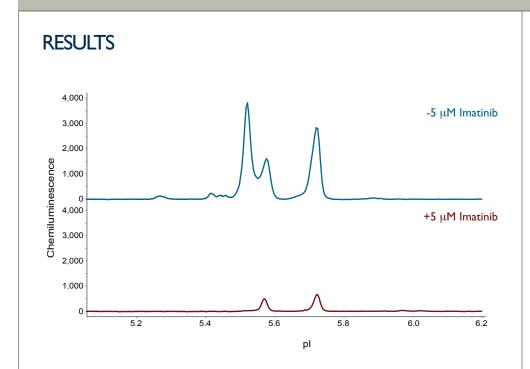
NanoPro[™] Assay: p-STAT5

APPLICATION BRIEF No. 1013

SUMMARY

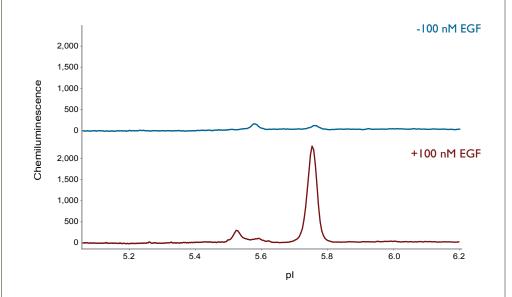
Primary Antibody: Anti-phospho STAT5a (Abcam, cat# ab30648) Detection Antibody: Anti-Rabbit HRP (Cell Biosciences, p/n 040-656)

The Signal Transducer and Activator of Transcription (STAT) proteins regulate many aspects of cell growth, survival and differentiation. The transcription factors in this family are activated by Janus Kinase (JAK). Dysregulation of this pathway is frequently observed in primary tumors and leads to increased angiogenesis, enhanced survival of tumors and immunosuppression. STAT5 is constitutively active in the overexpressing BCR-ABL K562 myelogenous leukaemia cell line. Imatinib (also known as Gleevec®), a BCR-ABL inhibitor, reduces STAT5 phosphorylation in these cells. STAT5 is also part of the Epidermal Growth Factor (EGF) signaling cascade as shown in MCF10A cells.



Imatinib treatment reduces STAT5 phosphorylation in K562 cells

K562 cells were treated -/+ 5 μM imatinib for 24 hours. The Abcam ab30648 anti-phospho STAT5a antibody recognized several peaks in the control cells (blue trace). These peaks were strongly inhibited by imatinib treatment (red trace). Peaks with similar pls were detected in EGF-treated MCF10A cells (see figure below).



EGF treatment increases STAT5 phosphorylation in MCFI0A cells

MCF10A cells were treated -/+ 100 nM EGF for 5 minutes. The Abcam ab30648 anti-phospho STAT5a antibody detected weak phospho STAT5 signals in the untreated MCF10A cells (blue trace). Exposure to EGF strongly induced phospho STAT5 signal (red trace).

NOTE: Detection of the chemiluminescent signal produced is relative. Absolute units may vary depending on cell line, treatment and assay conditions.

PROTOCOL

Cell Preparation

Cell culture: K562 cells (ATCC, cat# CCL-243) were cultured in RPMI 1640 media (Cellgro, cat# 10-041-CV) containing 10% FBS (Irvine

Scientific, cat# 3000-A) and 1x Penicillin/Streptomycin/Glutamine (JRS Scientific, cat# 20020). Cells were split 1:5 every 3 days by

removing an aliquot of cells and transferring them to fresh media. Data shown from cells at passage 5.

Pre-treatment: Cells were starved for 20 hours before stimulation at 37 °C, 5% CO₂ in starvation medium containing RPMI 1640 without serum.

Treatment: 5 µM Imatinib Methanesulfonate Salt (LC laboratories, cat# 1-5508) in starvation medium for 24 hours at 37 °C, 5% CO₂. Lysis buffer: Bicine/CHAPS Lysis Buffer (Cell Biosciences, p/n 040-764) plus 1x DMSO Inhibitor Mix (Cell Biosciences, p/n 040-510) and

Ix Aqueous Inhibitor Mix (Cell Biosciences, p/n 040-482).

Lysis details: Collect cells by centrifugation (1000 x g, 5 minutes). Transfer cells to a 15-mL centrifuge tube, spin (1000 x g, 5 minutes) to pellet

the cells. Aspirate media. Wash cell pellet with 1 mL of ice-cold PBS (Cellgro, cat# 21-031-CV). Transfer cells to a 1.5-mL centrifuge tube, spin (14,000 \times g, 2 minutes). Aspirate wash. Keeping tube on ice, add 400 μ L ice-cold lysis buffer to pellet, pipet up and down to resuspend. Incubate for an additional 30 minutes, rotating. Clarify by centrifugation (14,000 \times g, 15 minutes). Transfer supernatant

to a fresh microfuge tube. Immediately aliquot supernatant (10-30 µL) on ice and snap freeze on dry ice.

Storage: -80 °C

Cell culture: MCF10A cells (ATCC, cat# CRL-10317) were cultured in MEGM (Lonza, cat# CC-3150) containing 10% FBS,

1x Penicillin/Streptomycin/Glutamine, and MEGM SingleQuots. Cells were split 1:5 every 3 days using 0.25% Trypsin (Cellgro,

cat# 25-053-Cl) at 37 °C for 3-5 minutes. Data shown from cells at passage 4.

Pre-treatment: Cells were starved for 20 hours before stimulation at 37 °C, 5% CO₂ in starvation medium containing MEGM.

Treatment: 600 ng/mL EGF in starvation medium for 5 minutes at 37 °C, 5% CO₂.

Lysis buffer: Bicine/CHAPS Lysis Buffer (Cell Biosciences, p/n 040-764) plus Ix DMSO Inhibitor Mix and Ix Aqueous Inhibitor Mix.

Lysis details: Wash cells with 10 mL of ice-cold PBS (Cellgro, cat# 21-031-CV), aspirate well. Add 400 µL ice-cold lysis buffer to 10-cm plate on

ice, swirl around to ensure good coverage, and incubate 10 minutes on ice. Scrape plate, pipet up and down to mix. Transfer lysate to microfuge tube, lyse for an additional 30 minutes on ice. Clarify by centrifugation (14,000 \times g, 15 minutes) in a cooled centrifuge. Transfer supernatant to a fresh microfuge tube. Immediately aliquot supernatant (10–30 μ L) on ice and snap freeze on dry ice.

Storage: $-80 \,^{\circ}\text{C}$

Assay Reagents

NOTE: For specifics on sample preparation, please consult the addendum to this document.

Protein concentration: 0.1 mg/mL final in capillary by BCA assay

Sample diluent: Sample Diluent (Cell Biosciences, p/n 040-649) plus 1x DMSO Inhibitor Mix

Ampholyte premix: 5-8 (Cell Biosciences Premix G1, p/n 040-643 or Premix G2, p/n 040-972)

pl standards: pl Standard Ladder 3 (Cell Biosciences, p/n 040-646)

Wash: Wash Buffer (Cell Biosciences, p/n 040-654)

Primary antibody: Anti-phospho STAT5a (Abcam, cat # ab30648), 1:100 in Antibody Diluent (Cell Biosciences, p/n 040-309)

Detection antibody: Anti-Rabbit HRP (Cell Biosciences, p/n 040-656), 1:100 in Antibody Diluent

Anolyte: Phosphoric Acid, 10 mM (Cell Biosciences, p/n 040-650)

Catholyte: Sodium Hydroxide, 100 mM (Cell Biosciences, p/n 040-651)

Luminol/Peroxide: Mixed 1:1 (Cell Biosciences, p/n 040-652 and p/n 040-653)

NOTE: NanoPro XDR Assays are now available with optional reagents to provide optimized assay sensitivity, extended dynamic range and both low and high end signal enrichment. Additionally, Premix G2 provides enhanced protein resolution. For more information, visit cellbiosciences.com, contact your Cell Biosciences' Field Applications Specialist or call Technical Support at (888) 607-9692.

Assay Conditions

System: NanoPro 1000

Sample loading time: 10 seconds (Premix G1), 25 seconds (Premix G2)

Focus conditions: $15000 \mu W$, 40 minutes (Premix G1) or

21000 µW, 40 minutes (Premix G2)

Immobilization: 80 seconds

Wash I: 2×150 seconds (default)

Primary antibody incubation: 120 minutes

Wash 2: 2×150 seconds (default)

Detection antibody incubation: 60 minutes

Wash 3: 2×150 seconds (default) Chemiluminescence exposure: 60, 120, 240, and 480 seconds

Our favorite antibody

Anti-phospho STAT5a (Abcam, cat# ab30648)

Other antibody suggestions

Anti-STAT5 (Abcam, cat# 68465)

Anti-STAT5 (Santa Cruz Biotechnology, cat# sc-28685)

Anti-STAT5 (Genscript, cat# A00253)

Anti-phospho STAT5a/b (Cell Signaling Technology, cat# 9351)

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SAMPLE PREPARATION ADDENDUM

Sample preparation procedures are different depending on the premix used. The following updated instructions provide guidance on sample preparation for both Premix G1 and Premix G2.

Step	Premix G1 Procedure	Premix G2 Procedure
Step I	Dilute lysate with sample diluents to 0.1 mg/mL.	Dilute lysate with sample diluents to 0.2 mg/mL.
Step 2	In a separate tube, mix Premix GI and pI standards.	In a separate tube, mix Premix G2 and pl standards.
Step 3	Mix equal parts of diluted lysate prepared in Step I with the Premix GI + pl Standards prepared in Step 2 (I:I ratio) to create final protein concentration of 0.05 mg/mL.	Mix I part diluted lysate prepared in Step I with 3 parts Premix G2 + pl Standards prepared in Step 2 (1:3 ratio) to create final protein concentration of 0.05 mg/mL.

NOTES: When working with Premix G2, thorough mixing and vortexing during sample preparation is required. Additional sample volume may be required, 12-20 uL per sample well is recommended. Centrifugation of the sample plate (3000 x g, 10 minutes) is required.

For further assistance, please contact your Cell Biosciences' Field Applications Specialist or Technical Support at (888) 607-9692.

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