NanoPro[™] Assay: MEK I

SUMMARY

Primary Antibody: Anti-MEK1 (Millipore, cat# 07-641), Anti-phospho MEK1 (Thr292) (Millipore, cat# 07-852) and Anti-phospho MEK1 (Thr386) (Phospho Solution, cat# p180-386) Detection Antibody: Anti-Rabbit HRP (Cell Biosciences, p/n 040-656)

Dual-specificity mitogen-activated protein kinases (MEK) are members of the dual-specificity protein kinase family, which act upstream from the classical MAP kinases through phosphorylation and thus activation of ERK1 and ERK2 in response to a wide variety of extra- and intracellular signals. While the functions of MEK1 and MEK2 are very similar, these kinases differ significantly in the way they are regulated. For example, serum addition can specifically induce MEK1 activity in CHO cells. By contrast, MEK2 appears to be the functionally predominant isoform in formyl-methionyl-leucyl-phenylalanine treated neutrophils. Here we show MEK1 activation in MCF10A cells treated with EGF.

RESULTS phospho MEK I **MEKI** anti-MEKI (total) -EGF +EGF 2,000 1,500 1,000 500 0 5.8 6.4 6.6 6.8 5.4 .6 6 6.2 600 anti-phospho MEKI (Thr292) Chemiluminescence 500 -EGF +EGF 400 300 200 100 0 5.8 6.8 5.4 6 6.4 6.6 6 anti-phospho 500 MEK1 (Thr386) -EGF 400 +EGF 300 200 100 0 6.4 5.4 5.6 5.8 6 6.2 6.6 6.8 pl

EGF stimulation results in increased MEK1 phosphorylation in MCF10A cells

MCF10A cells were treated -/+ EGF (600 ng/mL, 10 minutes). Several phospho MEK1 peaks (green box) increased with EGF treatment, as detected by an anti-phospho MEK1 antibody (bottom two traces) and anti-total MEK1 antibody (top trace). The non-phospho peaks at pl 6.25 decreased dramatically after EGF treatment (orange box).

NOTE: Detection of the chemiluminescent signal produced is relative. Absolute units may vary depending on cell line, treatment and assay conditions

APPLICATION BRIEF No. 1016

Cell **P**Biosciences

PROTOCOL

Cell Preparation

Cell culture:	MCF10A cells (ATCC, cat# CRL-10317) were cultured in MEGM (Lonza, cat# CC3151) containing 10% FBS (Hyclone, cat# 1677-006), 1x Penicillin/Streptomycin/Glutamine (JRS Scientific, cat# 20020), and MEGM SingleQuots (Lonza, cat# CC4136).
	Cells were split 1:5 every 3 days using 0.25% Trypsin (Cellgro, cat# 25-053-Cl) at 37 °C for 3–5 minutes. Data shown from cells
	at passage 5.
Pre-treatment:	Before EGF stimulation, cells were placed at 37 °C, 5% CO $_2$ overnight in starvation medium containing MEGM.
Treatment:	600 ng/mL EGF (Sigma, cat# E1257) in MEGM for 10 minutes at 37 °C, 5% CO2.
Lysis buffer:	Bicine/CHAPS Lysis Buffer (Cell Biosciences, p/n 040-764) plus 1x DMSO Inhibitor Mix (Cell Biosciences, p/n 040-510) and
	I × Aqueous Inhibitor Mix (Cell Biosciences, p/n 040-482).
Lysis details:	Wash cells with 10 mL of ice-cold PBS (Cellgro, cat# 21-031-CV), aspirate well. Add 400 µL ice-cold lysis buffer to 10-cm plate on ice,
	swirl around to ensure good coverage, and incubate 10 minutes on ice. Scrape plate, pipet up and down to mix. Transfer lysate to
	microfuge tube, lyse for an additional 30 minutes on ice. Clarify by centrifugation (14,000 \times g, 15 minutes) in a cooled centrifuge.
	Transfer supernatant to a fresh microfuge tube. Immediately aliquot supernatant (10–30 µL) on ice and snap freeze on dry ice.
Storage:	-80 °C

Assay Reagents

NOTE: For specifics on sample preparation, please consult the addendum to this document.

Protein concentration	: 0.05 mg/mL final in capillary by BCA assay
Sample diluent:	Bicine/CHAPS Lysis Buffer plus 1 × DMSO Inhibitor Mix
Ampholyte premix:	Premix 5-8 (Cell Biosciences Premix G I, p/n 040-327 or Premix G2, p/n 040-973)
pl standards:	pl Standard Ladder 3 (Cell Biosciences, p/n 040-646)
Wash:	Wash Buffer (Cell Biosciences, p/n 040-654)
Primary antibody:	Anti-MEKI (Millipore, cat# 07-641), 1:100, anti-phospho MEKI (Thr292, Millipore, cat# 07-852), 1:50,
	anti-phospho MEKI (Thr386) (Phospho Solution, cat# p180-386), 1:50 and anti-phospho MEKI (S298)
	(Cell Signaling Technology, cat# CS9128), 1:50 in Antibody Diluent (Cell Biosciences, p/n 040-309)
Detection antibody:	Anti-Rabbit HRP (Cell Biosciences, p/n 040-656), 1:100 in Antibody Diluent
Anolyte:	Phosphoric Acid, 10 mM (Cell Biosciences, p/n 040-650)
Catholyte:	Sodium Hydroxide, 100 mM (Cell Biosciences, p/n 040-651)
Luminol/Peroxide:	Mixed I:I (Cell Biosciences, p/n 040-652 and p/n 040-653)

NOTE: NanoPro XDR Assays are now available with optional reagents to provide optimized assay sensitivity, extended dynamic range and both low and high end signal enrichment. Additionally, Premix G2 provides enhanced protein resolution. For more information, visit cellbiosciences.com, contact your Cell Biosciences' Field Applications Specialist or call Technical Support at (888) 607-9692.

Assay Conditions

System:	NanoPro 1000			
Sample loading time:	le loading time: 10 seconds (Premix G1), 25 seconds (Premix G2)			
Focus conditions:	15000 μW, 40 minutes (Premix GI) or 210	00 μW, 40 minutes (Premix G2)		
Immobilization:	80 seconds			
Wash I:	2×150 seconds (default)			
Primary antibody incubation:	60 minutes			
Wash 2:	2×150 seconds (default)			
Detection antibody incubation:	60 minutes			
Wash 3:	2×150 seconds (default)			
Chemiluminescence exposure:	60, 120, and 240 seconds	Our favorite antibodie		

Our favorite antibodies Anti-MEKI (Millipore, cat# 07-641) Anti-phospho MEKI (Thr292) (Millipore, cat# 07-852) Anti-phospho MEKI (Thr386) (Phospho Solution, cat# p180-386) Anti-phospho MEKI (S298) (Cell Signaling Technology, cat# CS9128)

Other antibody suggestions

Anti-phospho MEKI (Abcam, cat# 32088)

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SAMPLE PREPARATION ADDENDUM

Sample preparation procedures are different depending on the premix used. The following updated instructions provide guidance on sample preparation for both Premix G1 and Premix G2.

Step	Premix GI Procedure	Premix G2 Procedure
Step I	Dilute lysate with sample diluents to 0.1 mg/mL.	Dilute lysate with sample diluents to 0.2 mg/mL.
Step 2	In a separate tube, mix Premix GI and pI standards.	In a separate tube, mix Premix G2 and pl standards.
Step 3	Mix equal parts of diluted lysate prepared in Step I with the Premix GI + pl Standards prepared in Step 2 (1:1 ratio) to create final protein concentration of 0.05 mg/mL.	Mix I part diluted lysate prepared in Step I with 3 parts Premix G2 + pl Standards prepared in Step 2 (1:3 ratio) to create final protein concentration of 0.05 mg/mL.

NOTES: When working with Premix G2, thorough mixing and vortexing during sample preparation is required. Additional sample volume may be required, 12-20 uL per sample well is recommended. Centrifugation of the sample plate (3000 x g, 10 minutes) is required.

For further assistance, please contact your Cell Biosciences' Field Applications Specialist or Technical Support at (888) 607-9692.

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