）
4. 将板内液体吸去，使用洗瓶、多通道洗板器或自动洗板机洗板。每孔加洗涤液400 μ L，然后将板内洗涤液吸去。重复操作3次，共洗4次。每次洗板尽量吸去残留液体会有助于得到好的实验结果。最后一次洗板结束，请将板内所有液体吸干或将板倒置，在吸水纸拍干所有残留液体；
5. 在每个微孔内加入100 μ L配置好的检测抗体（1 $\times$ ）。用封板胶纸封住反应孔，**室温500 $\pm$ 50rpm水平振荡孵育2小时**；
6. 重复第4步洗板操作；
7. 在每个微孔内加入100 μ L稀释好的链霉亲和素- HRP A工作液。用封板膜封住反应孔，**室温500 $\pm$ 50rpm水平振荡孵育30分钟，注意避光**；
8. 重复第4步洗板操作；
9. 在每个微孔内加入100 μ L TMB底物溶液，**室温500 $\pm$ 50rpm水平振荡孵育20分钟，注意避光**；
10. 在每个微孔内加入50 μ L终止液，请轻拍微孔板，使溶液混合均匀。孔内溶液颜色会从蓝色变为黄色；
11. 加入终止液后10分钟内，使用酶标仪测量450nm的吸光度值，设定540nm或570nm作为校正波长。如果没有使用双波长校正，结果准确度可能会受影响；
12. **计算结果**：将每个标准品和样品的校正吸光度值（OD₄₅₀-OD₅₄₀/OD₅₇₀），复孔读数取平均值，然后减去平均零标准品OD值。使用计算机软件作四参数逻辑（4-PL）曲线拟合创建标准曲线。另一种方法是，可以通过绘制标准品浓度做对数与相应OD值对数生成曲线，并通过回归分析确定最佳拟合线。通过样本的OD值，可从标准曲线上得到样本中人Tpo的浓度。如果样品被稀释，从标准曲线读取的浓度必须乘以稀释倍数。

VIII. 参考文献

1. De Sauvage, F.J. *et al.* (1994) *Nature* 369:533.
2. Bartley, T.D. *et al.* (1994) *Cell* 77:1117.
3. Kaushansky, K. *et al.* (1994) *Nature* 369:568.
4. Wendling, F. *et al.* (1994) *Nature* 369:571.
5. Shimada, Y. *et al.* (1995) *Exp. Hematol.* 23:1388.
6. Nagahisa, H. *et al.* (1996) *Blood* 87:1309.
7. Sohma, Y. *et al.* (1994) *FEBS Lett.* 353:57.
8. Lok, S. *et al.* (1994) *Nature* 369:565.
9. Ogami, K. *et al.* (1995) *Gene* 158:309.
10. Angchaisuksiri, P. *et al.* (1996) *Br. J. Haematol.* 93:13.
11. Ito, T. *et al.* (1996) *Br. J. Haematol.* 94:387.
12. Choi, E.S. *et al.* (1996) *Br. J. Haematol.* 95:227.
13. Kosugi, S. *et al.* (1996) *Br. J. Haematol.* 93:704.
14. Kunishima, S. *et al.* (1996) *Eur. J. Hematol.* 57:68.
15. Ichikawa, N. *et al.* (1996) *Thromb. Haemost.* 76:156.
16. Tahara, T. *et al.* (1996) *Br. J. Haematol.* 93:783.
17. Shimodaira, S. *et al.* (1996) *Thromb. Haemost.* 76:545.
18. Mukai, H.Y. *et al.* (1996) *Thromb. Haemost.* 76:675.
19. Fielder, P.J. *et al.* (1996) *Blood* 87:2154.

96 孔模板图

请使用 96 孔模板图来记录标准品及样本在板内的位置

