# Co-detection of PD1/PD-L1 interaction, immune cell markers and cytokine mRNAs using RNAscope<sup>TM</sup> Multiomic LS assay is important for understanding anti-PD-L1 treatment response in muscle invasive bladder cancer patients

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### INTRODUCTION

- Network of protein-protein interactions (PPIs) and the underlying stoichiometry both within a tissue and with the immune repertoire is adversely impacted in many diseases, including cancer. While screening technologies to predict and map the protein interactome are advancing our understanding of intercellular/intracellular PPIs, no orthogonal technologies exist that can validate inferred PPIs and add context using known mRNA/protein markers.
- Technology that uses modifications to RNAscope<sup>™</sup> a validation technology known for its high sensitivity and specificity – has been developed to visualize PPIs, proteins, and mRNA *in situ* on the same tissue section.
- The assay was deployed on samples from patients with localized, muscle-invasive bladder cancer who received neoadjuvant anti-PD-L1 checkpoint inhibitor therapy prior to planned surgical cystectomy on a clinical trial (NCT02451423). To characterize tumor immune microenvironment of these patients, both responders and non-responders to the treatment, PD-1/PD-L1 interactions were assessed using a panel of targets to include cell phenotyping protein markers for both immune and tumor cells and mRNA markers for effector molecules. We visualized PD-1/PD-L1 interactions, and changes in cellular interactions between cytotoxic T cells or regulatory T cells and CK+ tumor cells before and after immunotherapy treatment.

### METHOD



Figure 1. RNAscope Multiomic LS Assay workflow. FFPE sections are first pretreated, followed by application of conjugated antibodies and target RNA specific probes. RNA transcripts and protein-protein proximity/interaction appear as punctate dots.

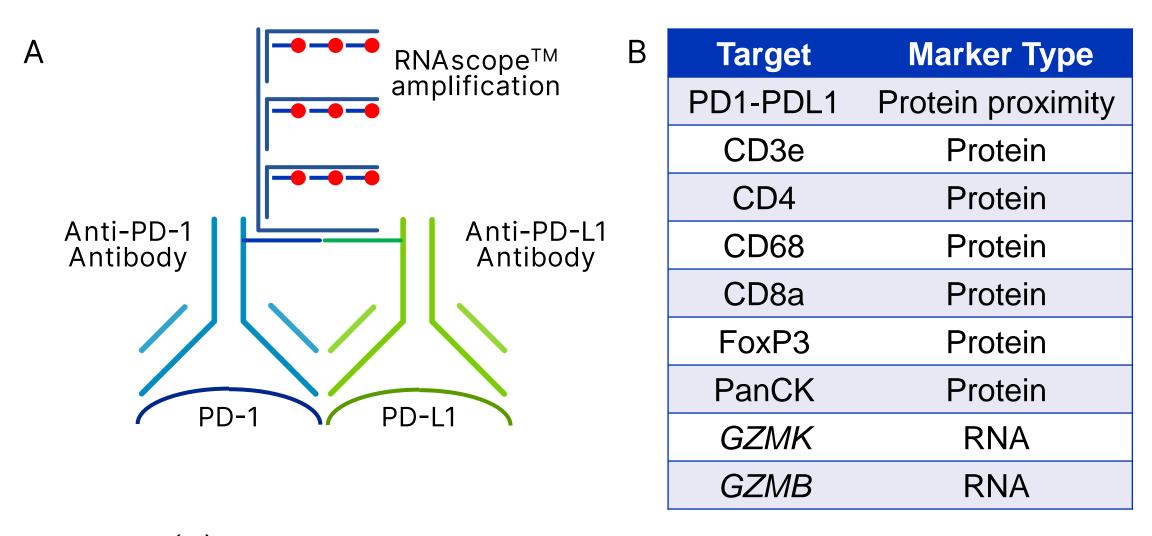
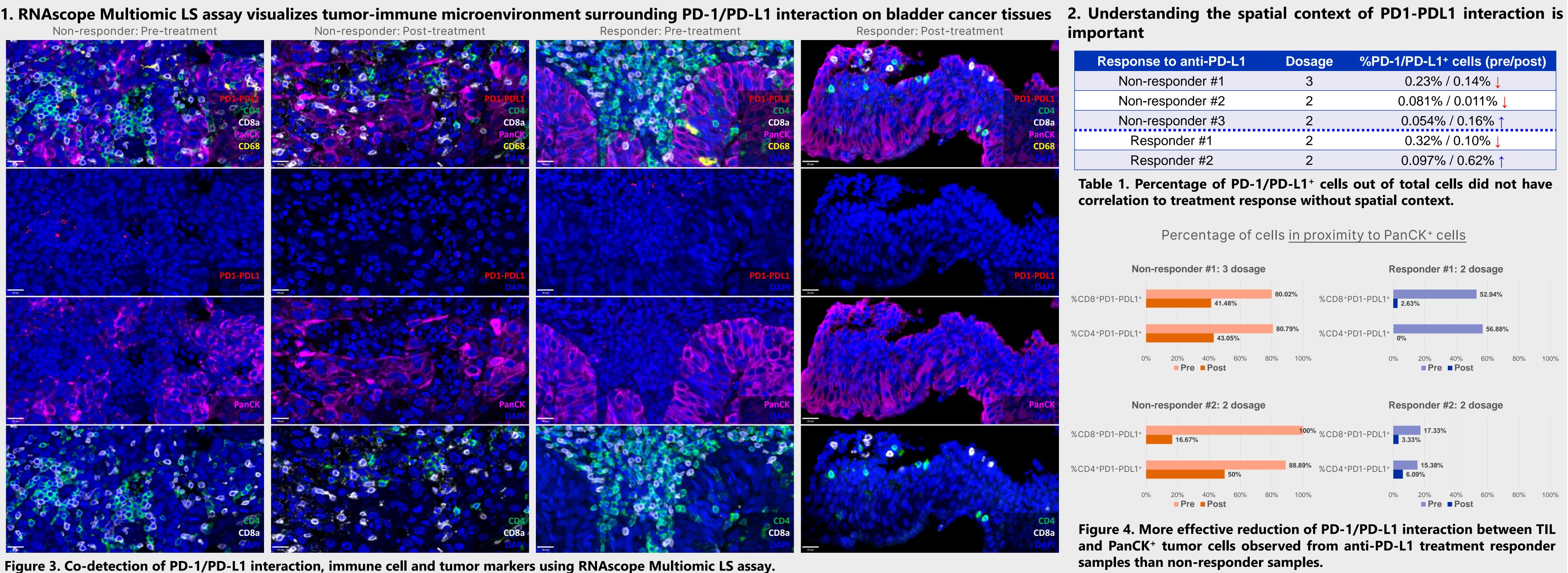


Figure 2. (A) Schematic for detection of PD-1/PD-L1 interaction using oligonucleotide-conjugated primary antibodies targeting PD-1 and PD-L1. (B) List of target markers detected on the patient samples.





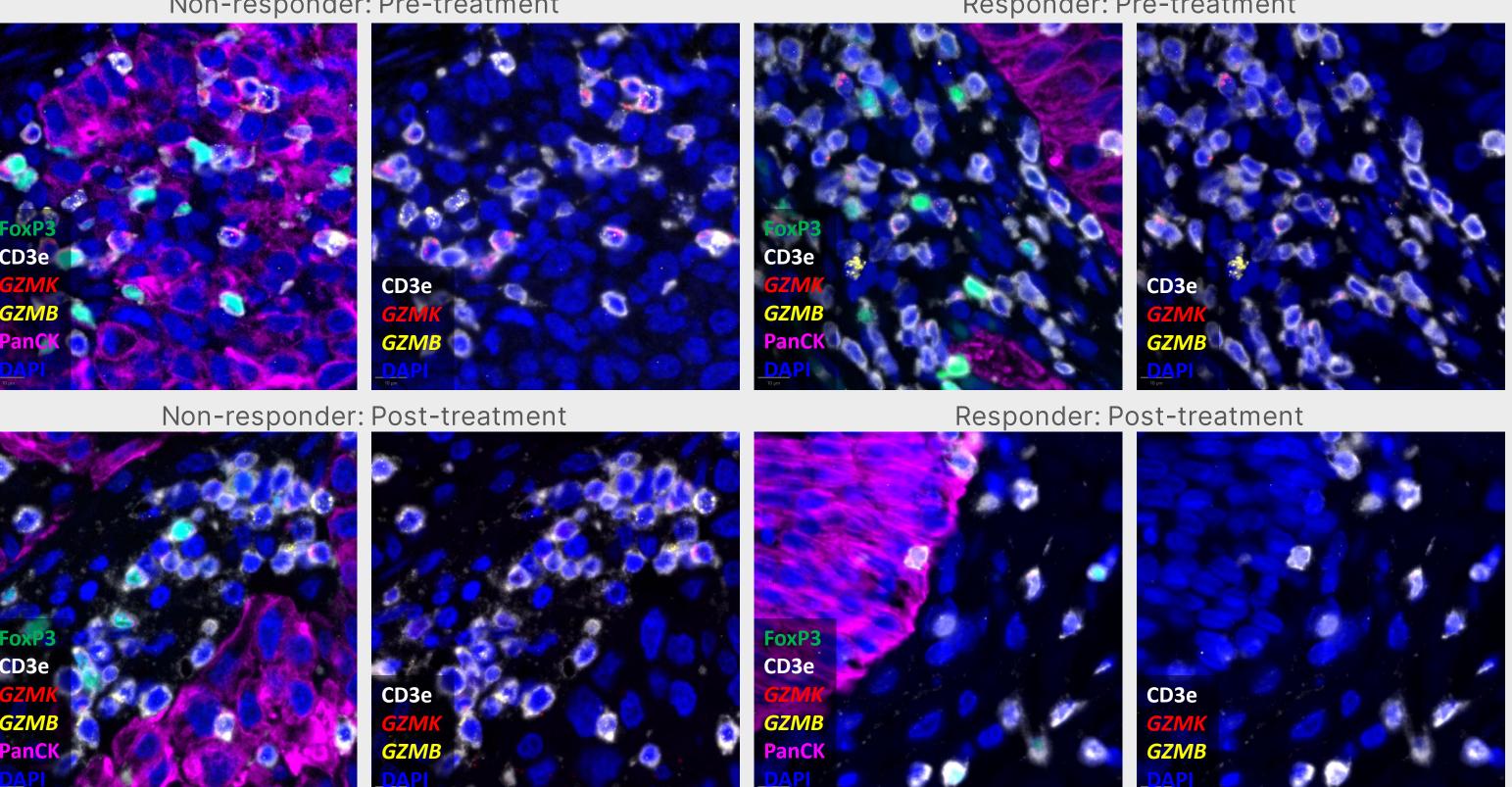
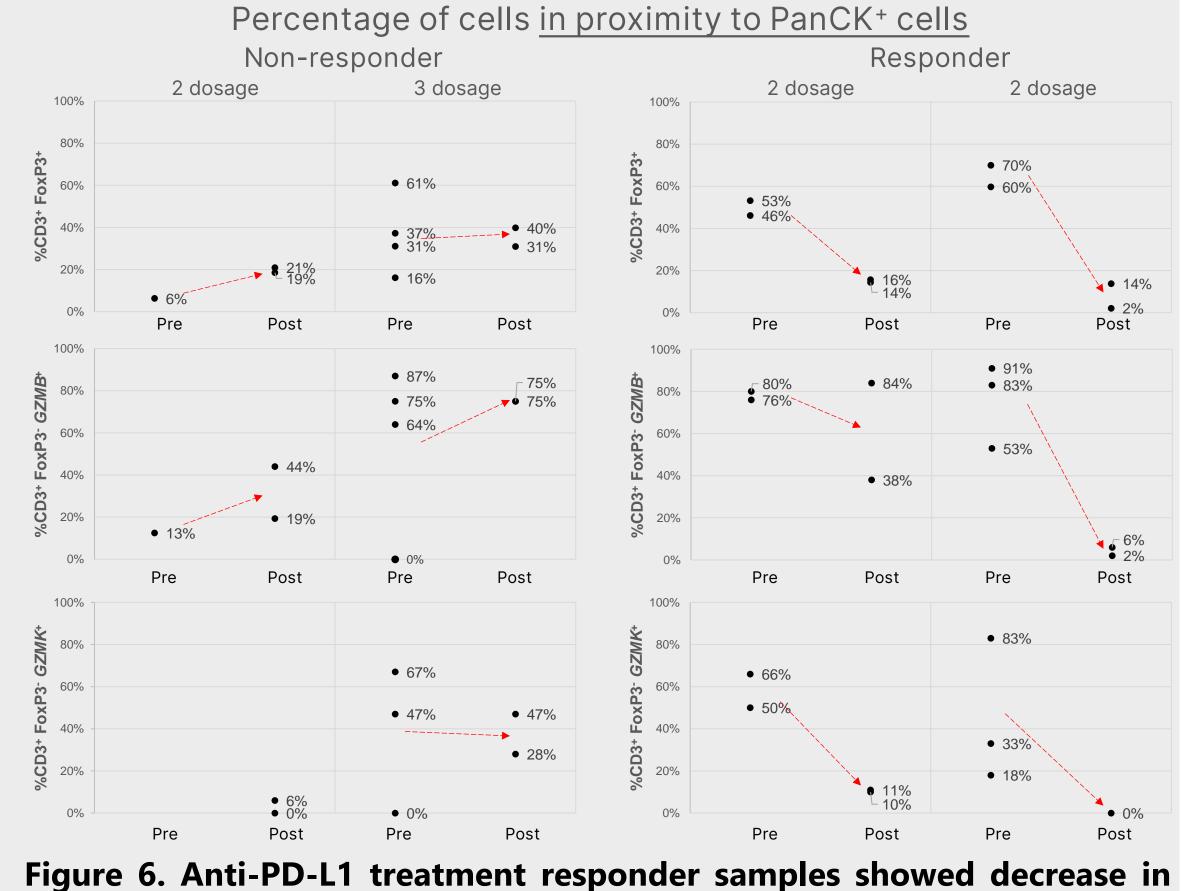


Figure 5. Protein (CD3e, FoxP3, PanCK) and RNA (GZMB, GZMK) markers for regulatory T cells and cytotoxic T cells were visualized using RNAscope Multiomic LS assay.



percentage of tumor-infiltrating regulatory T cells and cytotoxic T cells expressing granzyme B (GZMB) or granzyme K (GZMK).

## important

Response to anti-PD-L1	Dosage	%PD-1/PD-L1+ cells (pre/post)
Non-responder #1	3	0.23% / 0.14% ↓
Non-responder #2	2	0.081% / 0.011% 👃
Non-responder #3	2	0.054% / 0.16% ↑
Responder #1	2	0.32% / 0.10% 👃
Responder #2	2	0.097% / 0.62% ↑

Table 1. Percentage of PD-1/PD-L1<sup>+</sup> cells out of total cells did not have correlation to treatment response without spatial context.

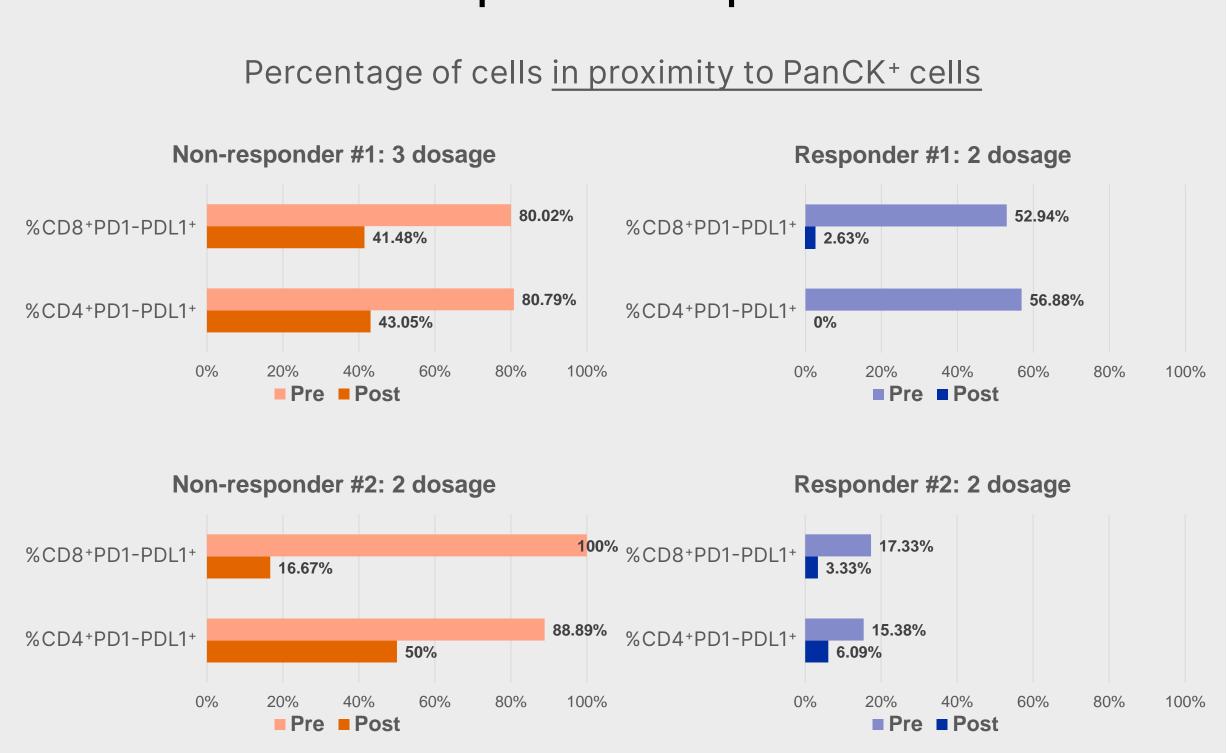


Figure 4. More effective reduction of PD-1/PD-L1 interaction between TIL and PanCK<sup>+</sup> tumor cells observed from anti-PD-L1 treatment responder samples than non-responder samples.

### **SUMMARY**

- This fully-automated assay on BOND RX enables simultaneous detection of 1 protein-protein proximity/interaction and up to 5 targets of RNA and individual protein on a single FFPE tissue section.
- PD-1/PD-L1 interaction is visualized at high resolution with oligonucleotide-conjugated primary antibodies for PD1 and PD-L1 using RNAscope<sup>TM</sup> Multiomic LS assay on human bladder cancer patient tissues.
- PD-1/PD-L1 interaction evaluated along with spatial multiomics is essential to derive context around anti-PD-L1 treatment response.

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